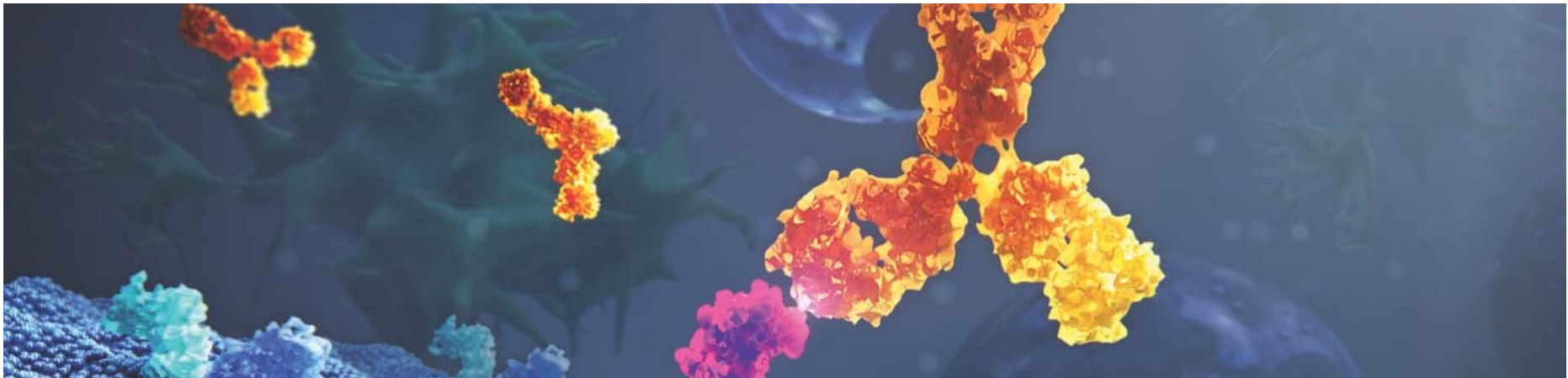


# Attack of the Clones: Understanding the kinetics of resistance to cancer treatment

Dr James Yates, Oncology R&D, AstraZeneca



## Introduction

**Treatment failure is a barrier to cure or at least long term disease control in cancer patients.**

**The evolution of drug resistant cancer cells is a predominant cause.**

**How can we understand this quantitatively?**



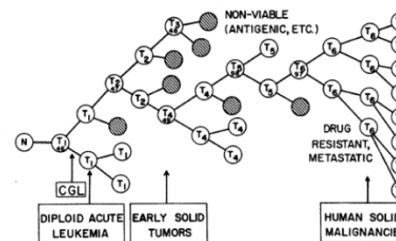
**What do we know about drug resistance in the clinic?**

# By the end of the 1970s there was evidence of clonal selection

## The Clonal Evolution of Tumor Cell Populations

Acquired genetic lability permits stepwise selection of variant sublines and underlies tumor progression.

1 OCTOBER 1976 Science Peter C. Nowell



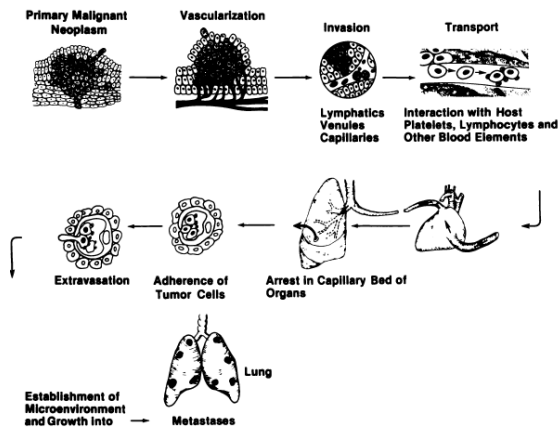
The acquired genetic instability and associated selection process, most readily recognized cytogenetically, results in advanced human malignancies being highly individual karyotypically and biologically. Hence, each patient's cancer may require individual specific therapy, and even this may be thwarted by emergence of a genetically variant subline resistant to the treatment. More research should be directed toward understanding and controlling the evolutionary process in tumors before it reaches the late stage usually seen in clinical cancer.

Cancer Research 38:2651-2660. 1978

### Tumor Heterogeneity and the Biology of Cancer Invasion and Metastasis<sup>1</sup>

Isaiah J. Fidler

Cancer Biology Program, National Cancer Institute Frederick Cancer Research Center, Frederick, Maryland 21701



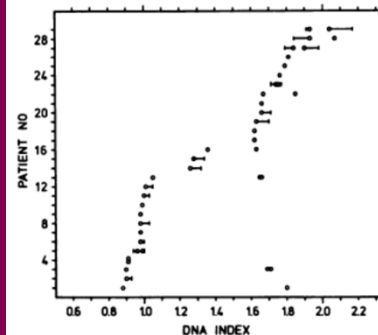
4

Cancer Research 40:4295-4300. 1980

### Clonal Heterogeneity of Small-Cell Anaplastic Carcinoma of the Lung Demonstrated by Flow-Cytometric DNA Analysis<sup>1</sup>

Lars L. Vindeløv,<sup>2</sup> Heine H. Hansen, Ib J. Christensen, Mogens Spang-Thomsen, Fred R. Hirsch, Mogens Hansen, and Nis I. Nissen

The Finsen Institute [L. L. V., H. H. H., I. J. C., F. R. H., M. H., N. I. N.] and the University Institute of Pathological Anatomy [M. S.], Copenhagen, Denmark



of the clones. To reach  $10^9$  to  $10^{12}$  cells (1 g to 1 kg of tumor), 30 to 40 tumor volume doublings have occurred. If the 2 clones have identical TD's, they must both have been present within the first 4 TD's of the tumor. Otherwise, one of them would constitute less than 6.3% (one-sixteenth) of the total mass and would therefore escape detection. Similar doubling times there-



# TracerX trial: C21<sup>st</sup> Tracking clonal evolution in NSCLC

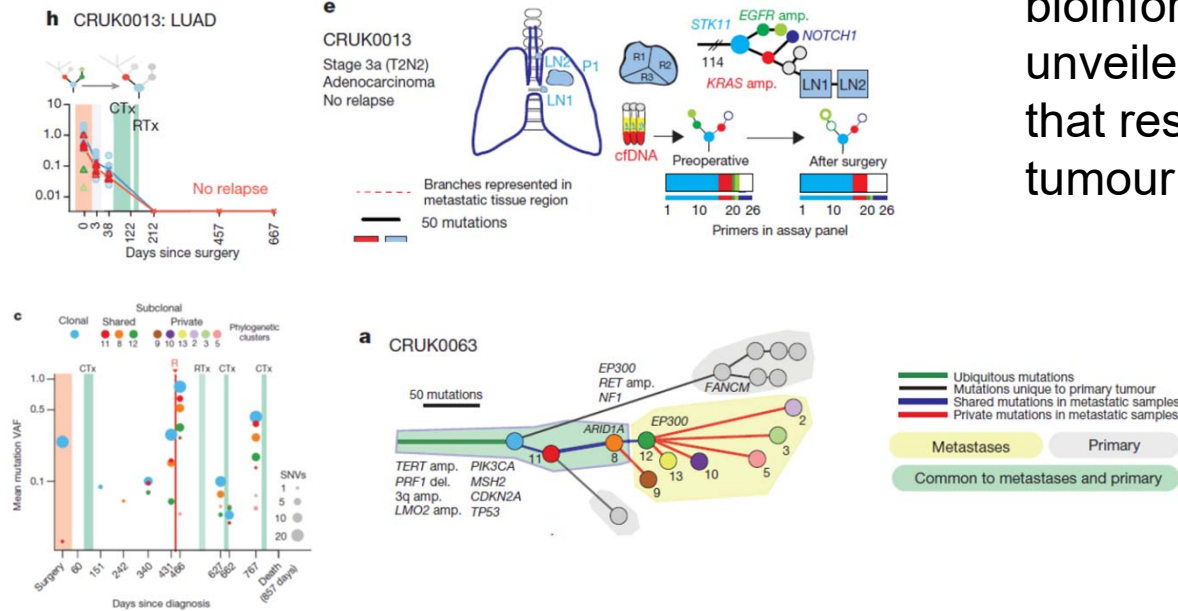
ARTICLE Nature 545: 446. 2017

doi:10.1038/nature22364

## Phylogenetic ctDNA analysis depicts early-stage lung cancer evolution

A list of authors and their affiliations appears in the online version of the paper.

