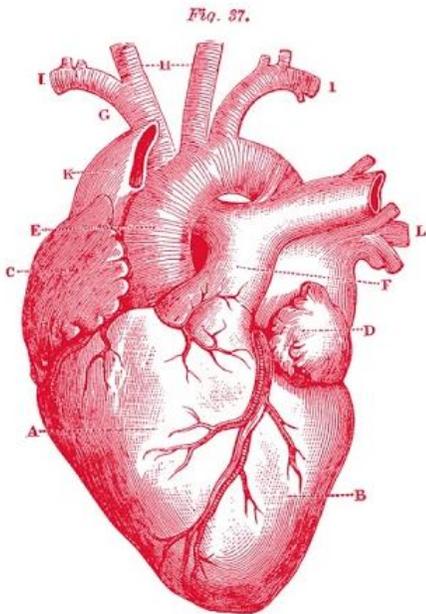


# Cross-species translation of drug-induced electrophysiological response in cardiac myocytes



**Stefano Morotti**

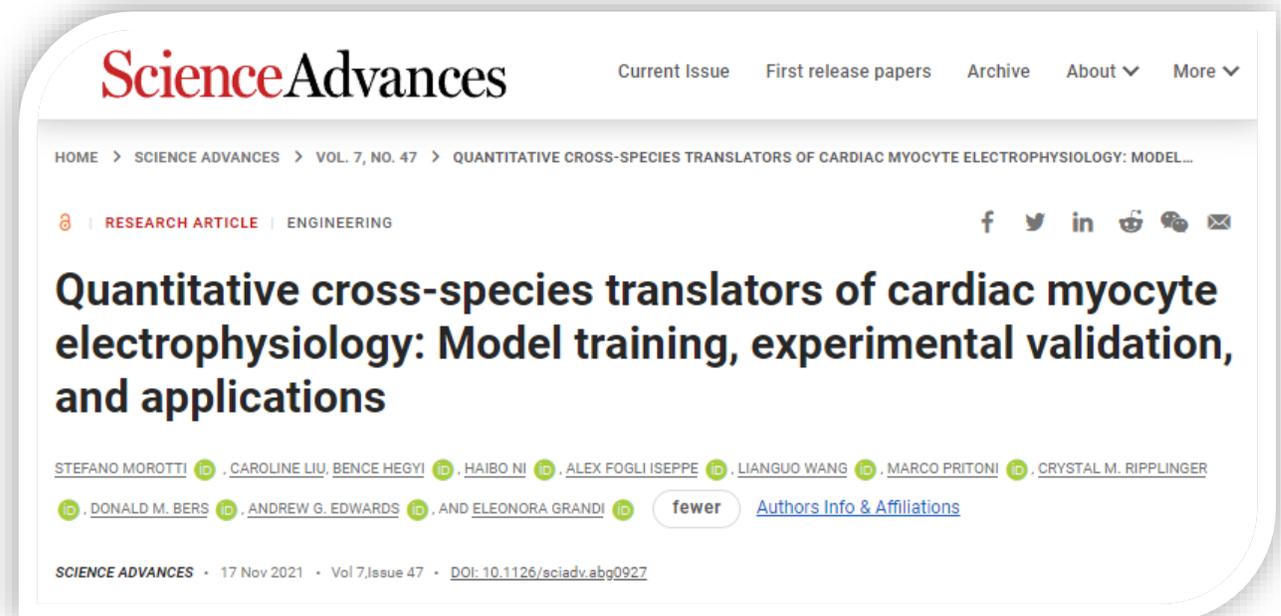
Assistant Professor in Residence  
Department of Pharmacology  
University of California Davis

[smorotti@ucdavis.edu](mailto:smorotti@ucdavis.edu)

 @MorottiLab

# Outline

- Inter-species differences in cardiac electrophysiology
- Impact on drugs' cardiotoxicity screening
- Development of cross-species translators
- Experimental validation
- Future directions



