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# Building kinetic models with complex drug-protein interactions

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ROSA Webinar, 2021, 20<sup>th</sup> January

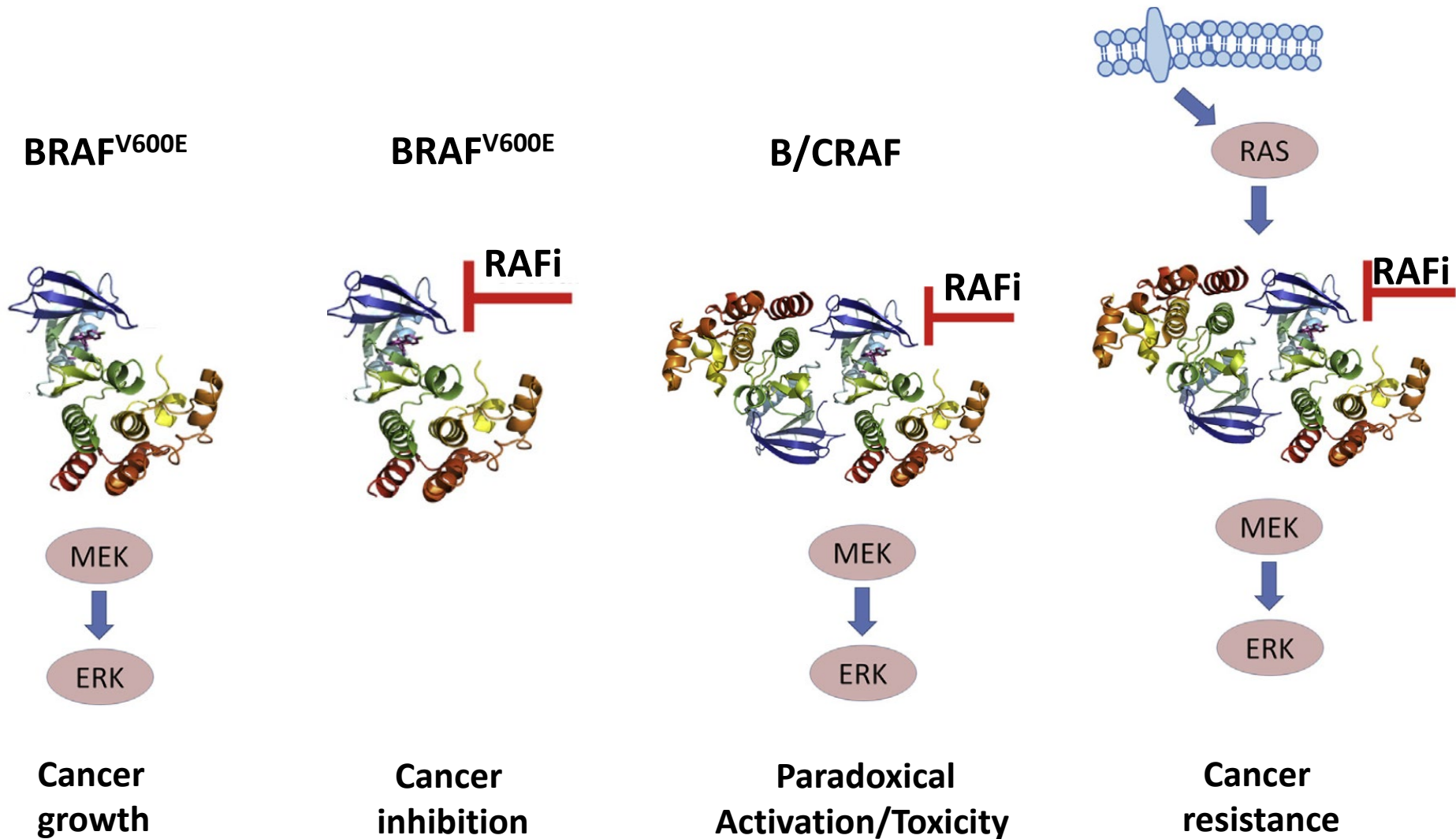
# Outline

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1. A **modeling framework** to build kinetic models with complex drug-protein interactions
2. A **predictive model** for the targeted inhibition of oncogenic MAPK signaling in cancer

# Understanding complex drug-protein interactions is essential for Quantitative Systems Pharmacology

PMID: 26343583



# Complex drug-protein interactions are due to positive and negative cooperativity and must be considered in drug studies

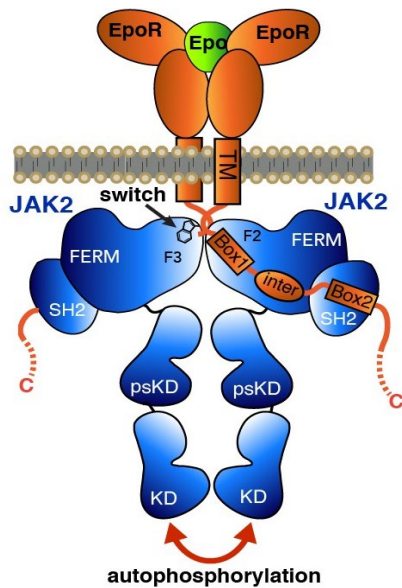
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**Modulate biological functions =  
contribute to drug efficacy**

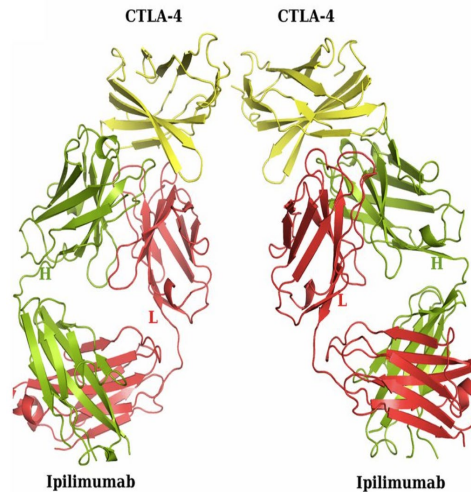
# Complex drug-protein interactions appear in multiple scenarios with therapeutic relevance

**Inhibitors of dimerizing kinases**  
(JAK, EGFR, MEK, etc..)



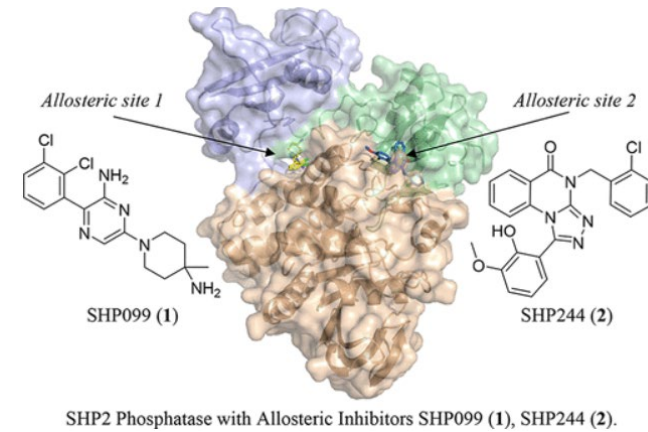
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**Immune checkpoint inhibitors**  
(CTLA4, PD1, etc..)



PMID: 28484017

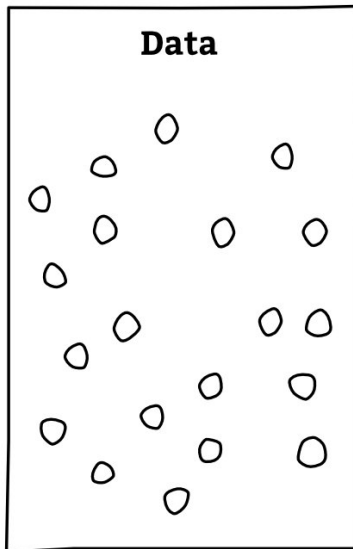
**Inhibitors of other signaling components**  
(SHP2, RAS, etc..)



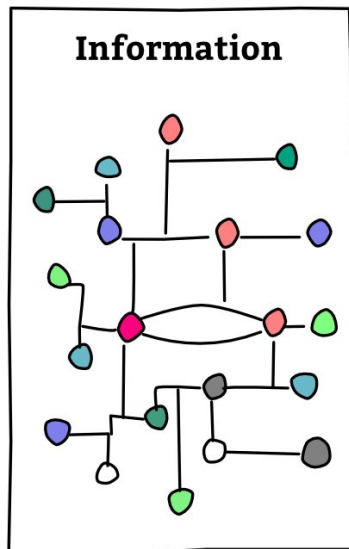
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# Quantitative Systems Pharmacology relies on kinetic modeling to describe and understand drug action

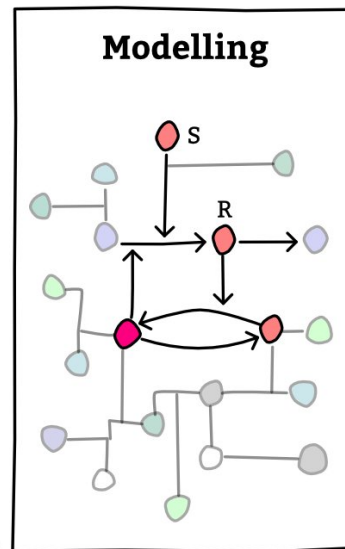
## QSP workflow



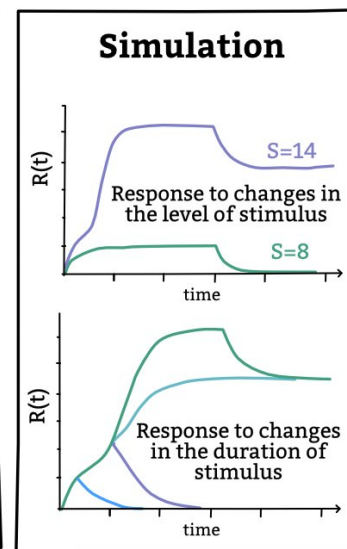
... from a molecular system



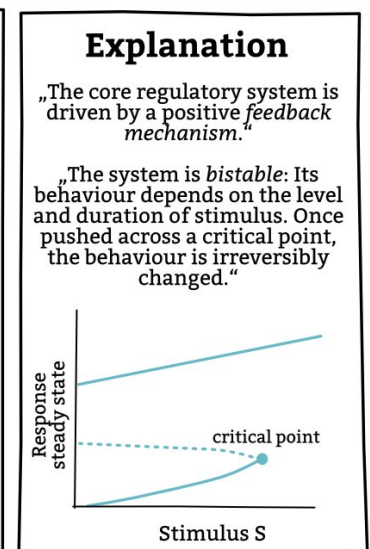
... enriched with bioinformatics



... using a systems biology approach



... to understand its (functioning) dynamics



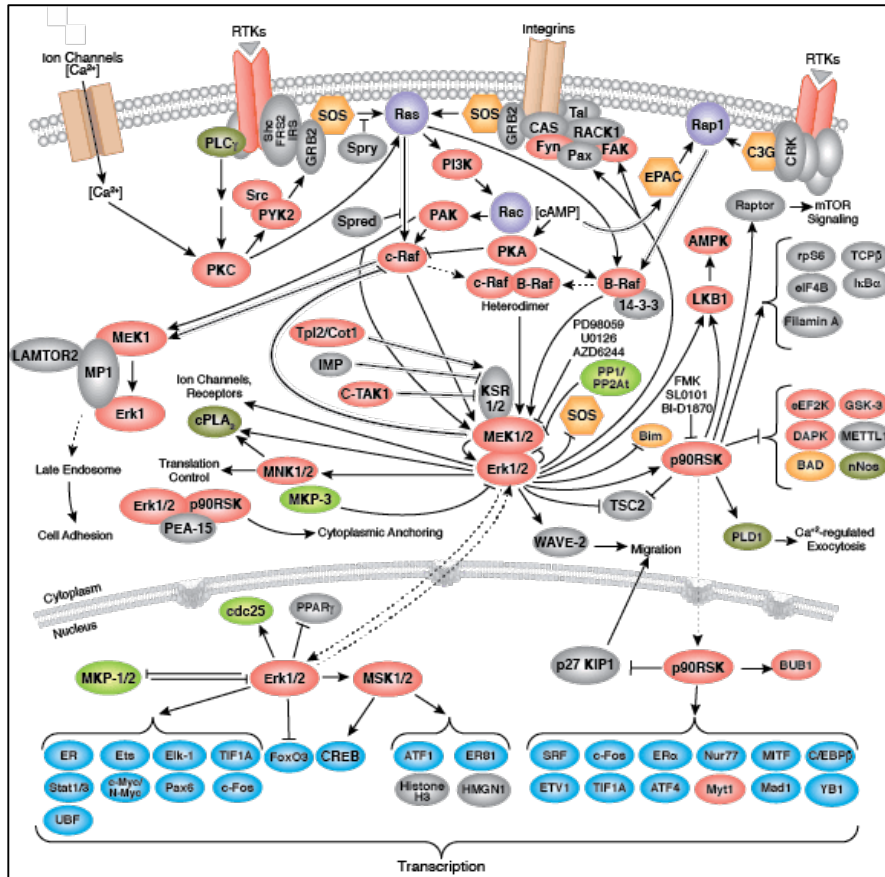
@OlafWolkenhauer

**i.e. Kinetic modeling**

# Kinetic modelling requires writing ODE systems that properly describe molecular complexity of biological system

Biological system

Ordinary differential equation systems



MAPK signaling pathway – from CST

$$\frac{d[\text{pEGFR}]}{dt} = k_{\text{on}} * [\text{EGFR}] * [\text{EGF}] - \dots$$