Longitudinal and multisite biopsies coupled with bioinformatic techniques unveiled the clonal evolution that results in heterogeneous tumour populations

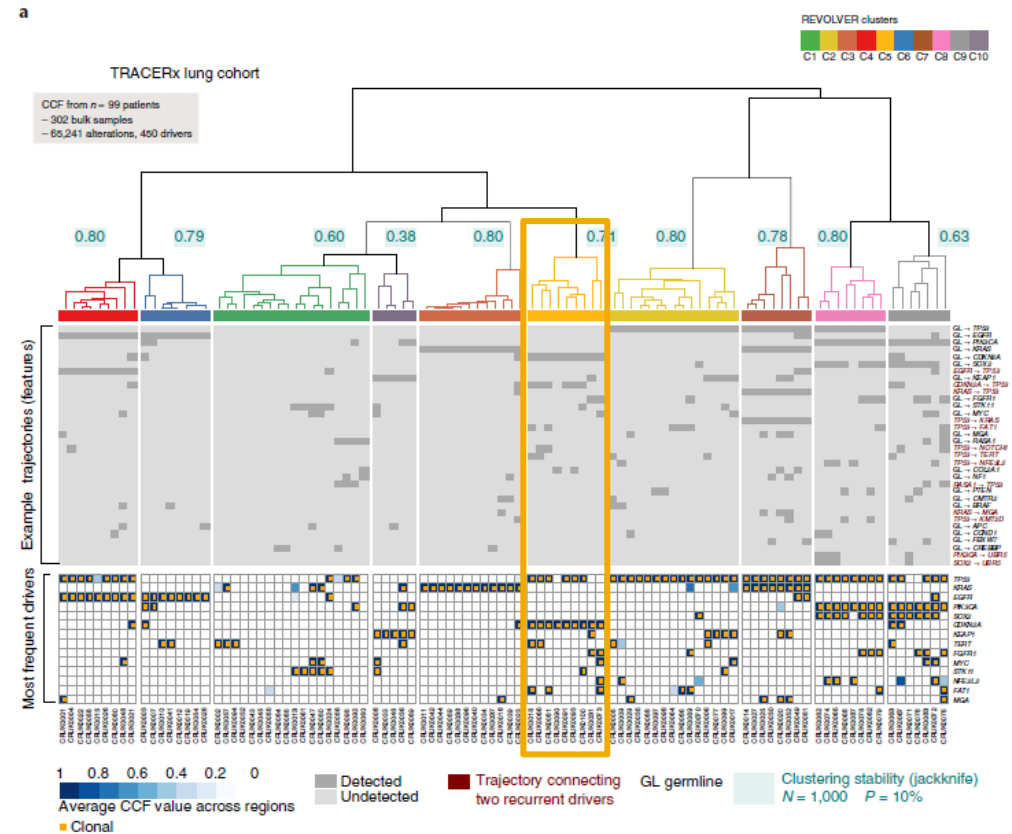
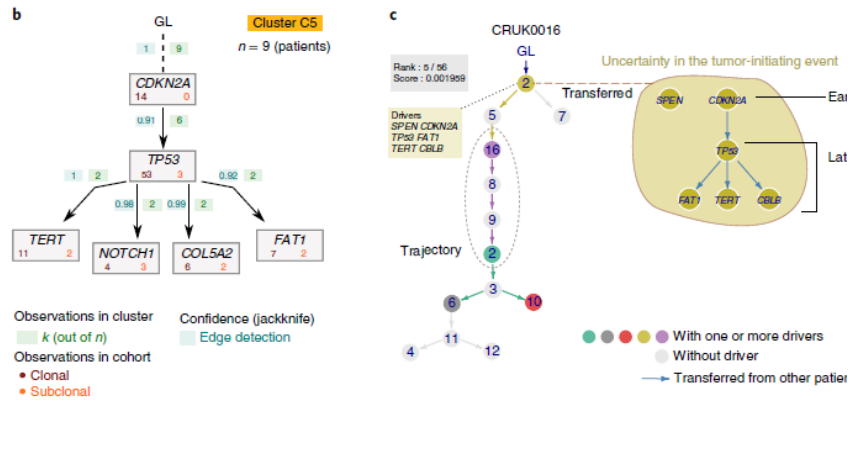


# TracerX: patient clusters of evolutionary history

nature methods Vol 15. 2018 ARTICLES  
<https://doi.org/10.1038/s41592-018-0108-x>

## Detecting repeated cancer evolution from multi-region tumor sequencing data

Giulio Caravagna<sup>1,2\*</sup>, Ylenia Giarratano<sup>2,3</sup>, Daniele Ramazzotti<sup>4</sup>, Ian Tomlinson<sup>5</sup>, Trevor A. Graham<sup>6</sup>, Guido Sanguinetti<sup>2\*</sup> and Andrea Sottoriva<sup>1\*</sup>

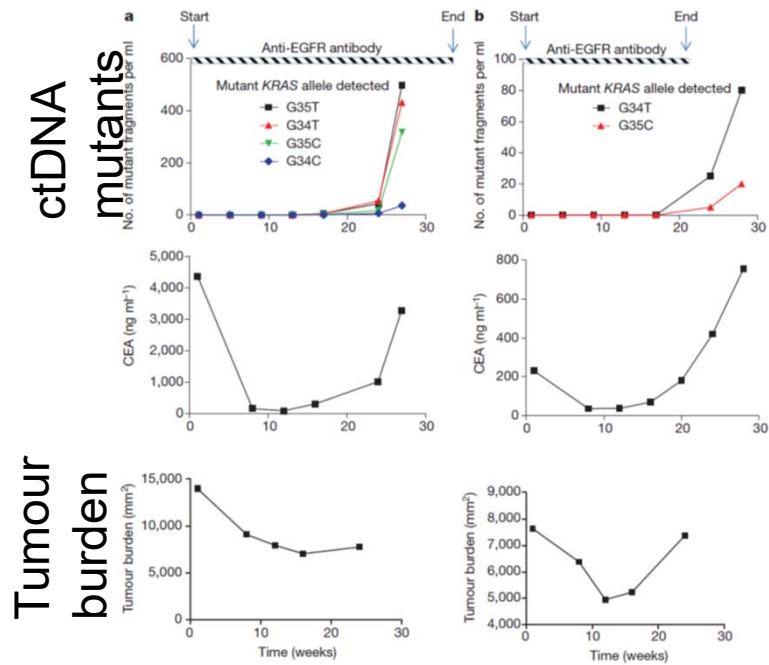


Suggests there are subgroups of patients with common treatment options  
 Anticipating this before treatment is key



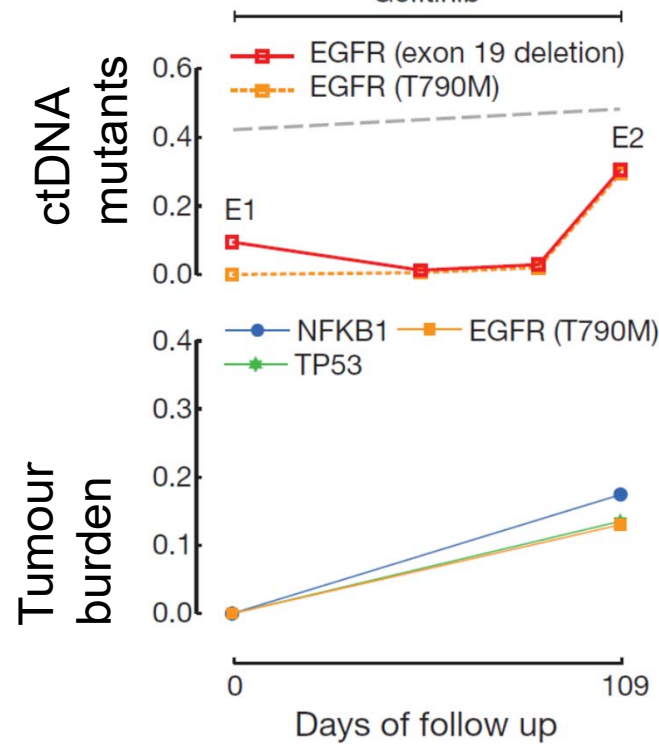
# Resistance kinetics observed in Circulating tumour DNA

## Colorectal cancer



[doi:10.1038/nature11219](https://doi.org/10.1038/nature11219)

## LUAD



[doi:10.1038/nature12065](https://doi.org/10.1038/nature12065)



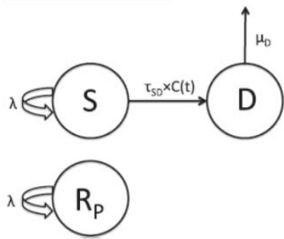
# CPT:PSP review on resistance models. Models reflecting tumour heterogeneity