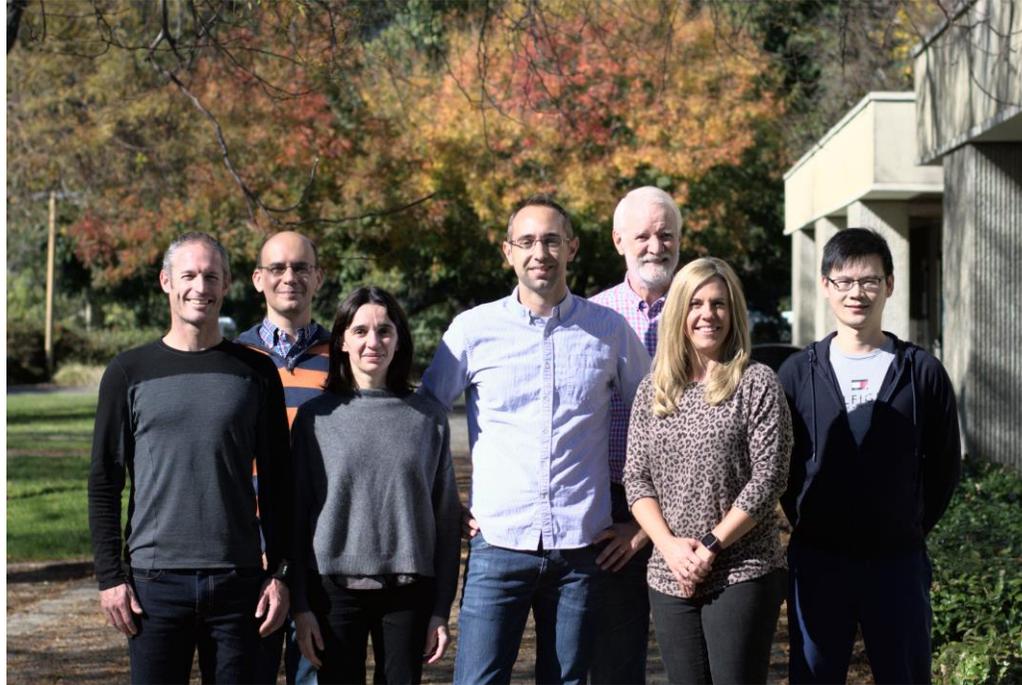
# Acknowledgements

## *Ele Grandi*

Haibo Ni  
Alex Fogli Iseppe  
Xianwei Zhang  
Lin-Lin Liu

## *Andy Edwards*

**UCDAVIS**  
**HEALTH**



## *Crystal Ripplinger*

Lianguo Wang

## *Don Bers*

Bence Hegyi  