$$\frac{d[\text{Ras-GTP}]}{dt} = k_{\text{on}} * [\text{Ras}] * [\text{GTP}] - \dots$$

$$\frac{d[\text{RAF}_2]}{dt} = k_{\text{on}} * [\text{RAF}]^2 - k_{\text{off}} * [\text{RAF}_2] + \dots$$

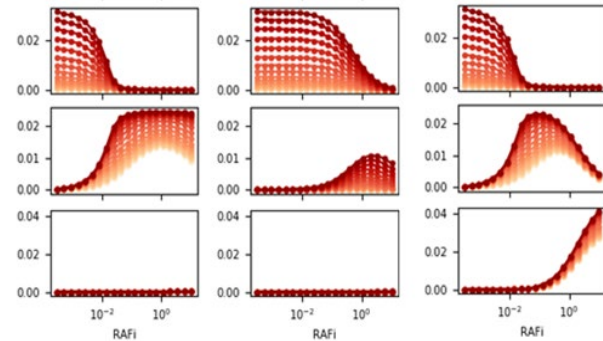
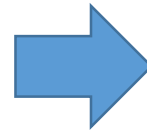
$$\frac{d[\text{BRAF}^{\text{V600E}}]}{dt} = k_{\text{exp}} - k_{\text{deg}} * [\text{BRAF}^{\text{V600E}}] + \dots$$

$$\frac{d[\text{pMEK}]}{dt} = k_{\text{on}} * [\text{MEK}] * [\text{BRAF}^{\text{V600E}}] - \dots$$

$$\frac{d[\text{pERK}]}{dt} = k_{\text{on}} * [\text{ERK}] * [\text{pMEK}] - \dots$$

$$\frac{d[\text{CyclinD1}]}{dt} = k_{\text{exp}} - k_{\text{deg}} * [\text{CyclinD1}] + \dots$$

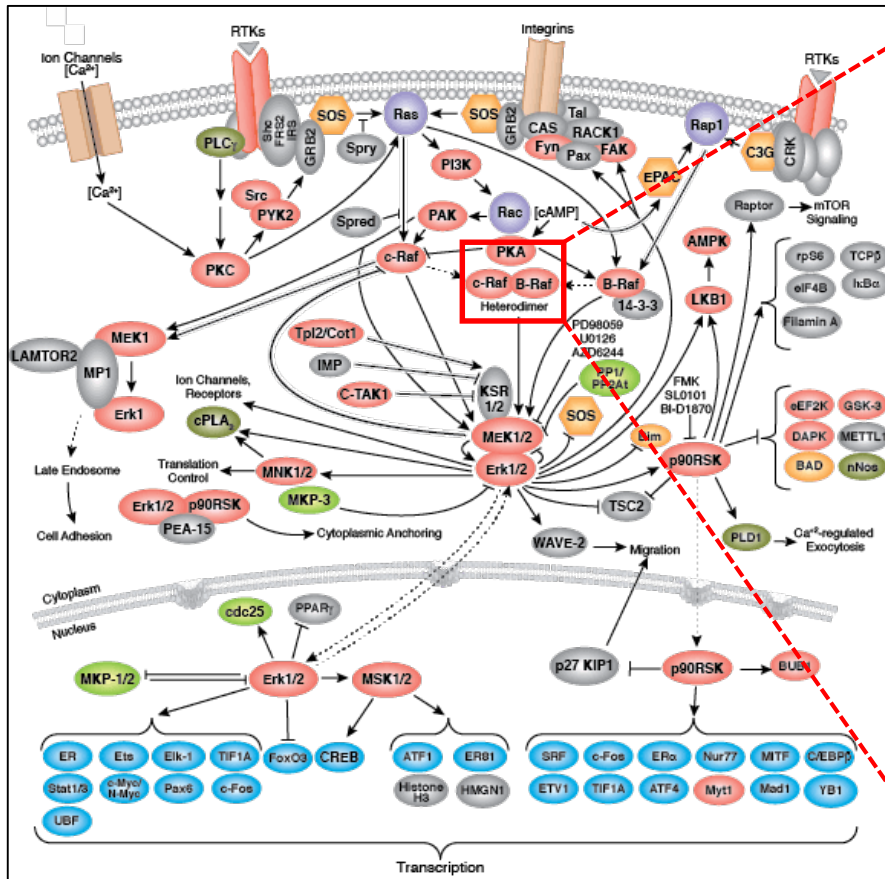
**Kinetic model**



# Kinetic modeling for QSP requires describing complex drug-protein interactions

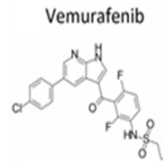
Pathway interactions

Drug-protein and protein-protein interactions

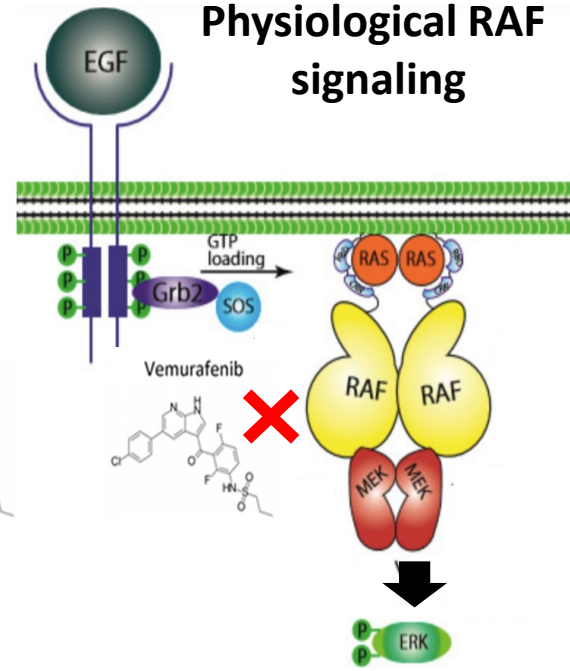


MAPK signaling pathway – from CST

1)  
BRAF<sup>V600E</sup>  
oncogene



2)  
Physiological RAF  
signaling



Degirmenci et al, 2020 Cells



# A toy example of RAF and RAF inhibitors to introduce a framework for kinetic modeling with energy description

Components:

**R**: RAF (target)

**I**: RAF Inhibitor (drug)

Binding schema:



Reaction

Dissociation constant

Thermodynamic factors



$$K_{RI}$$



$$K_{RR}$$



$$f \cdot K_{RI}$$



$$g \cdot K_{|RR|}$$

$f = K_{RR|} / K_{RI}$  fold change in drug affinity to 1<sup>st</sup> RAF in dimer

$g = K_{|RR|} / K_{RI}$  fold change in drug affinity to 2<sup>nd</sup> RAF in dimer

*Cooperative reactions*

*Kholodenko, 2015, Cell Reports*

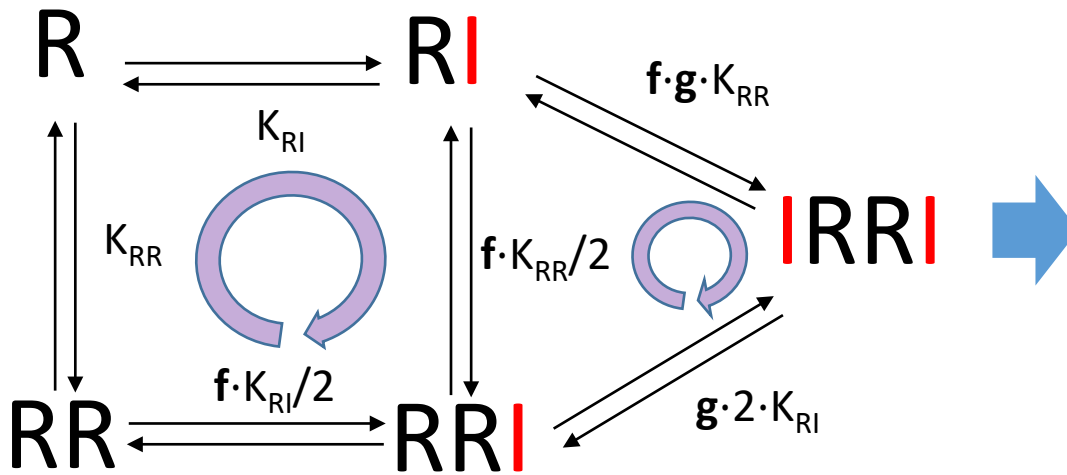
# The two problems in properly describing drug-protein interactions: combinatorial and contextual complexity

## 1. Combinatorial complexity:

the explosion of the reaction network size due to combinations of multi-domain protein and drug interactions

## 2. Contextual complexity:

the constraints in cooperative reaction rates due to context-dependency of cooperative interactions



## ODE model

$$d[R]/dt = k_{RR}^- [RR] + \dots$$

$$d[I]/dt = k_{RI}^- [RI] + \dots$$

$$d[RI]/dt = k_{RI}^+ [R][I] + \dots$$

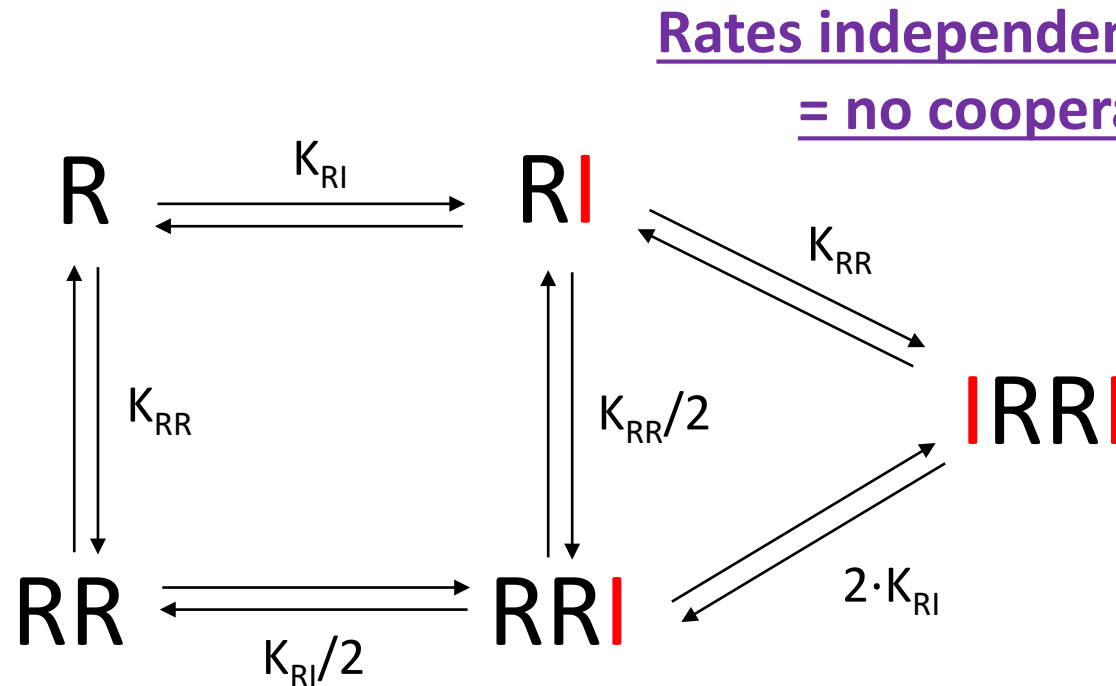
$$d[RRI]/dt = f k_{RI}^+ [RR][I] + \dots$$

$$d[IRRI]/dt = fgk_{RI}^+ [IRR][I] + \dots$$

Detailed balance must be satisfied!