Citation: CPT Pharmacometrics Syst. Pharmacol. (2019) XX, 1–18; doi:10.1002/psp4.12450

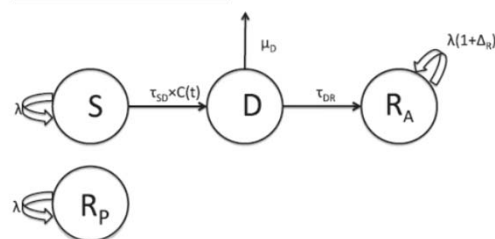
doi: 10.1111/jcp.12259

Analysis of temozolomide resistance in low-grade gliomas using a mechanistic mathematical model

Primary resistance only



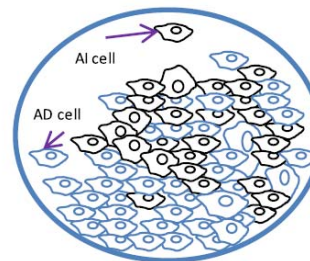
Primary and acquired resistance



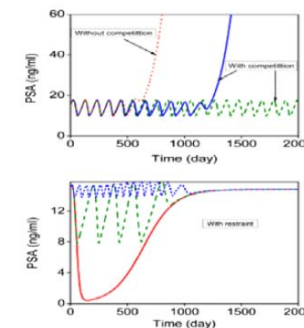
A nonlinear competitive model of the prostate tumor growth under intermittent androgen suppression

Jing Yang, Tong-Jun Zhao\*, Chang-Qing Yuan, Jing-Hui Xie, Fang-Fang Hao

Journal of Theoretical Biology 404 (2016) 66–72



Cancer cells compete with each other in a limited environment



Common motif is delineation between drug Sensitive and Resistant Cells  
 Various assumptions on whether resistance pre-exists treatment  
 Incorporation of both mechanisms + reversal of resistant phenotype to sensitive raises question of parameter identifiability

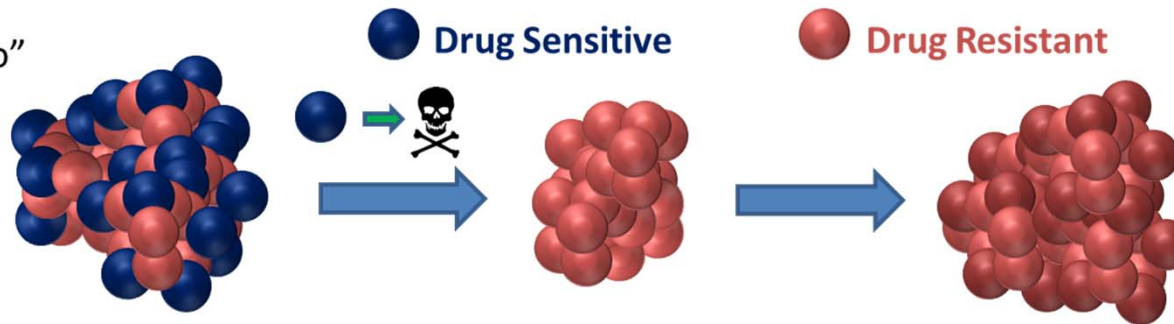


## Application: early or late onset resistance for EGFRi?

- Tested 2 models that describe baseline “de novo” resistant fraction vs on treatment mutation.

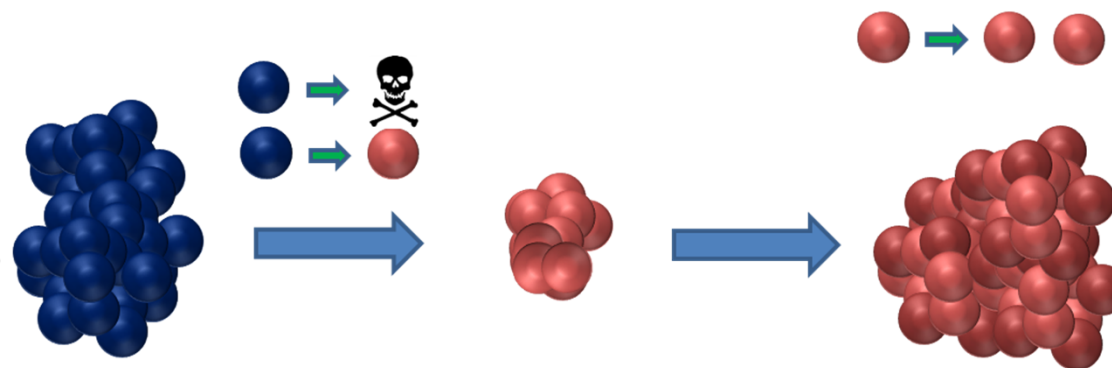
Model 1: “De-Novo”

Assumption:  
2 populations  
Infer:  
Proportion  
growing/decaying prior  
to treatment



Model 2: “Acquired”

Assumption:  
1 population that has  
ability to adapt



## Application: early or late onset resistance for EGFRi?

- Tested 2 models that describe baseline “de novo” resistant fraction vs on treatment mutation.

De Novo

$$Y(t) = Y(0)((1 - \phi)e^{-dt} + \phi e^{gt})$$

Initial size  $Y(0)$

Fraction resistant  $\phi$

Kill rate  $d$

Acquired

$$Y(t) = Y(0)e^{-(d+c)t} + \frac{cY(0)}{d+c+g}(e^{gt} - e^{-(d+c)t}),$$

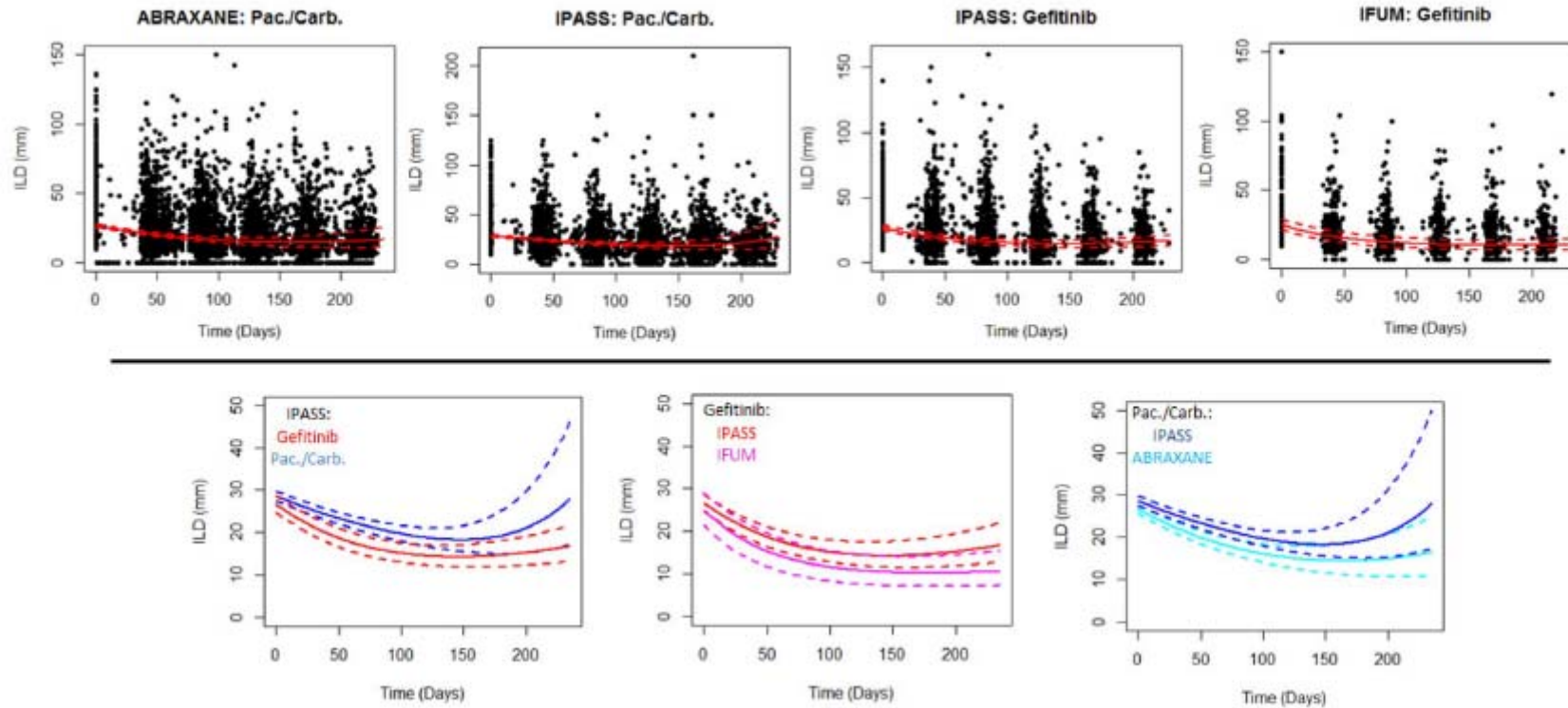
Outgrowth rate  $g$

“Mutation rate”  $c$

Can we distinguish between these two models?