Kim Hellgren

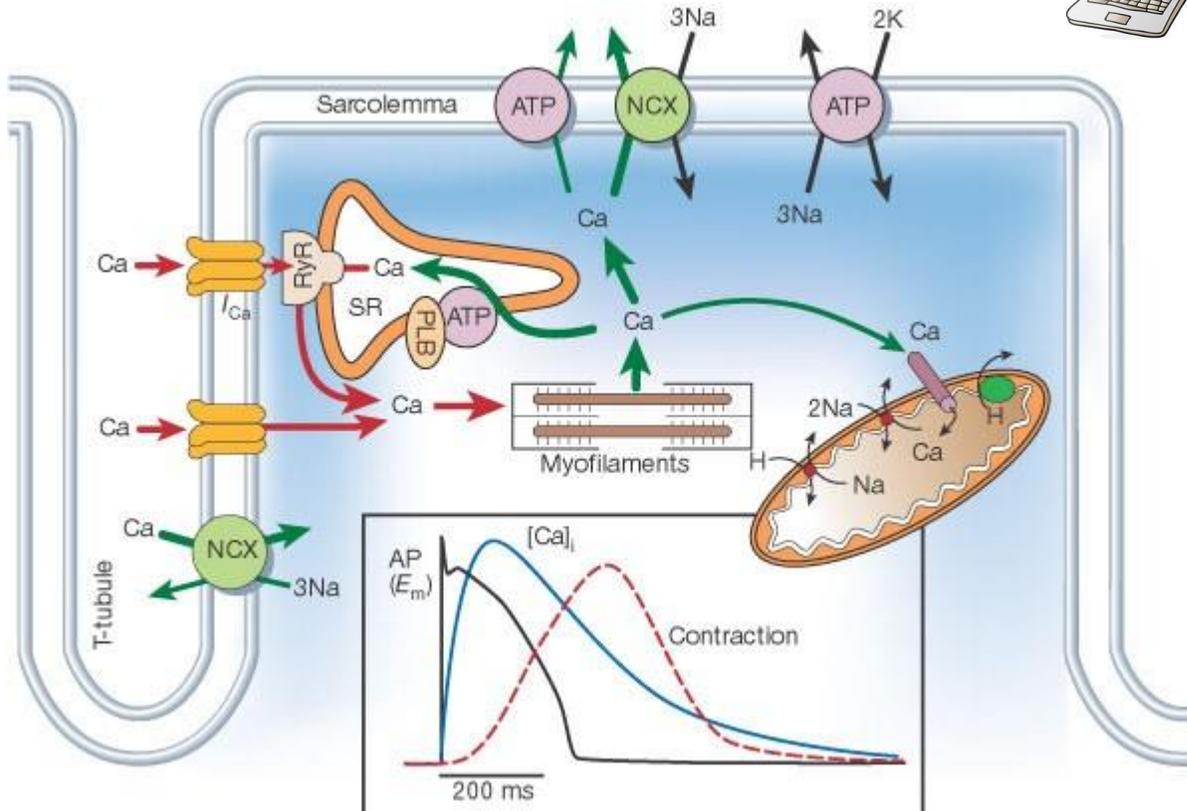


*Marco Pritoni*

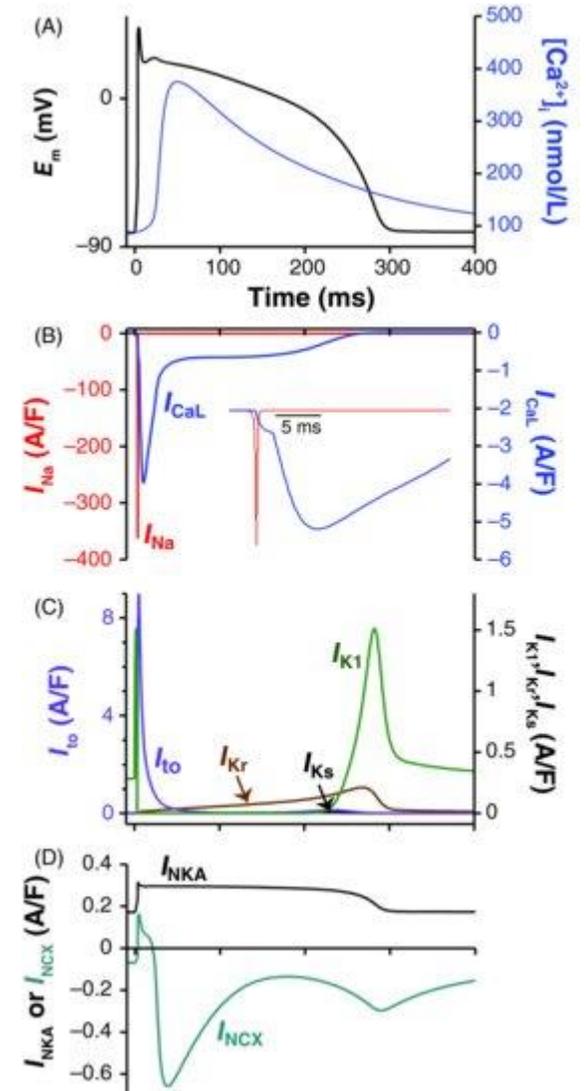


# Cardiomyocyte electrophysiology

## Excitation-contraction coupling

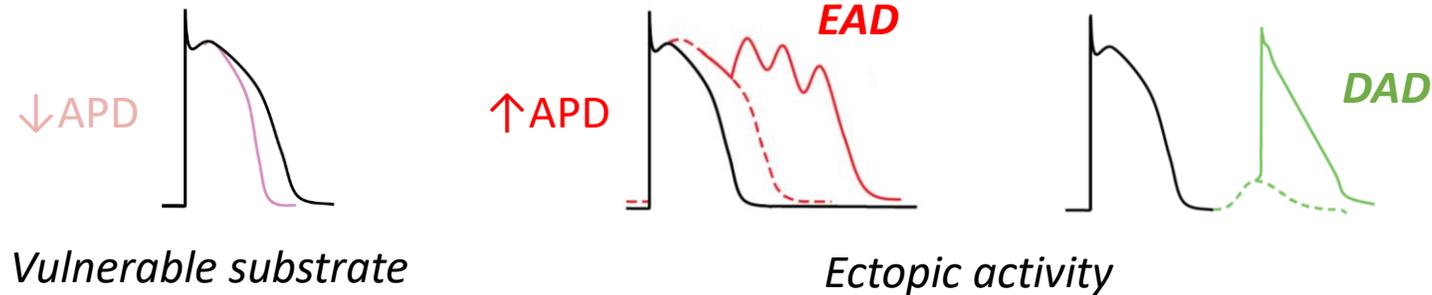