# Classic rule-based modeling solves combinatorial but not contextual complexity problem

Reaction rules:



# How to solve the problem of building kinetic models with combinatorial and contextual complexity?

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Solution:

**Rule-based modeling  
with energy descriptions\***

*\*Hogg JS, PhD Thesis, 2013, Faeder lab@UPitt*

# Rule-based modeling with energy descriptions solves combinatorial and contextual complexity at once

Reaction rules:



Baseline energies (kJ/mol):

$$E_0^{RR} = \emptyset \cdot \log(K_{RR}) - \log(k_{RR}^+)$$

$$E_0^{RI} = \emptyset \cdot \log(K_{RI}) - \log(k_{RI}^+)$$

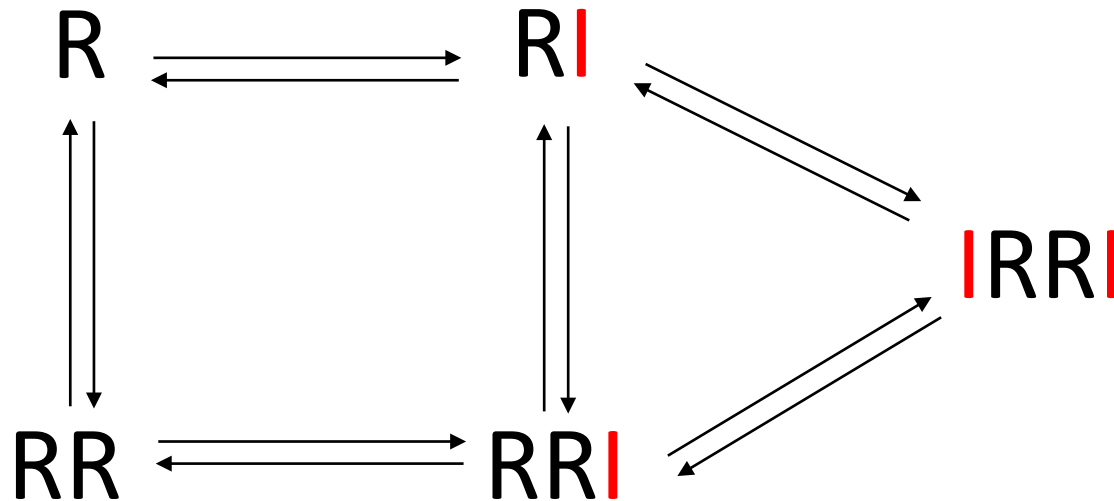
Energy Patterns (kJ/mol):

$$ep(RR) = \log(K_{RR})$$

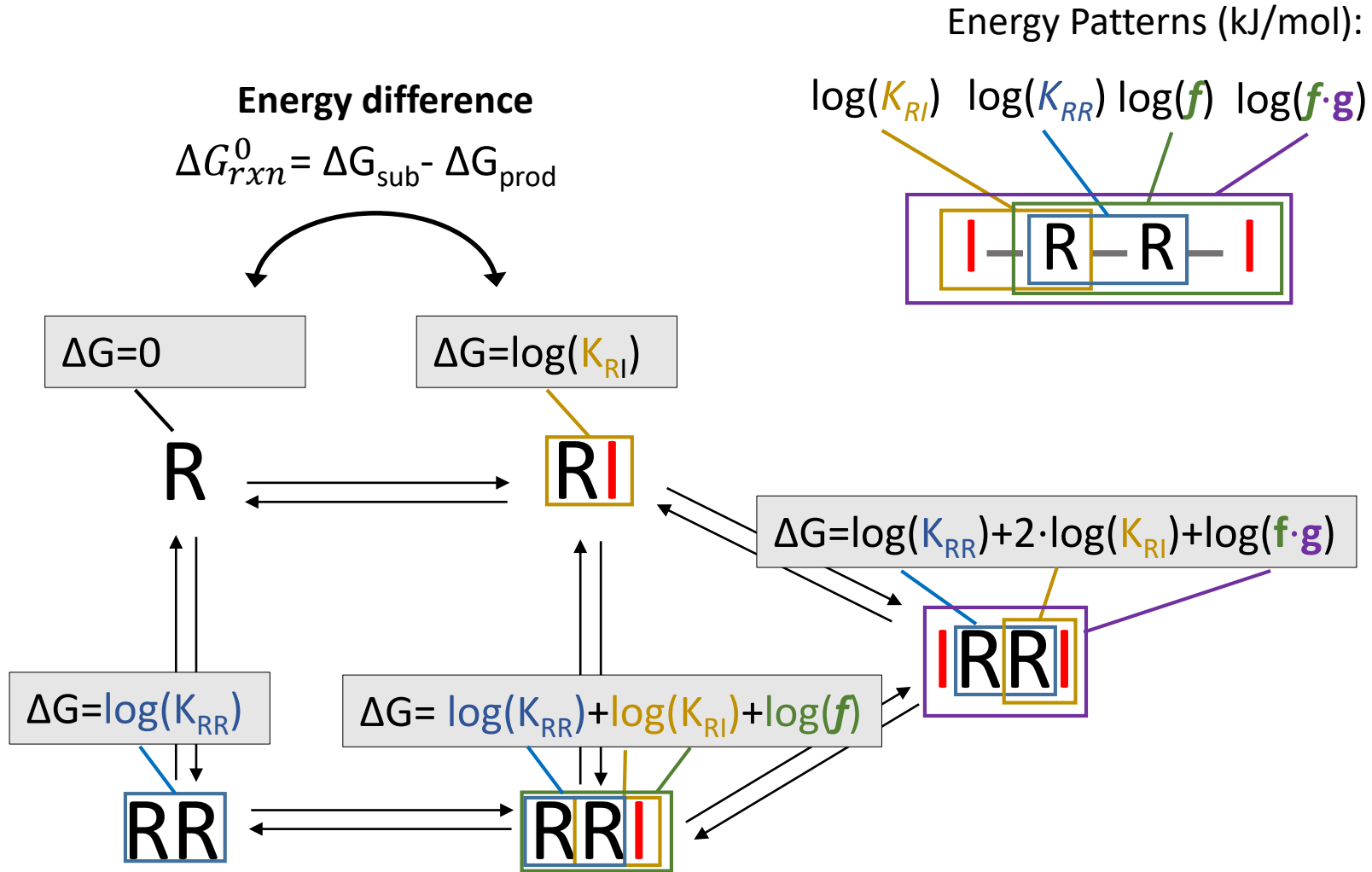
$$ep(RI) = \log(K_{RI})$$

$$ep(RRI) = \log(f)$$

$$ep(IRRI) = \log(f \cdot g)$$

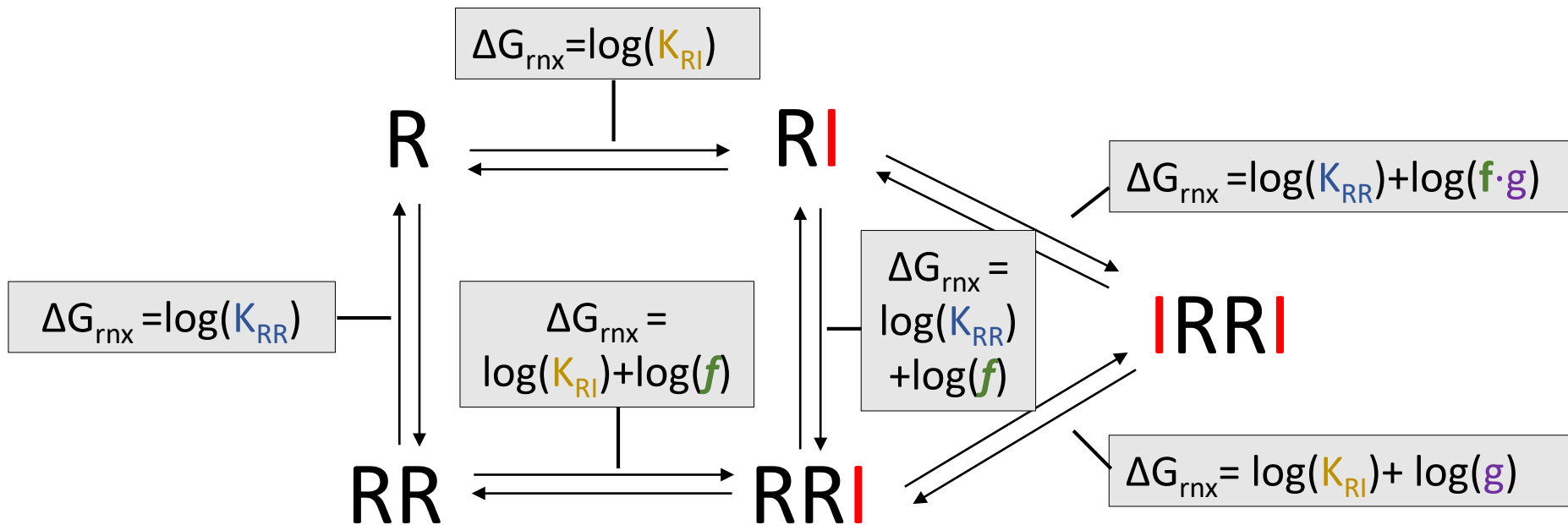
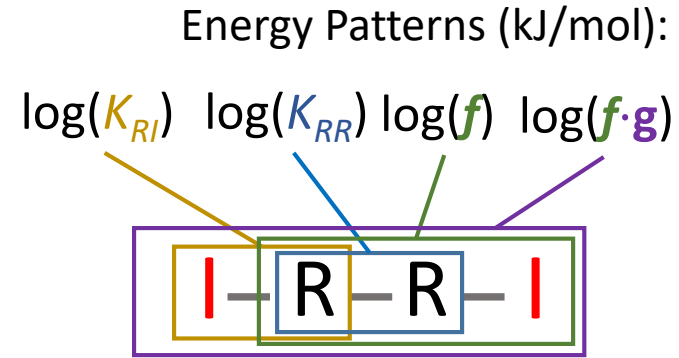