## Early onset “de novo” resistance kinetics observed

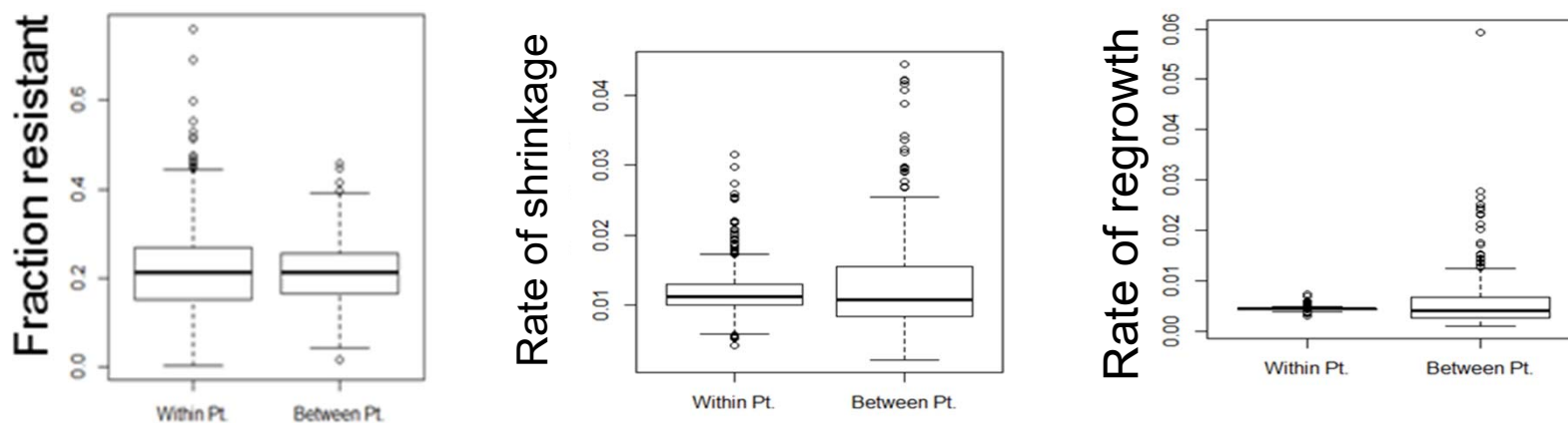


We see early “de novo” resistance and provides evidence that pre-clinical clone mixing approach is valid.

## Early onset “de novo” resistance kinetics observed

Gefitinib data described by “de novo” model

Individual lesion size modelled: within patient variability < between patient except for fraction resistant

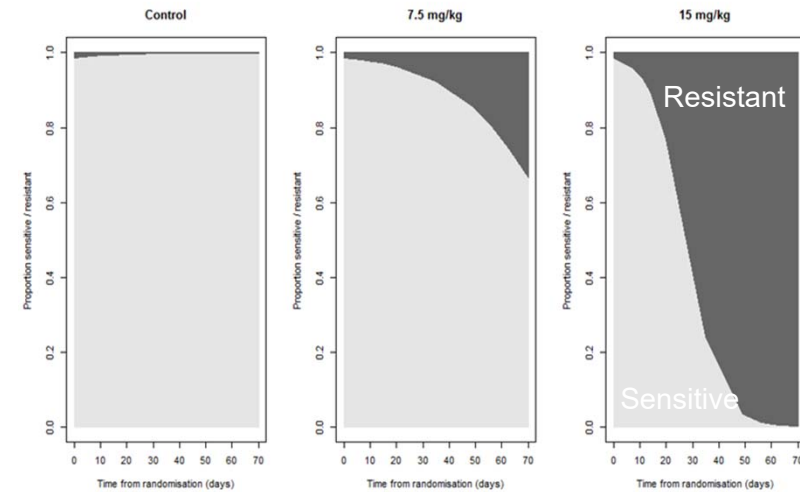
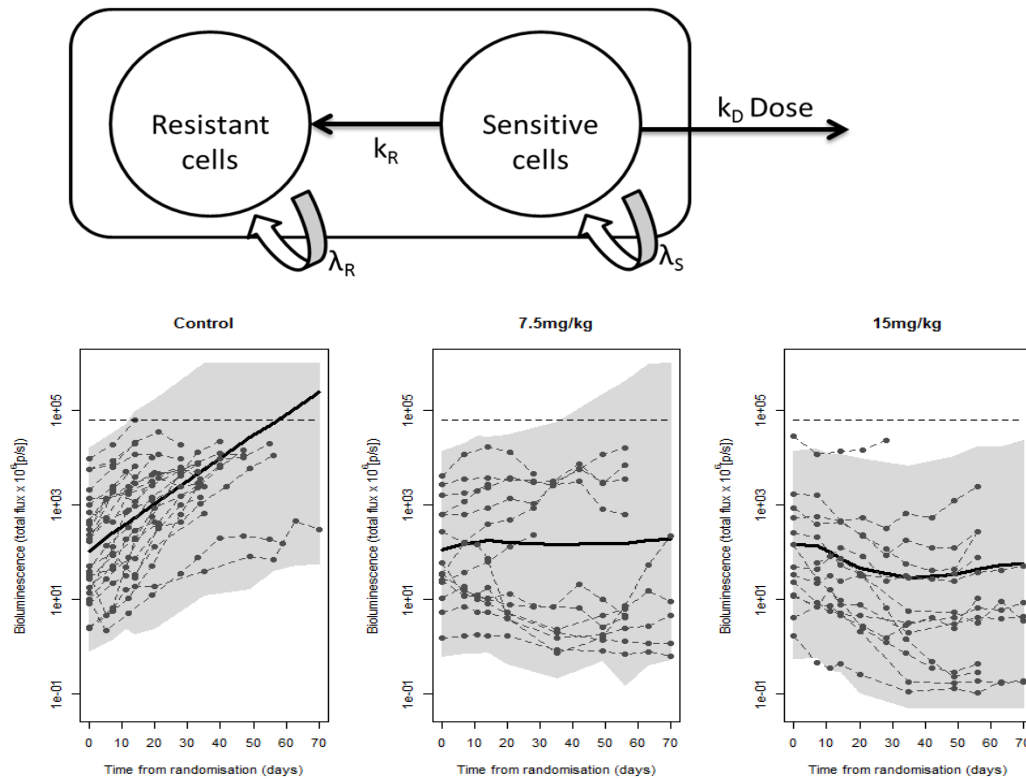


We see early “de novo” resistance and provides evidence that pre-clinical clone mixing approach is valid for investigating resistance.



# Resistance kinetics to EGFRi in mouse models

Pharmacodynamic modelling of resistance to epidermal growth factor receptor inhibition in brain metastasis mouse models



- Again: Baseline resistance better explanation
- Resistant cells have reduced proliferation rate
- Lower dose gives greater disease control



# The impact of doubling time on resistance fraction

Bull Math Biol (2012) 74:1379–1395  
DOI 10.1007/s11538-012-9717-1

ORIGINAL ARTICLE

On the Probability of Random Genetic Mutations  
for Various Types of Tumor Growth

Cristian Tomasetti

## Model

$$S'(t) = \left[ L(1-u)(1-a-b) - \left( D + bL + \frac{uaL}{2} \right) \right] \left( 1 - \frac{S(t)}{K} \right) S(t),$$

$$R'(t) = \left[ \left[ L(1-a-b) - (D + bL) \right] R(t) + uL \left( 1 - \frac{a}{2} - b \right) S(t) \right] \times \left( 1 - \frac{S(t)}{K} \right). \quad (7)$$

$$P_R = 1 - \exp \left( -uM \left( \frac{1 - \frac{a}{2} - b}{1 - a - b} \right) \frac{1}{C} \ln \left( \frac{1}{1 - C} \right) \right), \quad C = \frac{D + bL}{L(1 - a - b)}$$

$$E(T | \text{resistance}) \approx \frac{M}{P_R} \left( \frac{u(1 - \frac{a}{2} - b)}{(1 - a - 2b) - D/L} \right) \ln(M).$$

Probability of resistance at diagnosis  
independent of law of growth kinetics

It is dependent on underlying balance of  
proliferation (L) vs death (D)

There is literature evidence that human  
tumours have significant cell death  
(D/L~0.9) whereas in animal models it is  
lower (D/L~0.5)

Consequences for resistance  
development in animal models vs clinic?