Action Potential (AP) – Ca Transient (CaT) – Myofilament Contraction

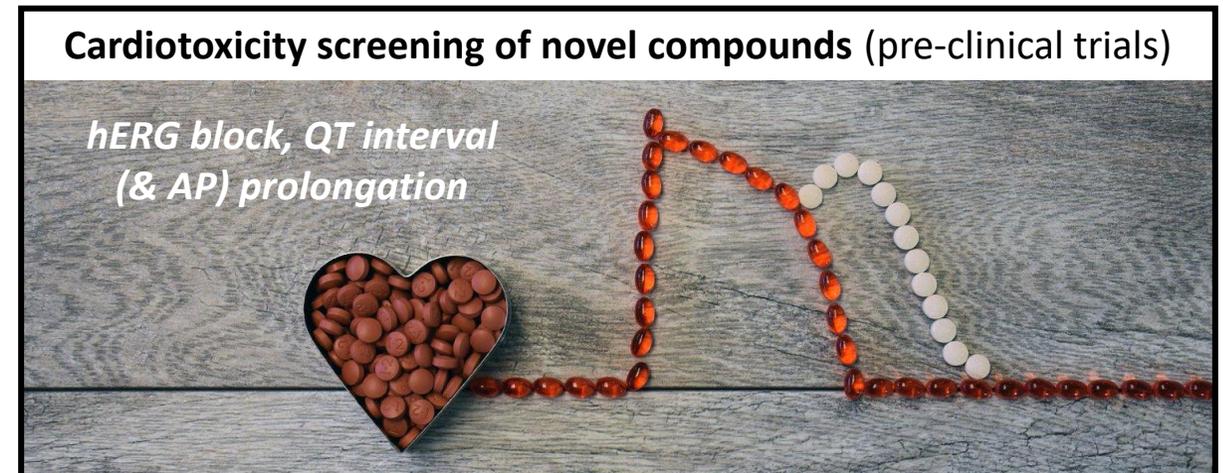


# AP regulation & arrhythmias

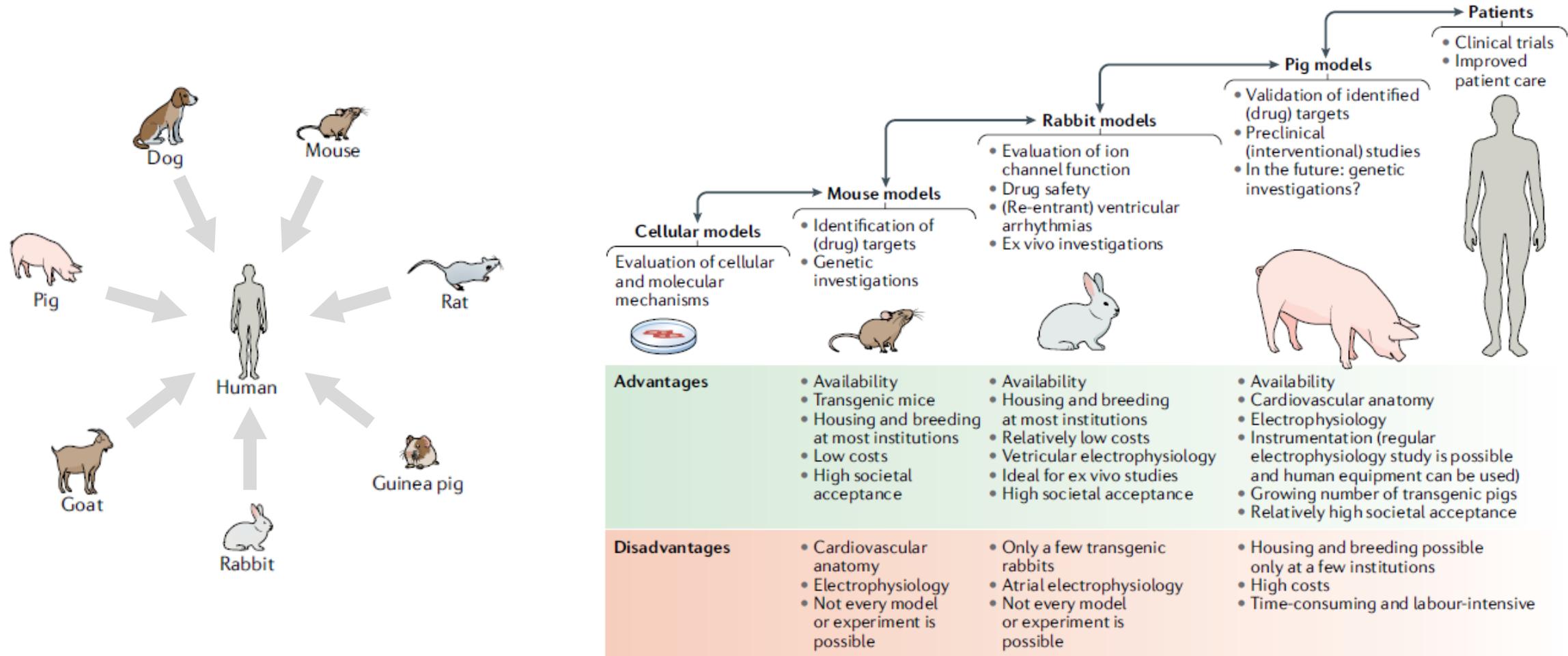
- Impaired AP regulation facilitates both development & maintenance of arrhythmias