# Rule-based modeling with energy descriptions solves combinatorial and contextual complexity at once



# Rule-based modeling with energy descriptions solves combinatorial and contextual complexity at once

Energy difference  
 $\Delta G_{rxn}^0 = \Delta G_{sub} - \Delta G_{prod}$



# The theory behind the energy formulation in BioNetGen

Biological Network Generator

$$E_0$$

baseline activation  
energy

$$\Delta G_{rxn}^0$$

free energy  
change

$$\phi$$

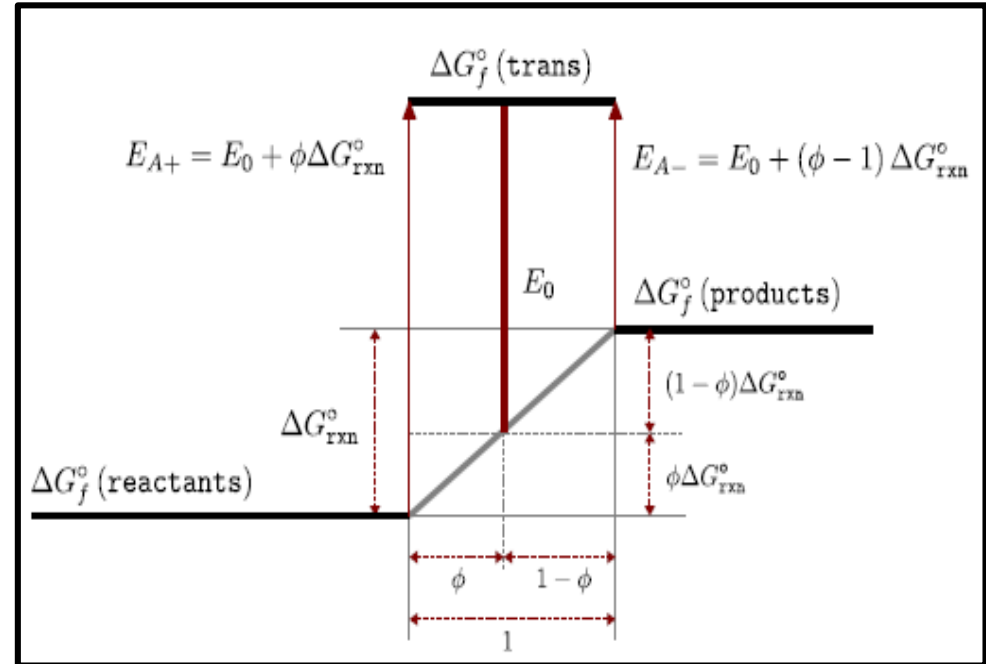
rate  
distribution

**Linear transition state theory.** The activation energy is linearly related to the standard change in free energy of a reaction:

$$E_A = E_0 + \phi \cdot \Delta G_{rxn}^0$$

**Arrhenius equation.** The reaction rate constant of a reaction is determined by the activation energy:

$$k = C \cdot \exp\left(\frac{-E_A}{RT}\right)$$



**Conversion rules from energies to kinetics:**

$$k_{on} = C \cdot \exp\left(\frac{-E_0 + \phi \cdot \Delta G_{rxn}^0}{RT}\right)$$

$$k_{off} = C \cdot \exp\left(\frac{-E_0 + (\phi - 1) \cdot \Delta G_{rxn}^0}{RT}\right)$$

From Justin Hogg PhD Thesis 2012 &  
Sekar et al 2017 BIBM -Faeder Lab@UPitt



# Rule-based modeling with energy descriptions solves combinatorial and contextual complexity at once

Baseline energies (kJ/mol):

$$E_0^{RR} = \phi_{RR} \cdot \log(K_{RR}) - \log(k_{RR}^+)$$

$$E_0^{RI} = \phi_{RI} \cdot \log(K_{RI}) - \log(k_{RI}^+)$$

Rate distributions:

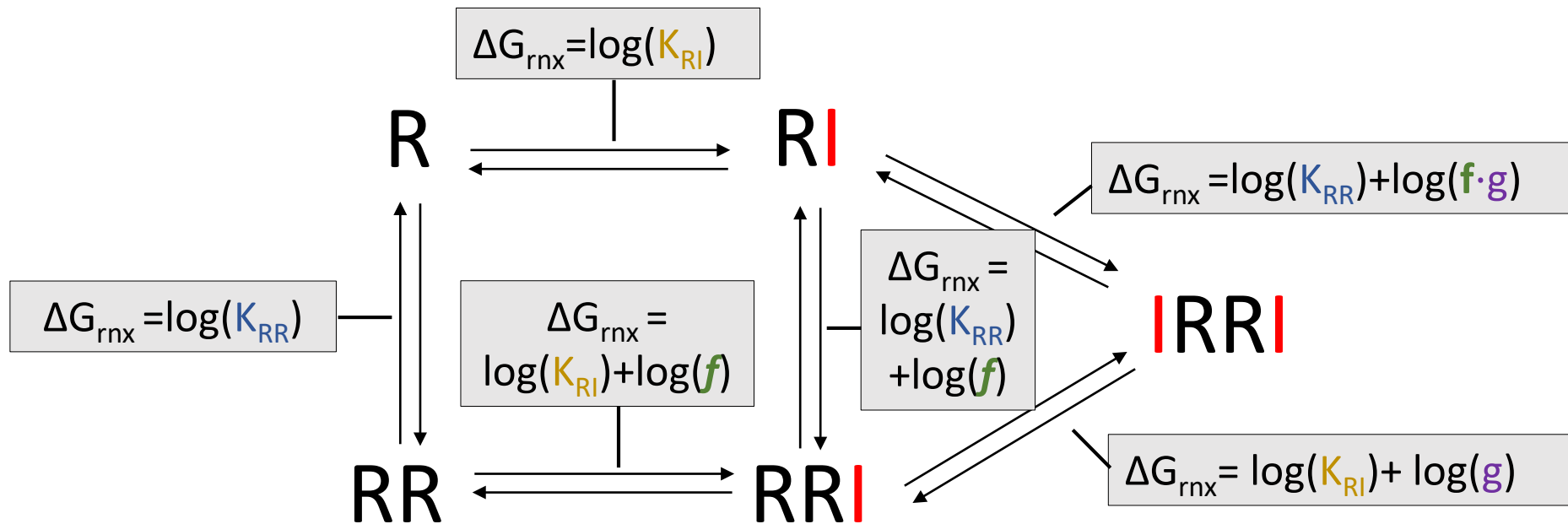
$$\phi_{RR}$$

$$\phi_{RI}$$

Conversion rules:

$$k_{on} = C \cdot \exp\left(\frac{-E_0 + \phi \cdot \Delta G_{rxn}^0}{RT}\right)$$

$$k_{off} = C \cdot \exp\left(\frac{-E_0 + (\phi - 1) \cdot \Delta G_{rxn}^0}{RT}\right)$$



# Rule-based modeling with energy descriptions solves combinatorial and contextual complexity at once

Baseline energies (kJ/mol):

$$E_0^{RR} = \phi_{RR} \cdot \log(K_{RR}) - \log(k_{RR}^+)$$

$$E_0^{RI} = \phi_{RI} \cdot \log(K_{RI}) - \log(k_{RI}^+)$$

Rate distributions:

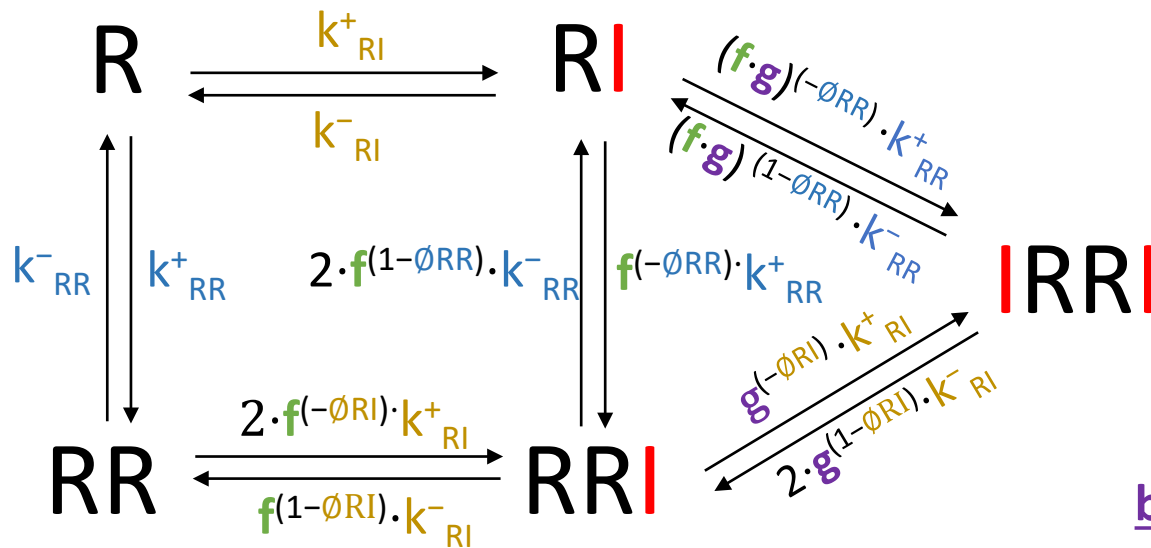
$$\phi_{RR}$$

$$\phi_{RI}$$

Conversion rules:

$$k_{on} = C \cdot \exp\left(\frac{-E_0 + \phi \cdot \Delta G_{rxn}^0}{RT}\right)$$

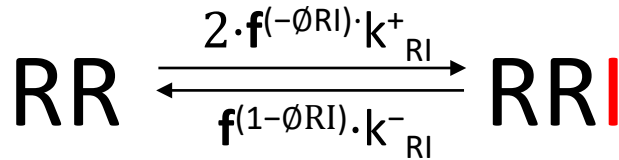
$$k_{off} = C \cdot \exp\left(\frac{-E_0 + (\phi - 1) \cdot \Delta G_{rxn}^0}{RT}\right)$$



Detailed balance is SATISFIED by construction!

# Rule-based modeling with energy descriptions solves combinatorial and contextual complexity at once

Automatically derived rates:



Dissociation constant:

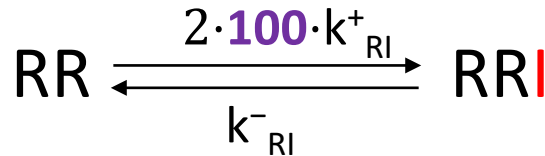
$$\frac{k^-}{k^+} = \frac{f^{(1-\emptyset_{RI})} \cdot k_{RI}^-}{2 \cdot f^{(-\emptyset_{RI})} \cdot k_{RI}^+} = \frac{f^{(1-\emptyset_{RI}+\emptyset_{RI})} \cdot k_{RI}^-}{2 \cdot k_{RI}^+} = \frac{f \cdot K_{RI}}{2}$$

Thermodynamic factor

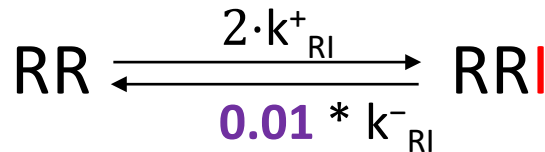
Distribution rate

$f=0.01$

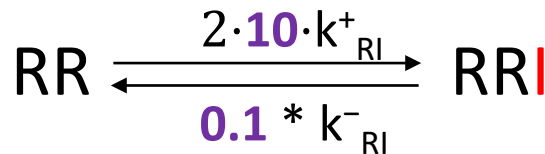
$\emptyset_{RI}=1$  (on  $k^+$ )



$\emptyset_{RI}=0$  (on  $k^-$ )