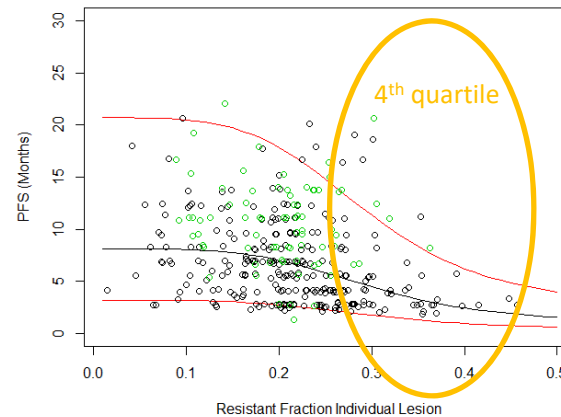
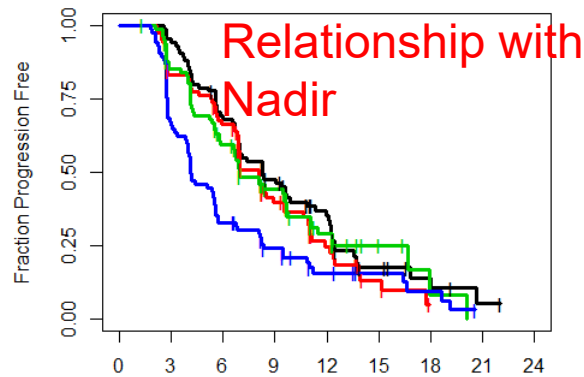
## The linkage between resistance and PFS

# Returning to EGFRi: Unclear relationship between tumour shrinkage and PFS

- Relationship between growing fraction and PFS exists – appears non-linear (KM-plots)
- Modelled relationship: acknowledge interval censored, data-descriptive approach, final model log-normal, used Emax model for functional relationship for covariate
  - Plot shows median, 2.5<sup>th</sup> and 97.5<sup>th</sup> PFS times as a function of growing fraction, black circles progressed patients, green circles patients right-censored

PFS is relative to nadir so no surprise not a strong relationship

Can the poor PFS patients be spotted after first 2 visits?



	Number at risk								
	1st Quartile:	2nd Quartile:	3rd Quartile:	4th Quartile:	0	3	6	9	12
Low RF	85	80	58	38	21	8	3	1	0
	84	69	54	26	12	4	0	0	0
	84	69	46	29	14	5	1	0	0
High RF	85	56	28	17	8	5	3	0	0



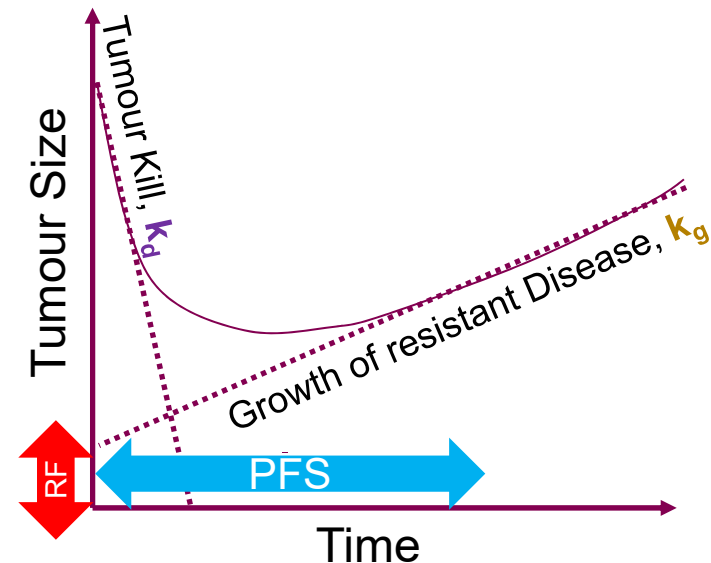


## A simple model explanation of why initial shrinkage and PFS are disconnected

$$Y(t) = Y(0)(RF \cdot e^{k_g t} + (1 - RF) \cdot e^{-k_d t})$$

Response kinetics defined by:

1. Baseline
2. Resistant fraction
3. Rate of shrinkage of drug sensitive fraction
4. Rate of regrowth of drug resistant fraction



This simple model has appeared several times in the literature.

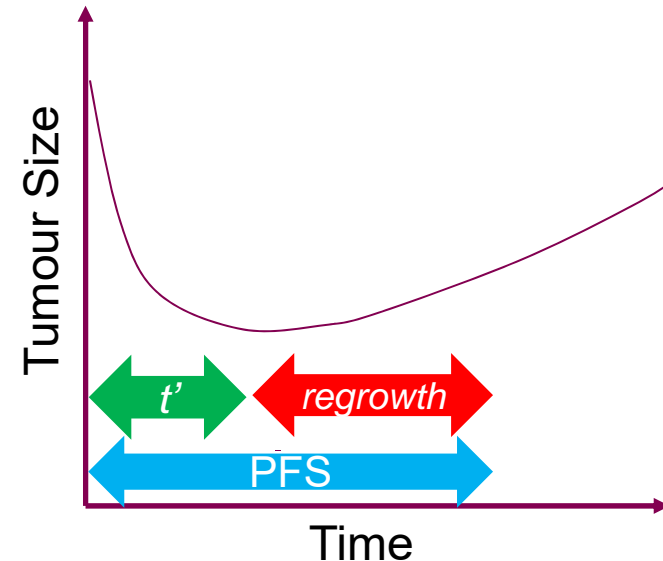


## PFS is time to nadir ( $t'$ ) + time to 20% increase

$$Y(t) = Y(0)(RF \cdot e^{k_g t} + (1 - RF) \cdot e^{-k_d t})$$

Response kinetics defined by:

1. Baseline
2. Resistant fraction
3. Rate of shrinkage of drug sensitive fraction
4. Rate of regrowth of drug resistant fraction



In the next few slides we will look at the relationship between these parameters



## PFS is not always sensitive to resistant fraction

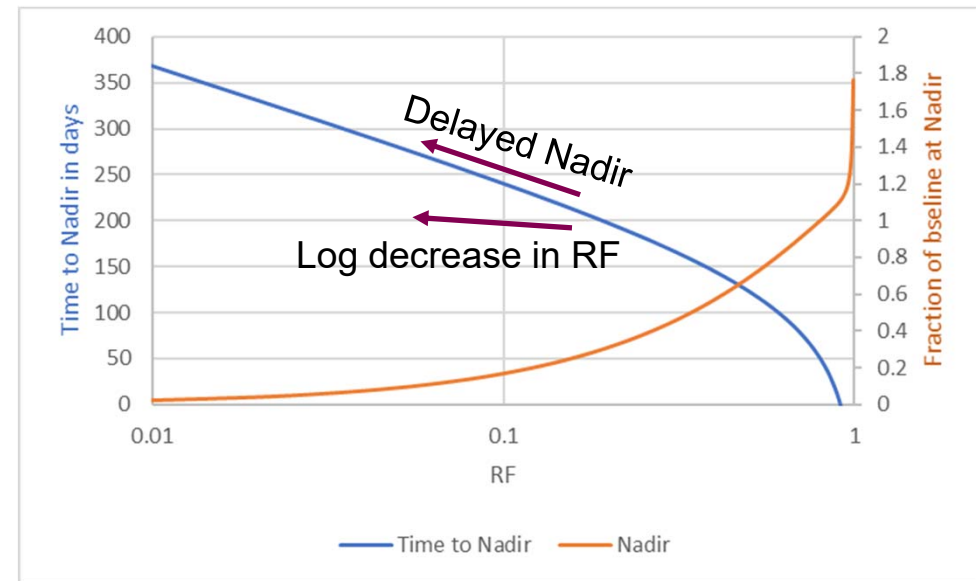
$$t' = \frac{1}{k_g + k_d} \ln \left[ \frac{k_d(1 - RF)}{k_g \cdot RF} \right]$$

We can see RF needs to be reduced by order of magnitude to increase  $t'$

$$RF = \frac{1}{1 + \frac{k_g}{k_d} e^{(k_g + k_d) \cdot t'}}$$

At  $t'$  the tumour will be this size

$$Y(0)(1 - RF)e^{-k_d t'}$$



## Time to progression post nadir is defined by $k_g$

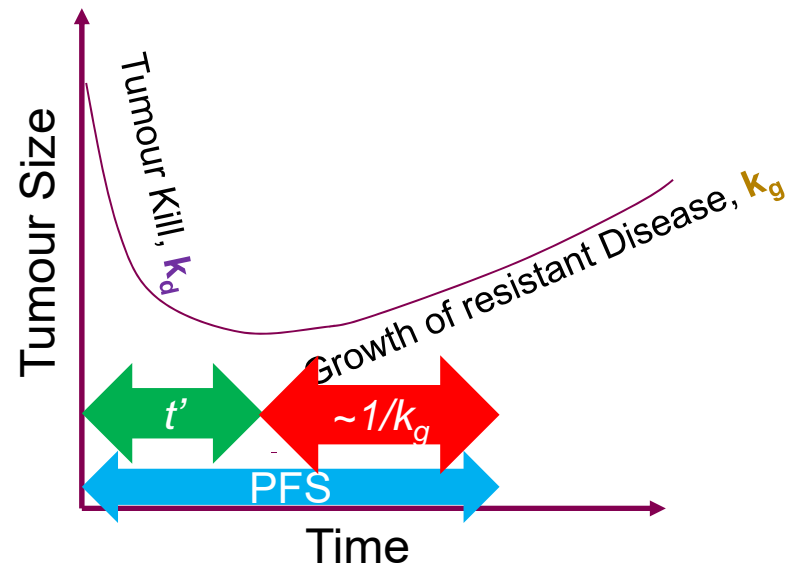
By nadir the tumour has been “purified” and now the resistant fraction is

$$RF' = \frac{k_d}{k_g + k_d}$$

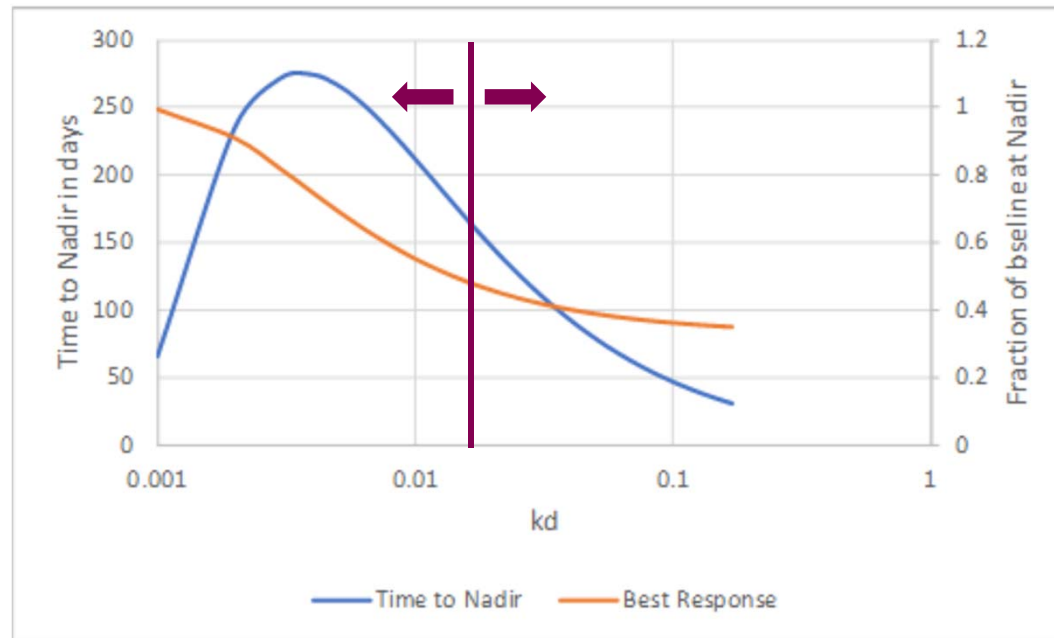
After this point the tumour will grow as:

$$Y(t') = Y(0)(1 - RF)e^{-k_d t'} \left[ \frac{k_d}{k_g} e^{k_g(t-t')} + e^{-k_d(t-t')} \right]$$

Progression at 20% increase from nadir  
Progression is a function of  $1/k_g$

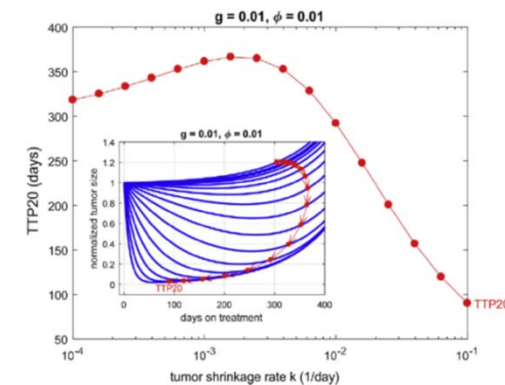


# Time to nadir and rate of tumour kill: Biphasic relationship



Original Research

Directional inconsistency between Response Evaluation Criteria in Solid Tumors (RECIST) time to progression and response speed and depth



Reducing tumour kill rate could increase time to nadir without impacting best response significantly.

Observed via simulation by Millennium Pharmaceuticals Researchers



## Linking Initial Response to OS

FDA NSCLC model with different outgrowth (resistance) rates

$$TS_i(t) = BASE_i \cdot e^{-SR_i \cdot t} + PR_i \cdot t,$$

Treatment	M_BASE (cm)	M_SR (1/week)	M_PR (cm/week)
PCB	9.1 (0.33)	0.06 (0.004)	0.13 (0.02)
PC	8 (0.3)	0.038 (0.01)	0.14 (0.04)
DC	8.7 (0.31)	0.052 (0.01)	0.16 (0.02)
DCb	9.2 (0.38)	0.047 (0.005)	0.16 (0.02)
VC	8.5 (0.28)	0.063 (0.01)	0.17 (0.02)
DT	8.5 (0.82)	0.033 (0.01)	0.13 (0.02)
PT	7.4 (0.47)	0.023 (0.01)	0.25 (0.05)
PB <sup>a</sup> Placebo	8.6 (0.44)	0.0047 slow (0.001) 0.13 fast (0.004)	0.20 (0.02)
ET <sup>a</sup>	8.4 (0.32)	0.0045 slow (0.001) 0.11 fast (0.05)	0.058 (0.02)

- Tumour reduction at 8 weeks shown to be predictive of OS
- Prentice criteria: Many of these studies do not show OS effect differences are fully account for by tumour response. i.e. show treatment not a covariate
- Why do these treatments show different progression growth rates?

DC, docetaxel and cisplatin; DCb, docetaxel and carboplatin; DT, docetaxel; ET, erlotinib; PB, placebo; PC, paclitaxel and carboplatin; PCB, paclitaxel, carboplatin, and bevacizumab; PT, pemetrexed; VC, vinorelbine and cisplatin.



## Key points:

PFS on target lesions as defined by RECIST 1.1 is time to nadir+time to 20% regrowth

PFS in a population will be increased by targeting more mutations because of an increased % objective response rate: Responders have PFS

PFS and OS in a patient will be increased by leaving slower growing disease: post nadir time  $\sim 1/k_g$



**Resistance in nonclinical cancer models**  
**Scope for understanding the clinical issues?**

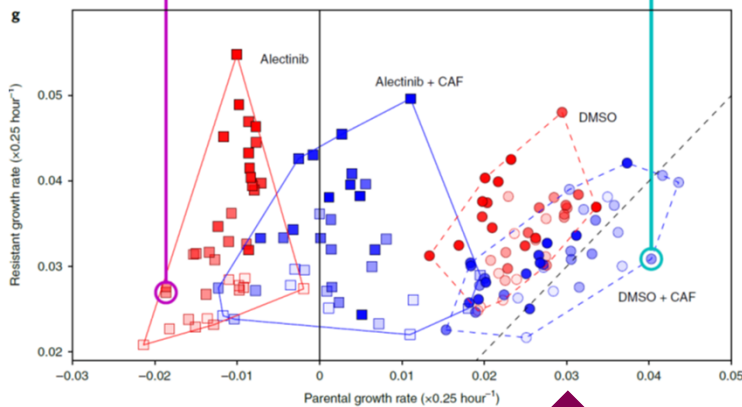


# Competition assays: ALK inhibitor example

Nature Ecology and Evolution 3: 450. 2019

Fibroblasts and alectinib switch the evolutionary games played by non-small cell lung cancer