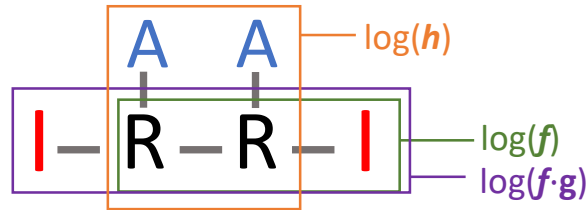
$\emptyset_{RI}=0.5$  (shared)



$$\frac{0.01 \cdot K_{RI}}{2}$$

# PySB with support for eBNG allows to write energy-balanced kinetic models as compact python programs

R: RAF  
 I: RAFi  
 A: Ras<sup>GTP</sup>



**PySB** Systems biology modeling in Python **BioNetGen** Biological Network Generator

PySB extended with energy BioNetGen  
 by *Jeremy Muhlich*

## PySB code (Python)

```
#Rules
Rule('RR', R(r=None)+R(r=None)<>R(r=1)%R(r=1), RR_phi, Ea0_RR)
Rule('RI', R(i=None)+I(r=None)<>R(i=1)%I(r=1), RI_phi, Ea0_RI)
Rule('RA', R(a=None)+A(r=None)<>R(a=1)%A(r=1), RA_phi, Ea0_RA)

#Energy Patterns
EnergyPattern('ep_RR', R(r=1)%R(r=1), Gf_RR)
EnergyPattern('ep_RI', R(i=1)%I(r=1), Gf_RI)
EnergyPattern('ep_RA', R(i=1)%I(r=1), Gf_RI)
EnergyPattern('ep_IRR', I(r=1)%R(r=2)%R(r=2,i=None), ep_IRR)
EnergyPattern('ep_IRRI', I(r=1)%R(i=1,r=2)%R(r=2,i=3)%I(r=3), ep_IRR)
EnergyPattern('ep_ARRA', A(r=1)%R(a=1,r=2)%R(r=2,a=3)%A(r=3), ep_ARRA)

#Expressions
Expression('Gf_RR', log(RR_kD))
Expression('Gf_RI', log(RI_kD))
Expression('Gf_RA', log(RA_kD))
Expression('Ea0_RR', -(RR_phi*log(RR_kD)+log(RR_kf)))
Expression('Ea0_RI', -(RI_phi*log(RR_kD)+log(RI_kf)))
Expression('Ea0_RA', -(RA_phi*log(RR_kD)+log(RA_kf)))
Expression('ep_IRR', log(f))
Expression('ep_IRR', log(g)+log(f))
Expression('ep_ARRA', log(h))
```

## ODE system

```
In [204]: model
Out[204]: <Model 'pysb_energy.model.example1'
          (monomers: 3, rules: 3,
           parameters: 13,
           expressions: 32)

In [205]: len(model.parameters)
Out[205]: 13

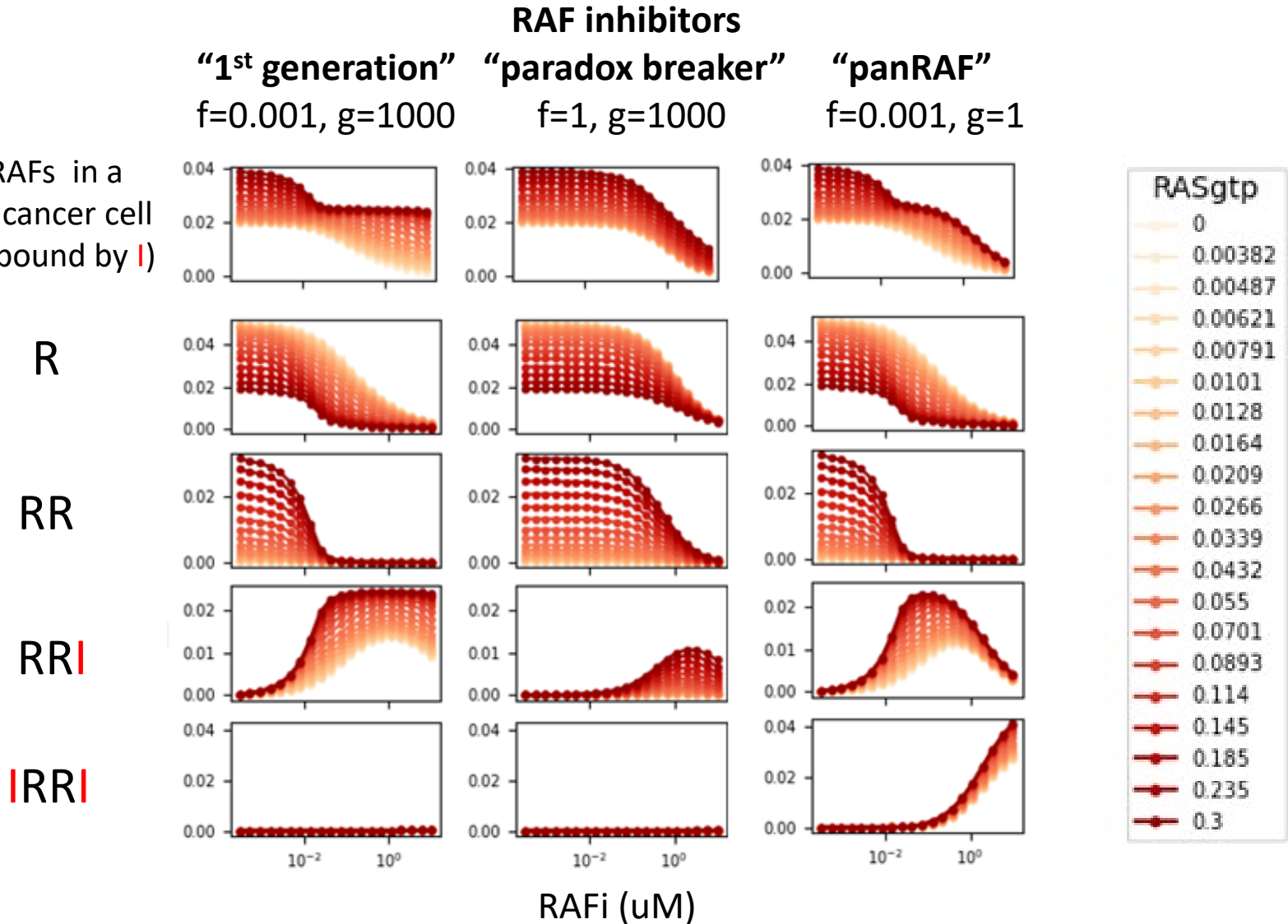
In [206]: len(model.species)
Out[206]: 16

In [207]: len(model.reactions)
Out[207]: 60
```



# A toy kinetic model with energies recapitulates the efficacy of different classes of RAF inhibitors

Active RAFs in a  
 BRAF<sup>V600E</sup> cancer cell  
 (all R not bound by I)



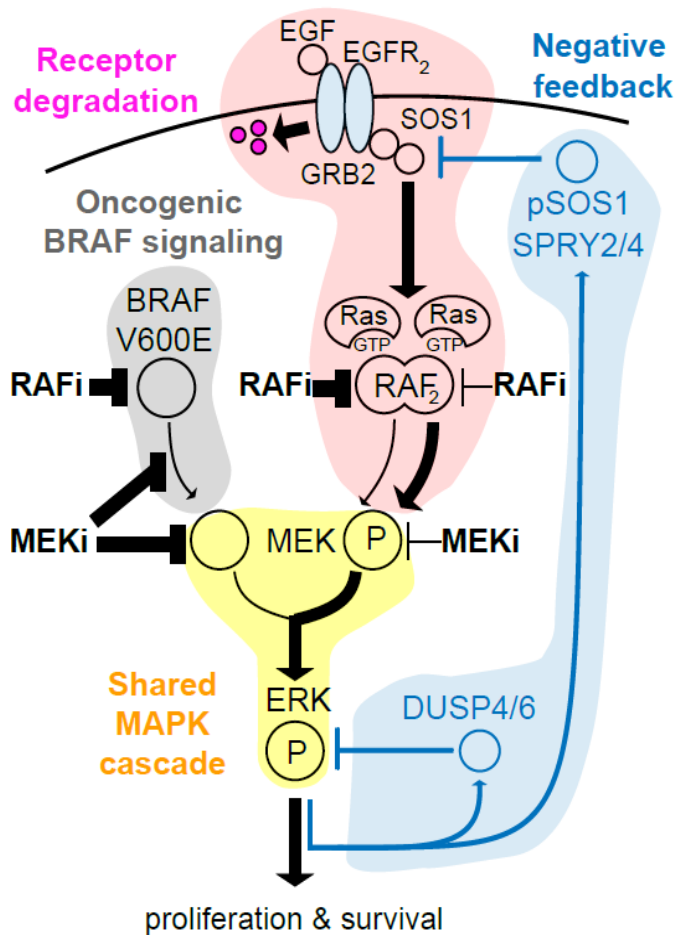
# Conclusions on rule-based modelling framework with energies

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- Remapping kinetic parameters as energies using energy patterns allows to solve the combinatorial and contextual complexity of kinetic models at once
- Convenient description of high-order cooperativity in drug-protein and protein-protein interactions
- Applicable to much more complex biological descriptions with multiple high-order cooperativities as for example:
  - Mutations, PTMs, asymmetric complexes, localization, etc..
- Newly implemented support for energy-BNG in PySB allows to write kinetic models with energy description as compact Python programs

# Construction of a mechanistic model of MAPK signaling with high-order cooperativity in drug-protein interactions

MAPK signaling cascade  
in BRAF-mutant cancer



Proteins: **11**

Drugs: **2**

Reaction Rules: **54**

Energy Patterns: **27**

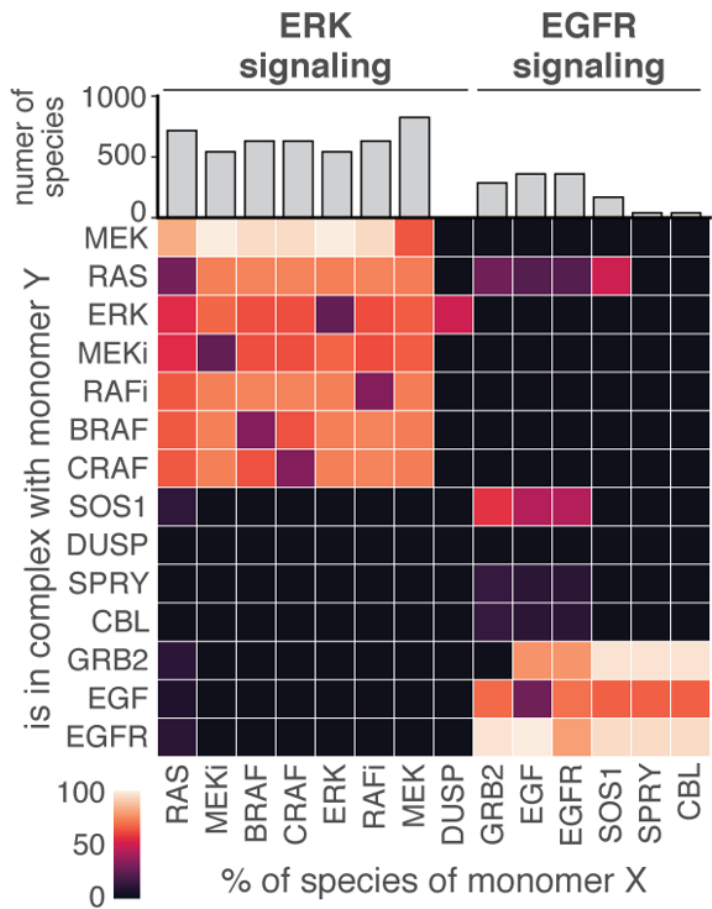
Parameters: **63**

Protein Complexes (ODEs): **1007**

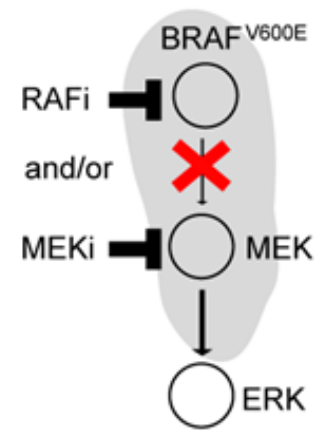
Reactions: **13164**

# The MAPK signaling model with energy descriptions properly accounts for complexity of protein and drug-protein assembly