Artem Kaznatcheev<sup>1,2\*</sup>, Jeffrey Peacock<sup>3</sup>, David Basanta<sup>4</sup>, Andriy Marusyk<sup>5\*</sup> and Jacob G. Scott<sup>2,6\*</sup>



- Growth rate determined using fluorescent protein expression
- Density of drug sensitive parental cell line (opacity of point) alters the growth rate of resistant cells

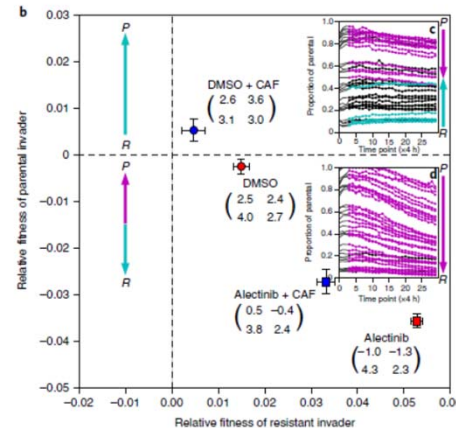
$$P \begin{pmatrix} A & B \\ C & D \end{pmatrix} R \Rightarrow \begin{cases} \frac{d}{dt} N_P = N_P \left( A \frac{N_P}{N_T} + B \frac{N_R}{N_T} \right) \\ \frac{d}{dt} N_R = N_R \left( C \frac{N_P}{N_T} + D \frac{N_R}{N_T} \right) \end{cases}$$

$\hat{w}_P$ : parental growth rate  
 $\hat{w}_R$ : resistant growth rate

$$\Rightarrow \frac{dp}{dt} = p(1-p) \left( \underbrace{(B-D)(1-p)}_{\text{relative fitness of parental invader}} - \underbrace{(C-A)p}_{\text{relative fitness of resistant invader}} \right)$$

gain function for  $p$

where  $N_T = N_P + N_R$  and  $p = \frac{N_P}{N_T}$



Mathematical model where exponential growth rate is dependent on the presence of other clones.

Results suggest proliferation (fitness) of drug resistant cells is altered in the presence of drug sensitive cells



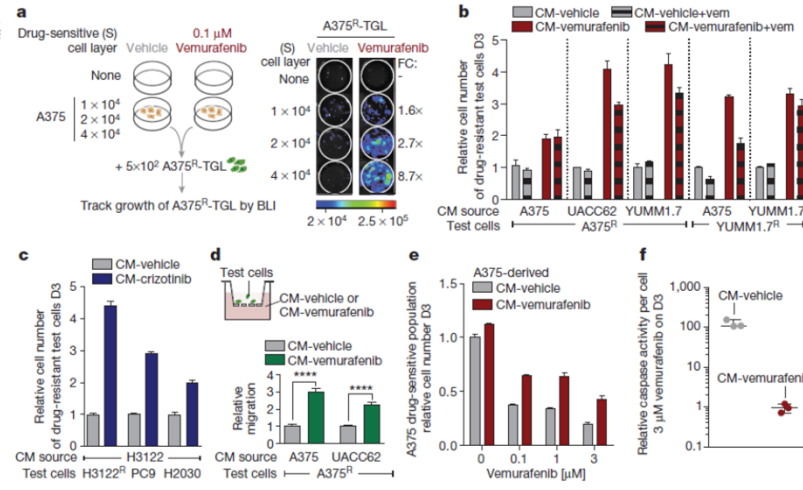
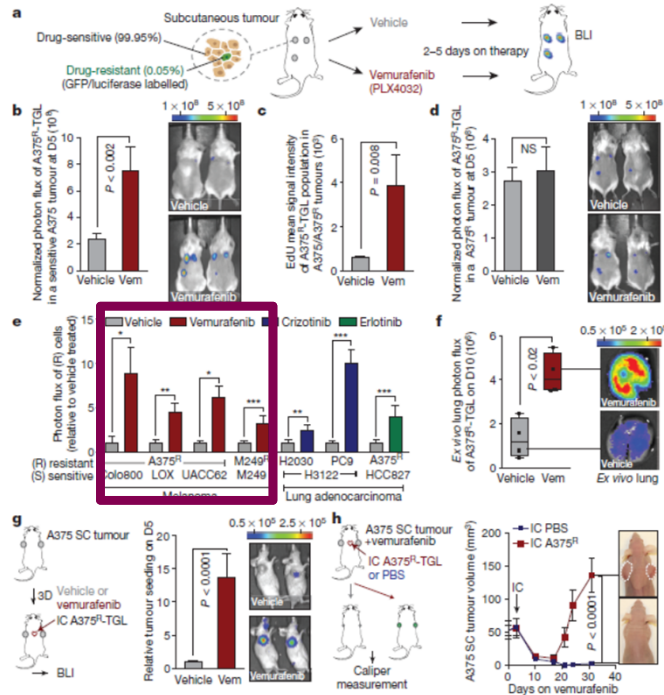
# Competition assays in vitro and in vivo in vivo

LETTER Nature 520: 368. 2015

doi:10.1038/nature14336

## Therapy-induced tumour secretomes promote resistance and tumour progression

Anna C. Obenaus<sup>1</sup>, Yilong Zou<sup>1,2\*</sup>, Andrew L. Ji<sup>4</sup>, Sakari Vanharanta<sup>1,3</sup>, Weiping Shu<sup>1</sup>, Hubing Shi<sup>4</sup>, Xiangju Kong<sup>4</sup>, Marcus C...

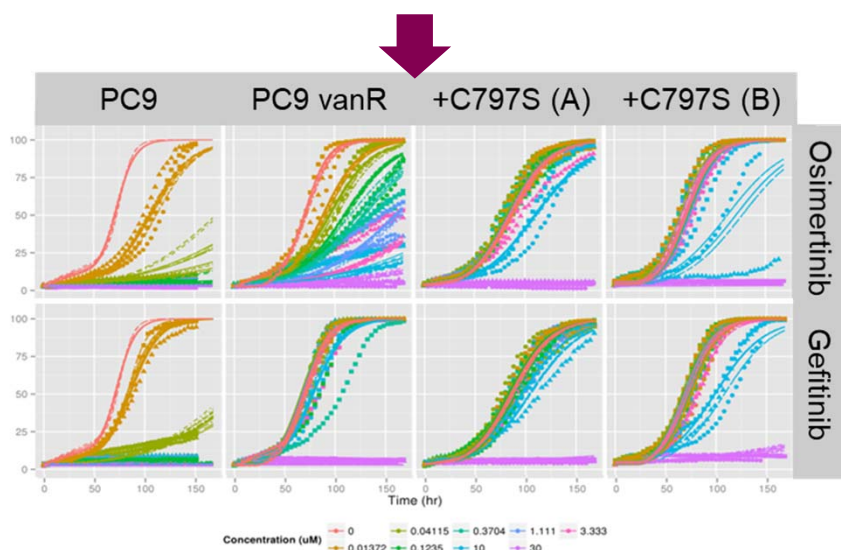


Similar interactions observed for EGFR, BRAF and ALK inhibitors  
 -Resistant cells grow more rapidly in the presence of Sensitive cells and drug

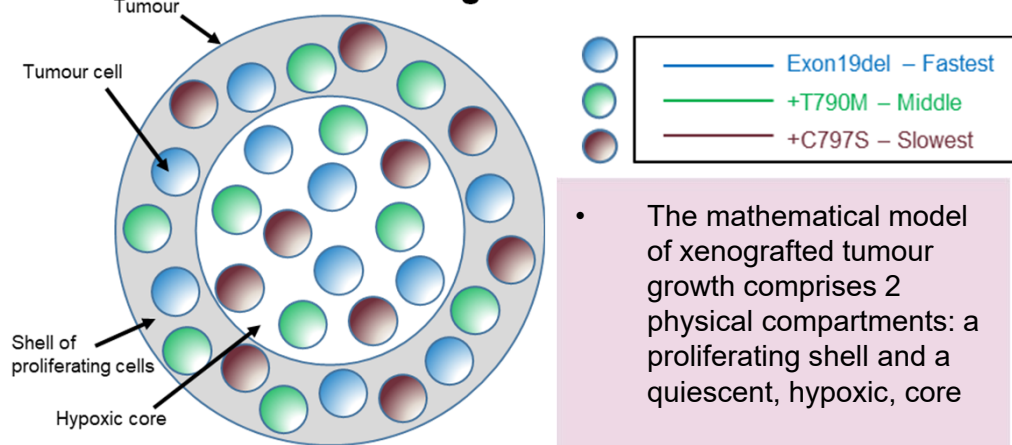


# Model Application: In vitro profiling – In vivo extrapolation

In vitro time course of cell confluence with model fits super-imposed.



Schematic of tumour growth model

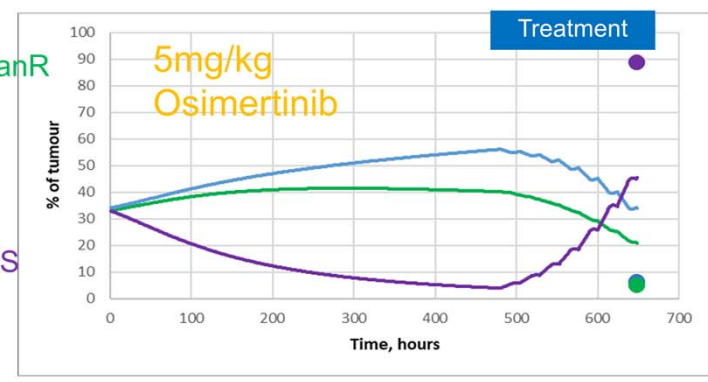
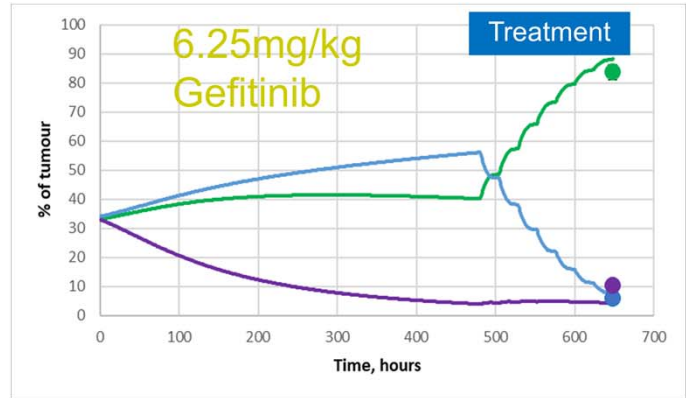
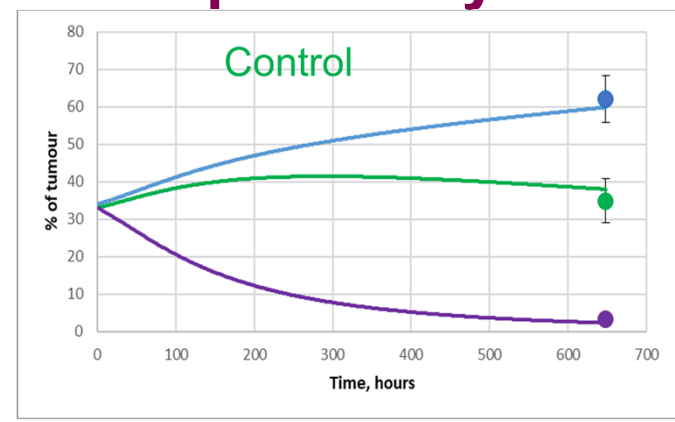
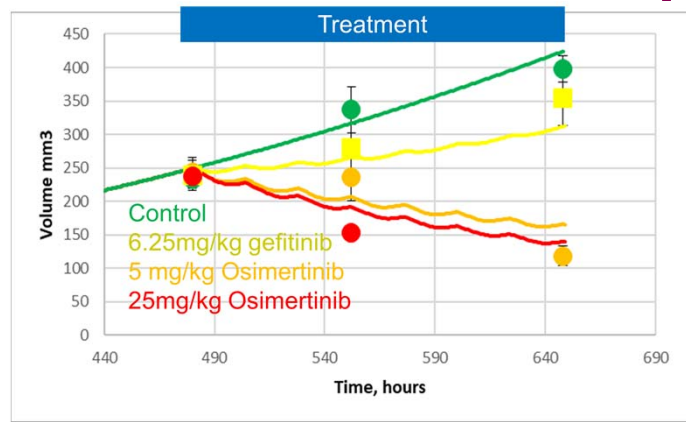


- The mathematical model of xenografted tumour growth comprises 2 physical compartments: a proliferating shell and a quiescent, hypoxic, core
- As the tumour grows the hypoxic region expands, thus the rate of growth reduces
- Intrinsic proliferation rates were set to those estimated from the in vitro experiments above

- PC9 is sensitive to both drugs and grows most rapidly
- PC9vanR is resistant to Gefitinib
- The addition of C797S mutation renders the cells resistant to both drugs but also slow growth

# In vivo experiment: treatment dependent tumour growth kinetics and clonal expansion captured by model

PC9s dominate untreated tumours due greatest proliferation as predicted by in vitro



PC9

PC9 vanR

+C797S

PC9 vanR

+C797S  
PC9

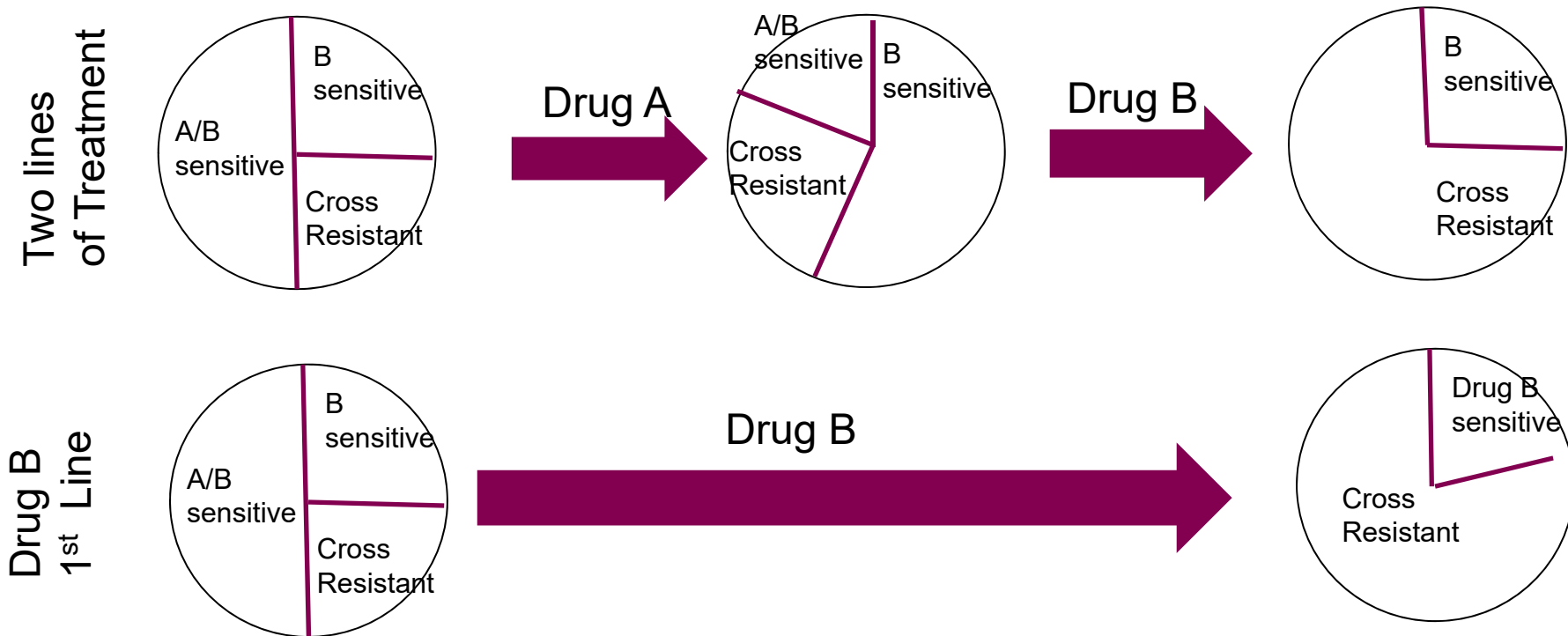
+C797S


PC9  
PC9 vanR



## **Challenges for the future: Modelling multiple lines of therapy**

# Can we integrate clinical efficacy data from multiple lines of treatment to infer evolution in tumours?



<sup>30</sup> Drug development starts in late line then moves to earlier patient populations   
Such modelling would help decision making in which populations are appropriate

## Conclusions

Understanding resistance kinetics will enable

- Optimisation of therapy
- Connection of early efficacy indicators to OS
- Predict the performance of treatments in earlier patient populations

Nonclinical assays and models exist to do this as well as Clinical data analysis



# Acknowledgements

Hitesh Mistry – University of Manchester  
Emma Martin – University of Leicester





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