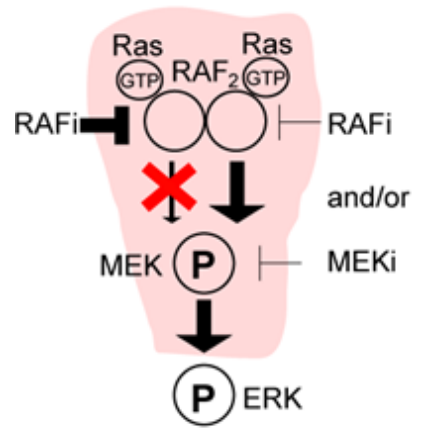
Interaction diagram



Examples of complex drug-protein species



Sensitive to  
RAFi and/or MEKi

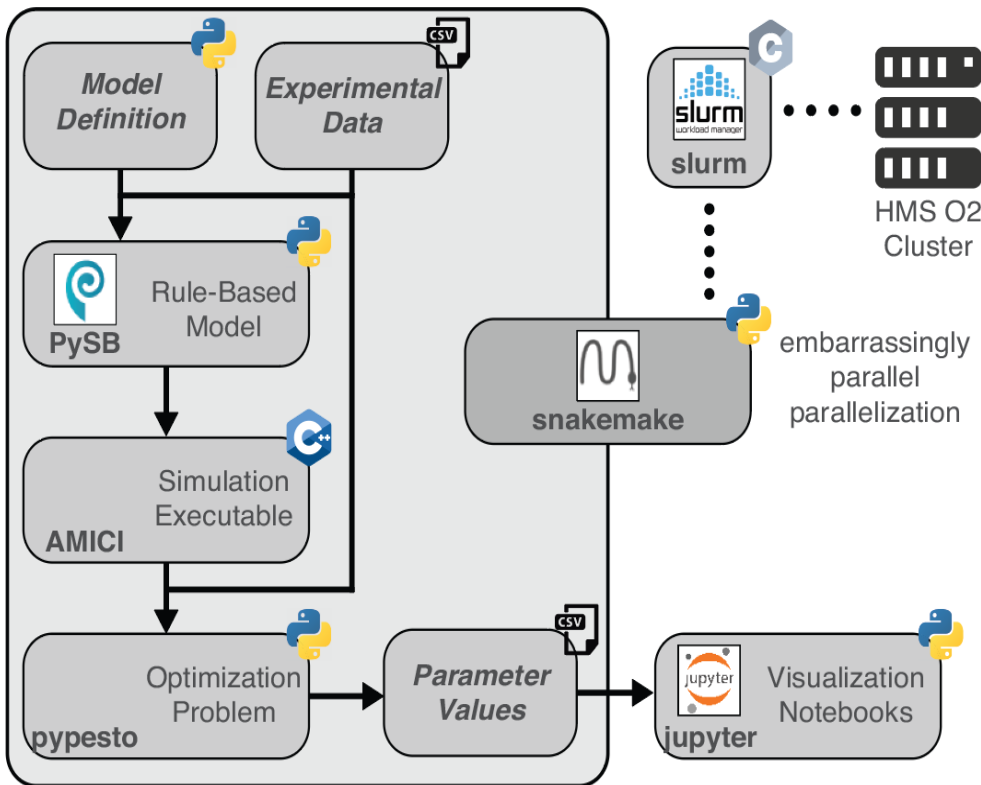


Resistant to  
RAFi and/or MEKi

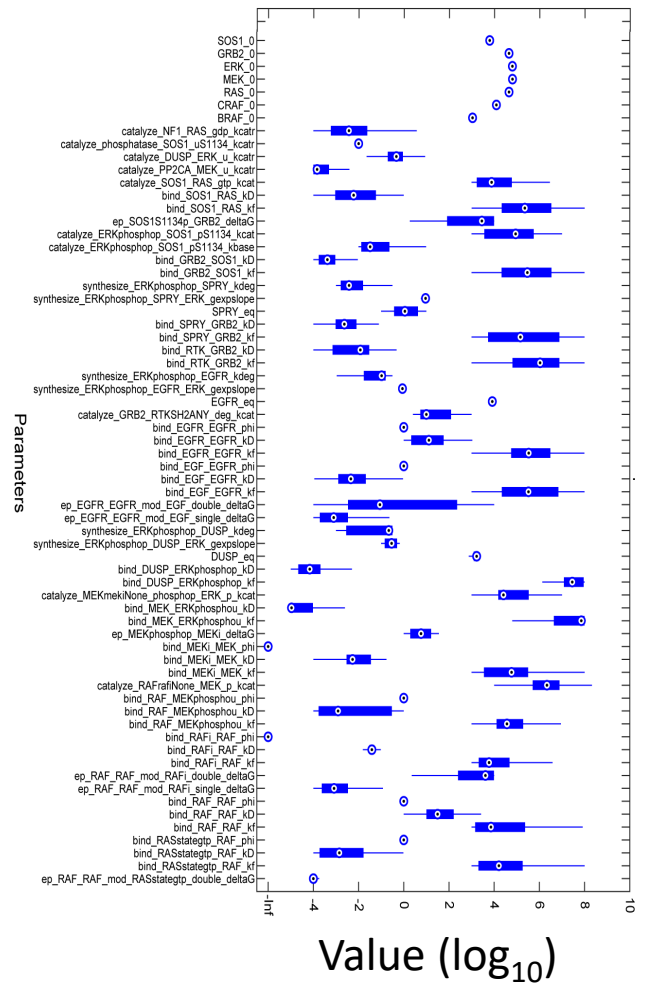


# Large-scale parameter fitting provides estimations for the 63 kinetic and energy parameters determining MAPK signaling

## Parameter fitting pipeline



## 63 estimated parameters



by Fabian Fröhlich

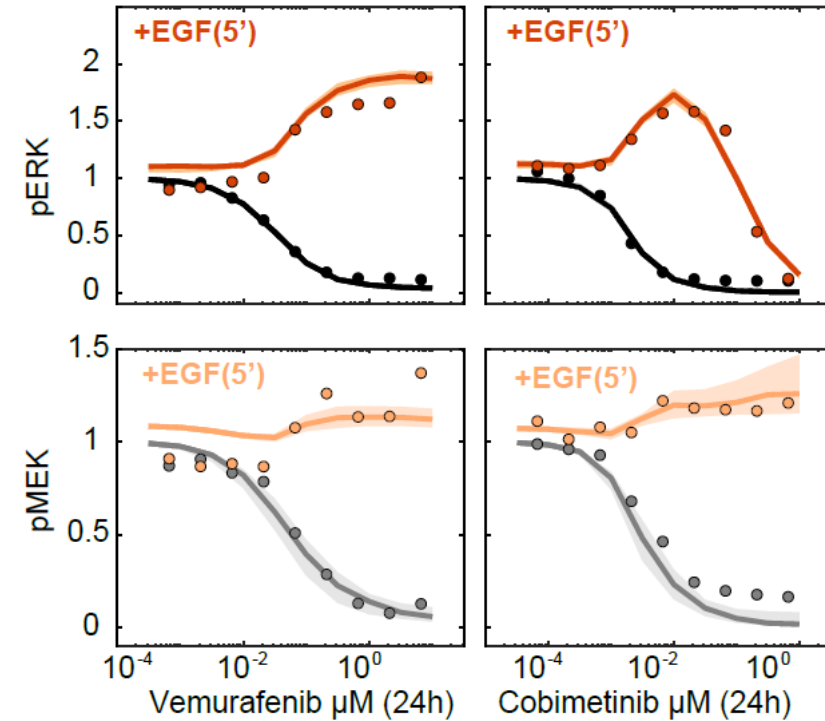
# Kinetic parameters show that drug resistance to RAF and MEK inhibitors is due to RAF dimers assembled by receptor activation

## Model fit to data

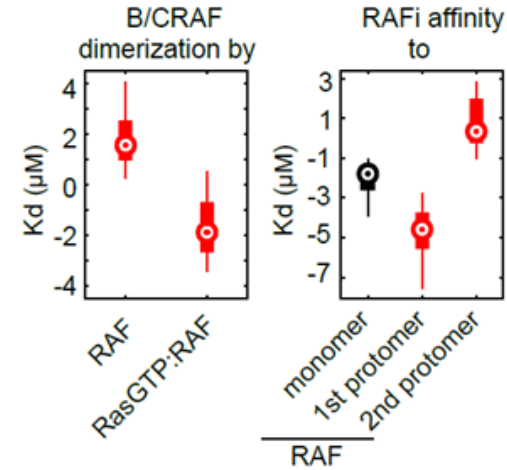
## Parameters

### Dose-Response

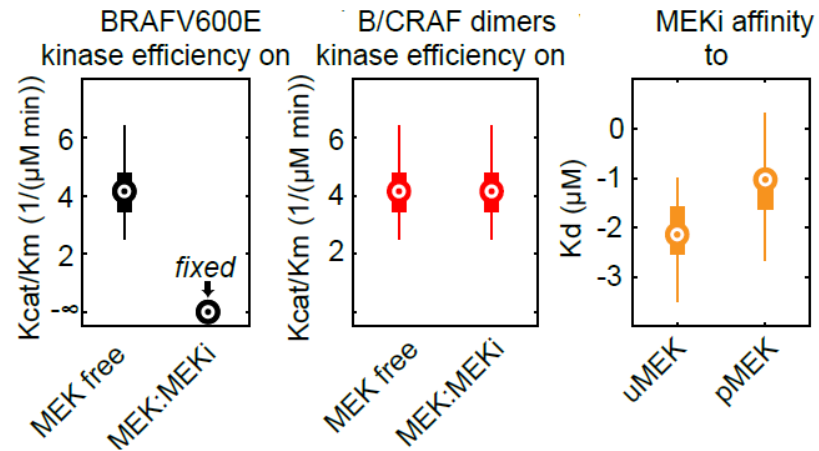
● Experiments — Model fit



A375 melanoma cell line

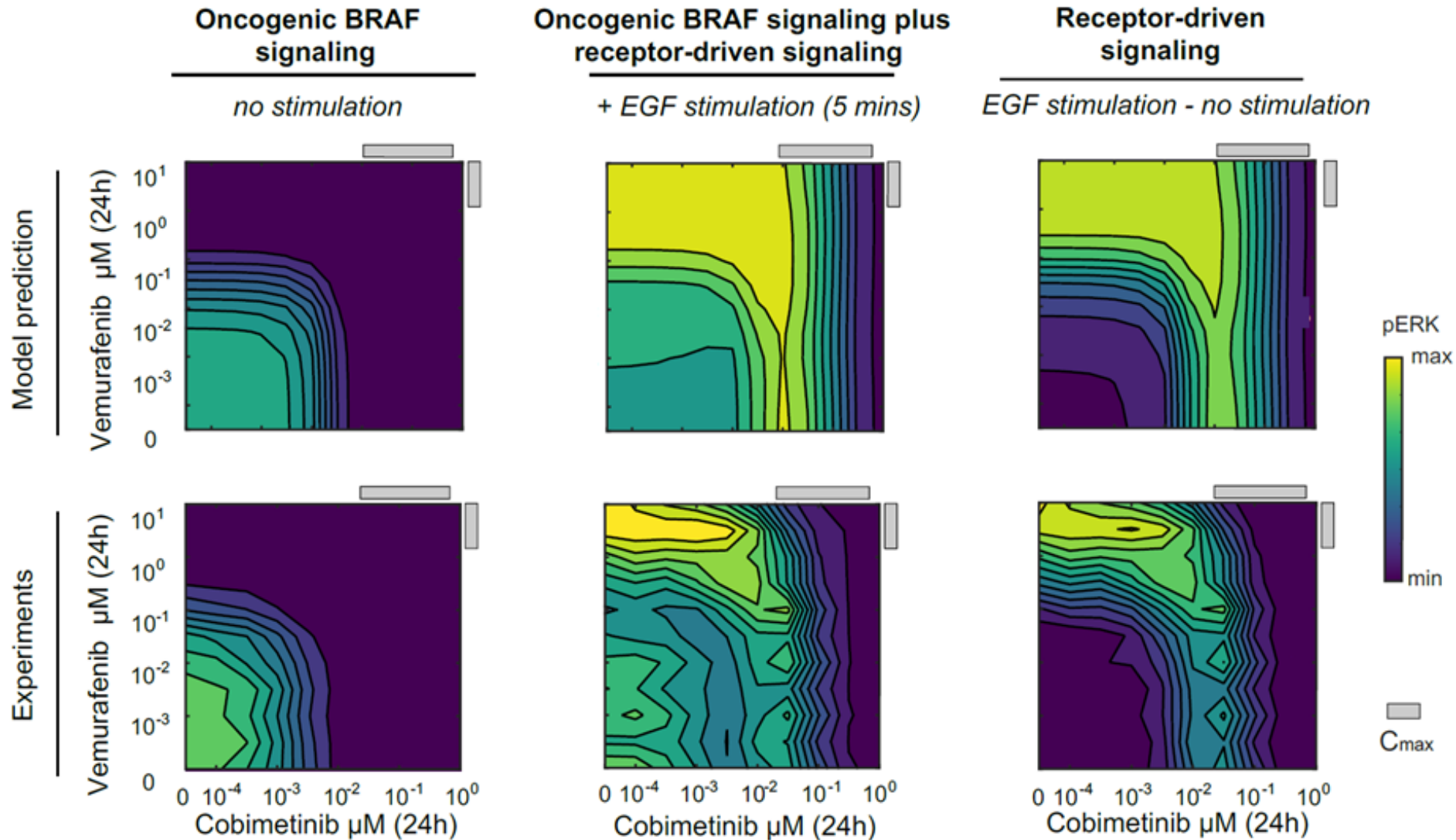


RAS-RAF-RAFi



MEK-MEKi

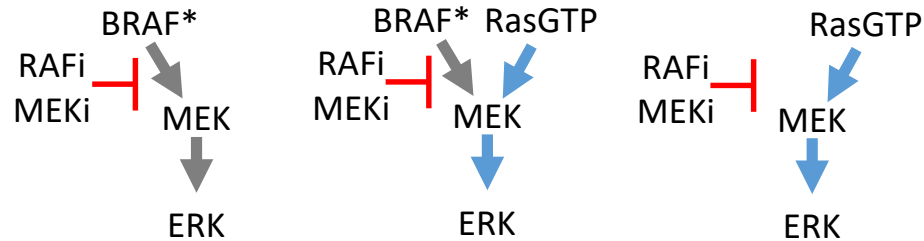
# Modelling predicts wide dose range of combined RAF/MEK inhibition in which receptor-driven ERK signaling is possible



A375 melanoma cell line

# RasGTP activation by receptors or mutations is resistant combined RAF and MEK inhibitors

Experimental validation of unified model

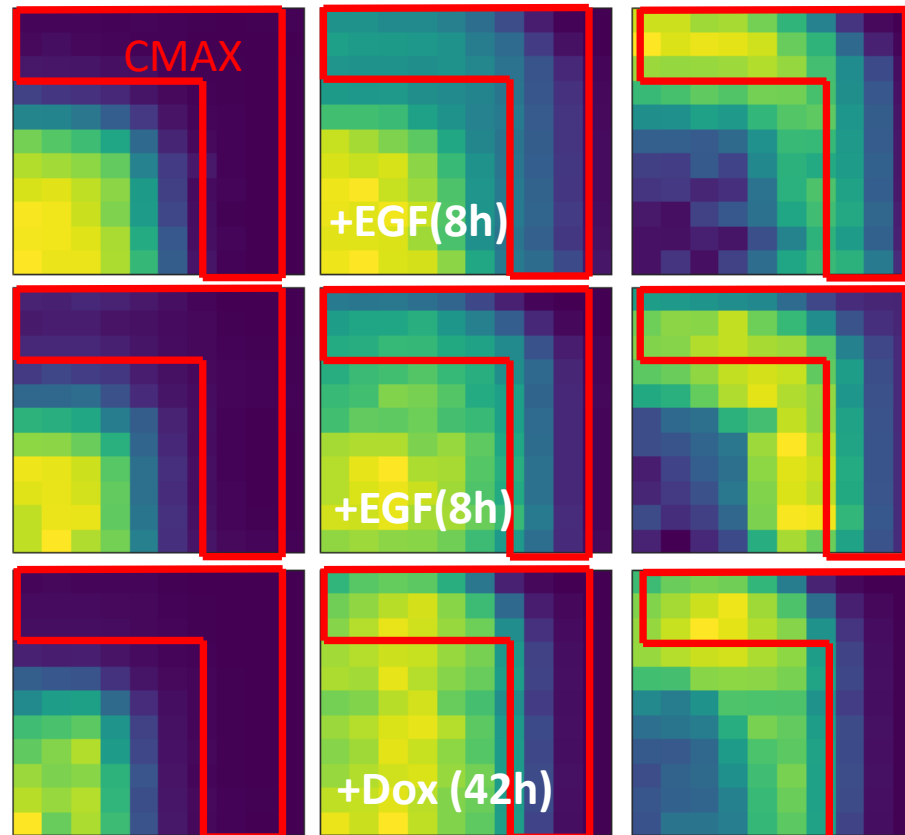


A375 EGFR overexpression (CRISPRa)

HT29 EGFR expressing (colorectal)

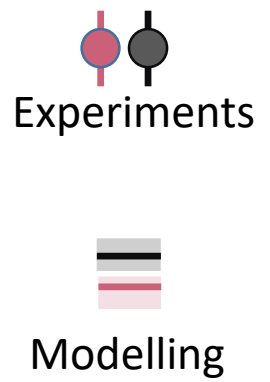
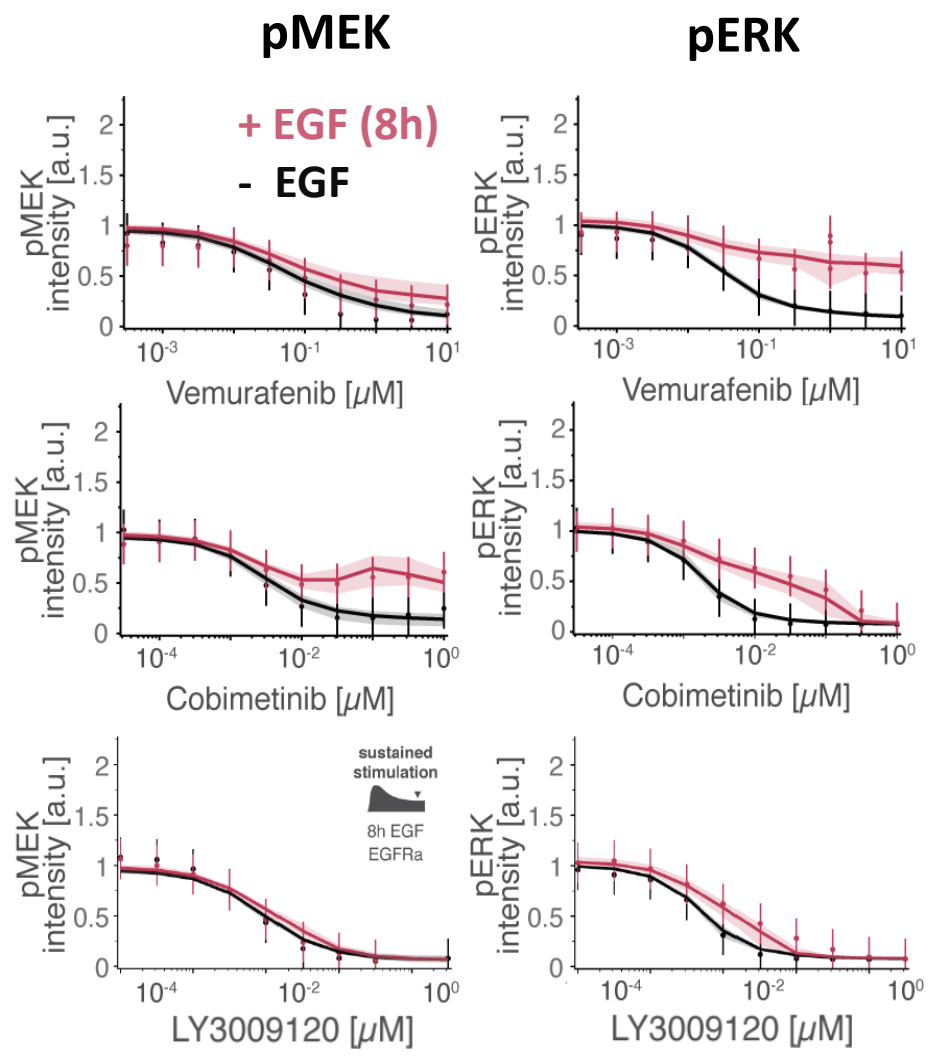
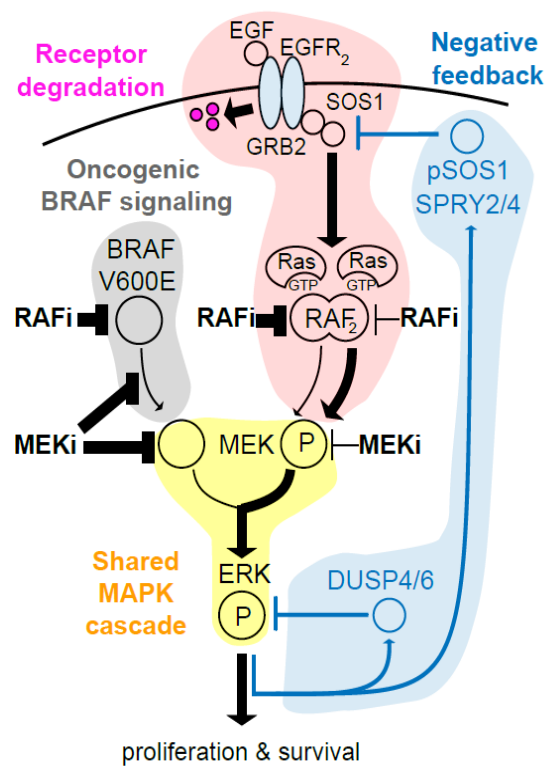
A375 with NRAS Q61K overexpression

Vemurafenib (RAFi)



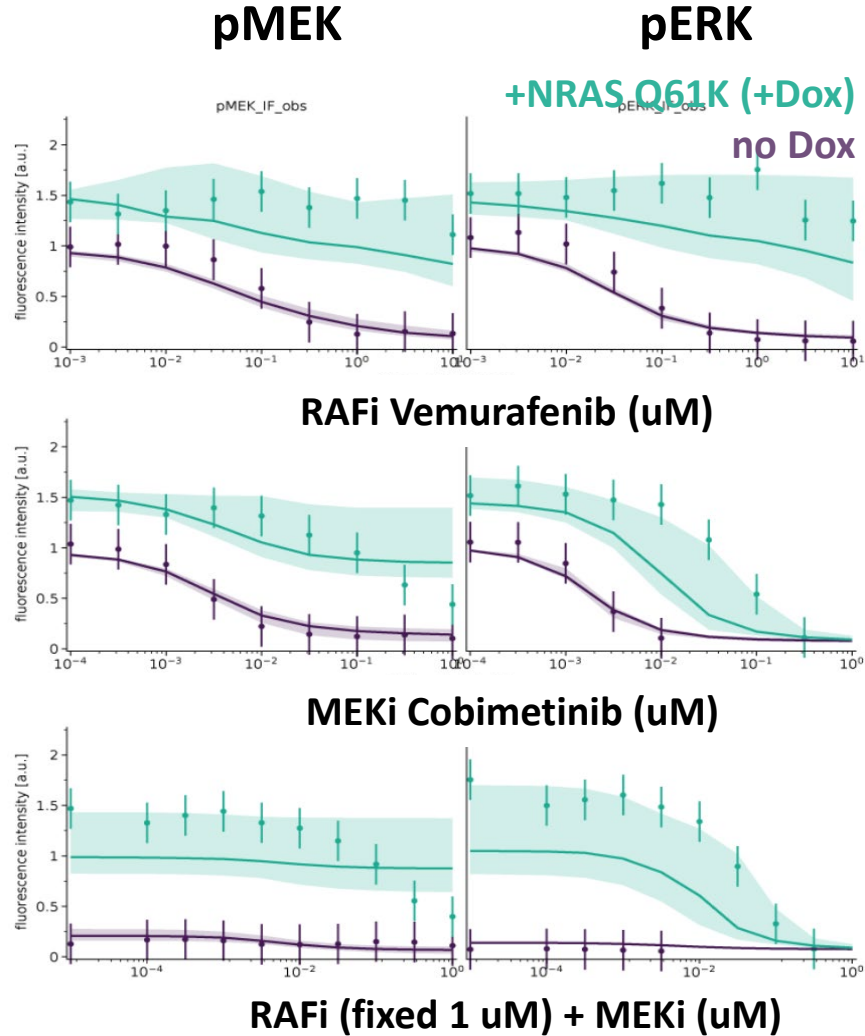
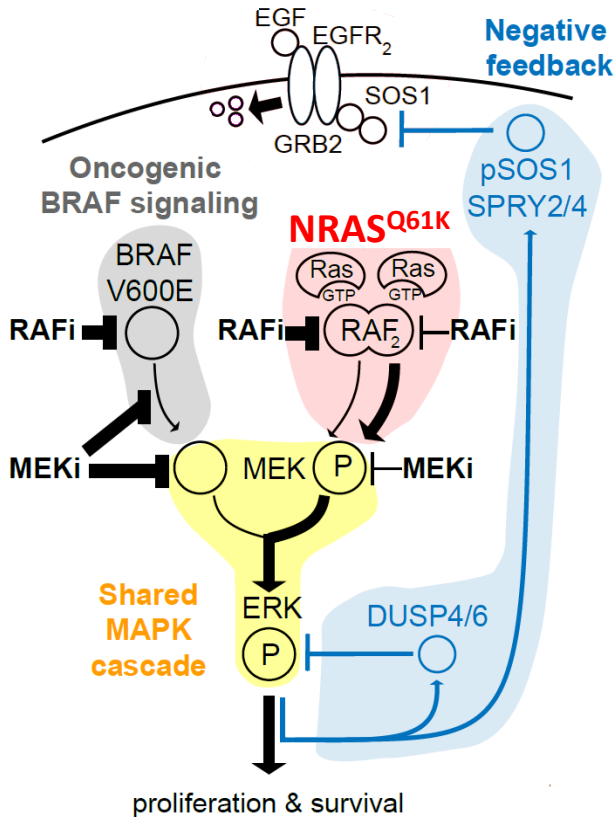
# The model accurately predicts resistance through MAPK reactivation driven by EGFR overexpression

**Simulated and experimental (CRISPRa) overexpression of EGFR by x10 times**



# The model accurately predicts resistance through MAPK reactivation driven by RAS mutations

**Simulated and experimental (Dox inducible) expression of NRAS<sup>Q61K</sup> with x10 lower GTPase activity**

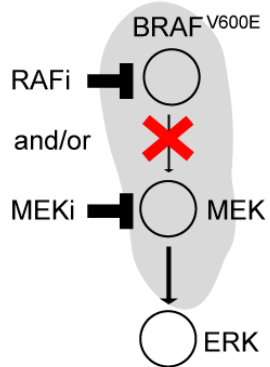


**Experiments**  
**Modelling**

# Conclusions on modeling complex drug-protein interactions in the MAPK signaling of BRAF-mutant cancers

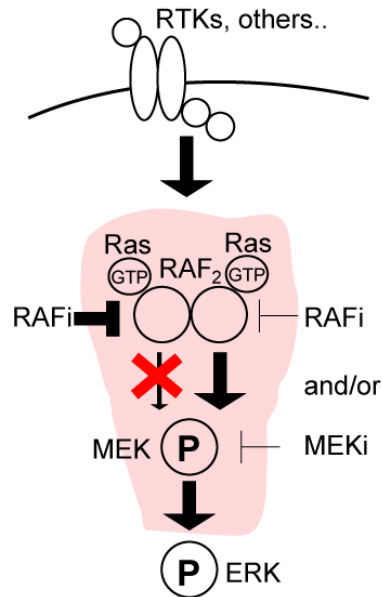
Pharmacological signaling specificity

**Oncogenic BRAF signaling**



**Sensitive to  
RAFi and/or MEKi**

**Ras-GTP signaling**

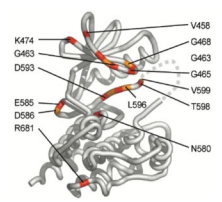


**Resistant to  
RAFi and/or MEKi**

1. MAPK reconfiguration by Ras signaling assemble RAS-RAF-MEK complexes with lower affinity for drugs.
2. Genetic and adaptive resistance to RAF/MEK inhibitors is through MAPK reconfiguration
3. Likely both a source of drug resistance and drug tolerability

# Outlook: Energy-based kinetic modeling is ideal to integrate structural biology information and omics data into predictive models

## Mutational status



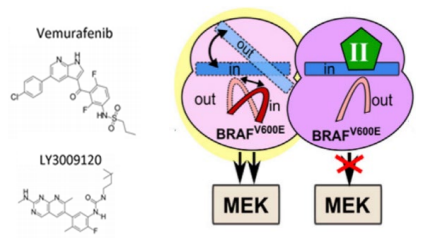
## Energy-based kinetic modeling

```
#Rules
Rule('RR', R(r=None)+R(r=None)<>R(r=1)%R(r=1), RR_phi, Ea0_rr)
Rule('RI', R(i=None)+I(r=None)<>R(i=1)%I(r=1), RI_phi, Ea0_ri)
Rule('RA', R(i=None)+A(r=None)<>R(i=1)%I(r=1), RI_phi, Ea0_ri)

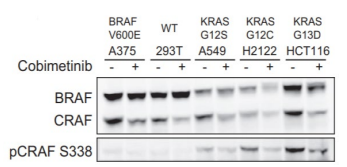
#Energy Patterns
EnergyPattern('ep_RR', R(r=1)%R(r=1), Gf_RR)
EnergyPattern('ep_RI', R(i=1)%I(r=1), Gf_RI)
EnergyPattern('ep_IRR', I(r=1)%R(r=2)%R(r=2,i=None), ep_IRR)
EnergyPattern('ep_IRRI', I(r=1)%R(r=2)%R(r=2,i=3)%I(r=3), ep_IRRI)

#Expressions
Expression('Gf_RR', log(RR_kd))
Expression('Gf_RI', log(RI_kd))
Expression('Ea0_RR', -(RR_phi*log(RR_kd) + log(RR_kf)))
```

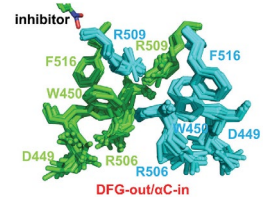
## Drug-protein conformations



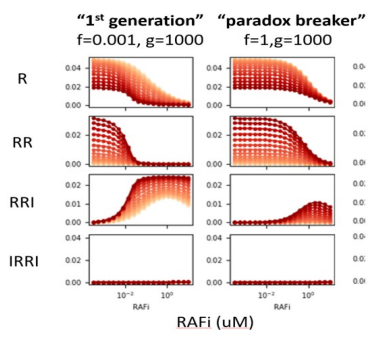
## Protein abundances and PTM



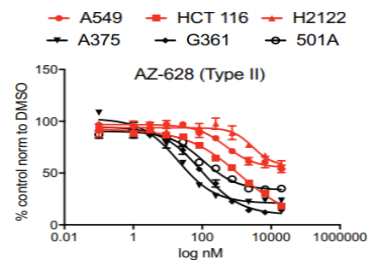
## Molecular dynamic simulations



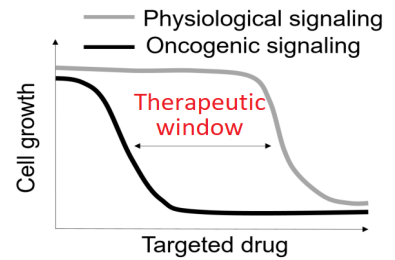
## Drug design



## Drug combinations



## PK/PD analysis



$$\frac{d[pEGFR]}{dt} = k_{on} * [EGFR] * [EGF] - \dots$$

$$\frac{d[Ras-GTP]}{dt} = k_{on} * [Ras] * [GTP] - \dots$$

$$\frac{d[RAF_2]}{dt} = k_{on} * [RAF]^2 - k_{off} * [RAF_2] + \dots$$

$$\frac{d[BRAF^{V600E}]}{dt} = k_{exp} - k_{deg} * [BRAF^{V600E}] + \dots$$

$$\frac{d[pMEK]}{dt} = k_{on} * [MEK] * [BRAF^{V600E}] - \dots$$

$$\frac{d[pERK]}{dt} = k_{on} * [ERK] * [pMEK] - \dots$$

$$\frac{d[CyclinD1]}{dt} = k_{exp} - k_{deg} * [CyclinD1] + \dots$$



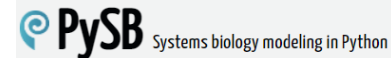
# Resources

- **Code for this webinar:** [https://github.com/lgerosa/rosa\\_webinar\\_20Jan2021](https://github.com/lgerosa/rosa_webinar_20Jan2021)
  - Toy examples of thermodynamic model implementations in PySB
- **MAPK model:** <https://github.com/labsyspharm/marm1-supplement>
  - [\*Receptor-Driven ERK Pulses Reconfigure MAPK Signaling and Enable Persistence of Drug-Adapted BRAF-Mutant Melanoma Cells\*](#)  
Gerosa L, et al. - Cell Systems, 2020
- **PySB:** <http://pysb.org/>
  - [\*Programming biological models in Python using PySB\*](#)  
Lopez CF, Muhlich JL, Bachman JA, Sorger PK - Molecular systems biology, 2013
- **Foundational work:**
  - Energy-BNG by **Faeder lab** @ UPittsburgh:
    - [\*Advances in Rule-based Modeling: Compartments, Energy, and Hybrid Simulation, with Application to Sepsis and Cell Signaling\*](#), Hogg JS , PhD Thesis , U Pitt - 2013
    - [\*Energy-based modeling in BioNetGen\*](#), Sekar JAP, Hogg JS , Faeder JR  
IEEE International Conference on Bioinformatics and Biomedicine - 2016
  - Thermodynamic derivations by **Kholodenko lab** @ UCDublin
    - [\*Drug resistance resulting from kinase dimerization is rationalized by thermodynamic factors describing allosteric inhibitor effects\*](#), Kholodenko BN , Cell reports - 2016
    - [\*Dissecting RAF inhibitor resistance by structure-based modeling reveals ways to overcome oncogenic RAS signaling\*](#), Rukhlenko OS , et al, Cell systems – 2018



**Cell Systems**

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# Acknowledgments

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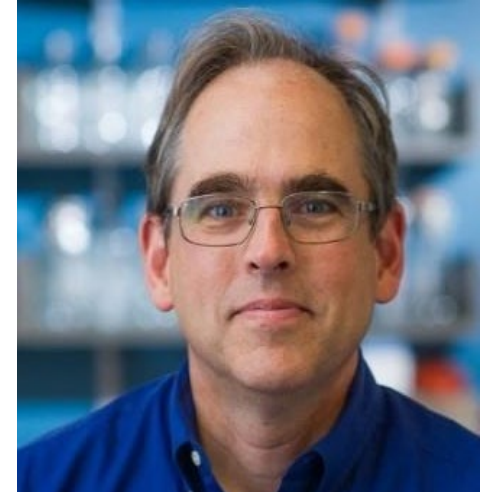
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*Head Bioinf.@LSP*

Implementation  
PysB with energies



**Fabian Fröhlich**  
*Postdoc@LSP*

Model refining &  
parameter estimation



**Peter Sorger**  
*Director@LSP*

Supervision &  
Leadership

## **External experimental collaborators:**

Steve Wiley@PNNL (Proteomics)

Miles Miller@MGH (In-vivo imaging)

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