- **Inherited conditions**  
(long QT, Brugada, etc.)
- **Acquired conditions**  
(heart failure, atrial fibrillation, etc.)
- **Drug-induced**  
(Torsade de Pointes, brady-arrhythmias, etc.)



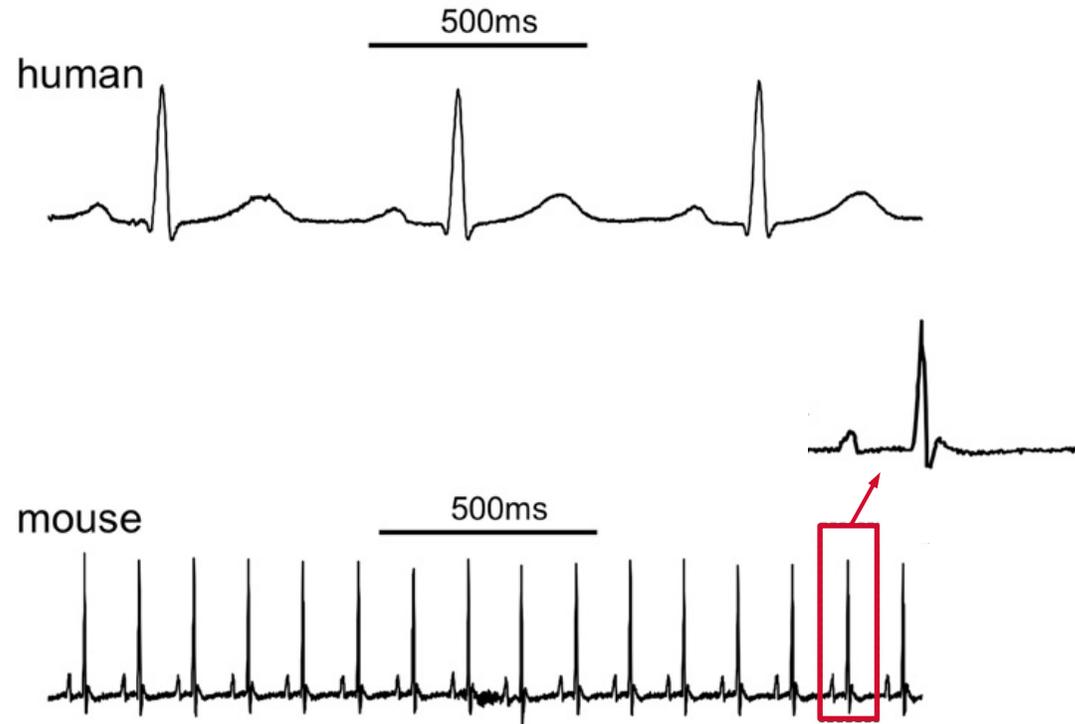
# Animal models in arrhythmia research



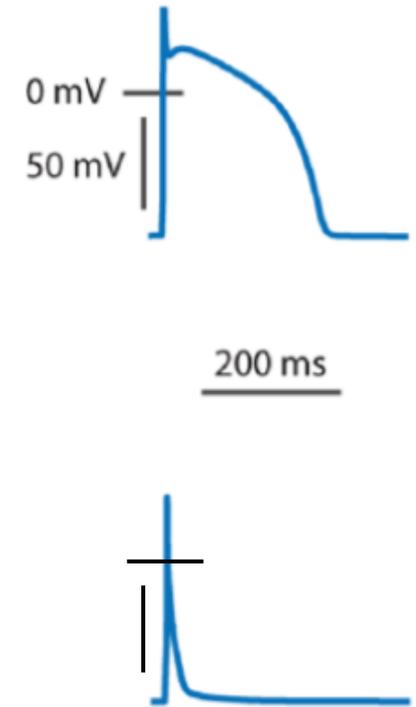
# Inter-species differences in cardiac electrophysiology



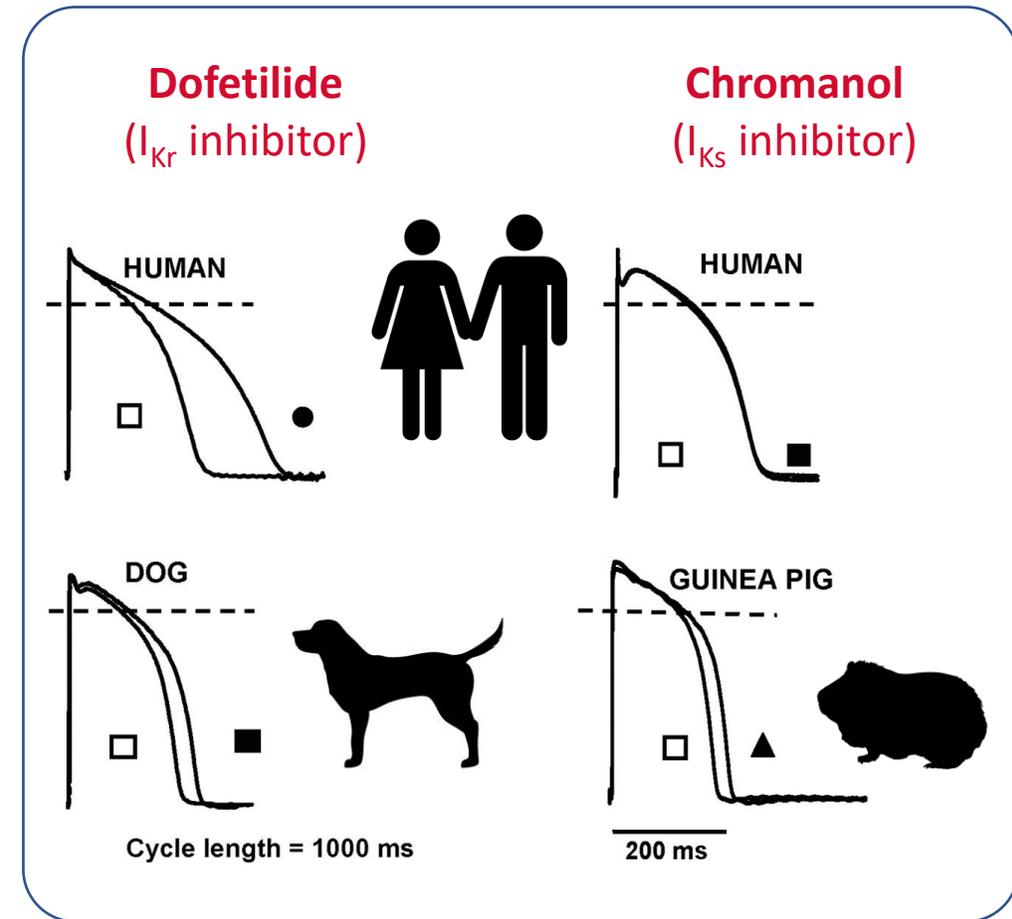
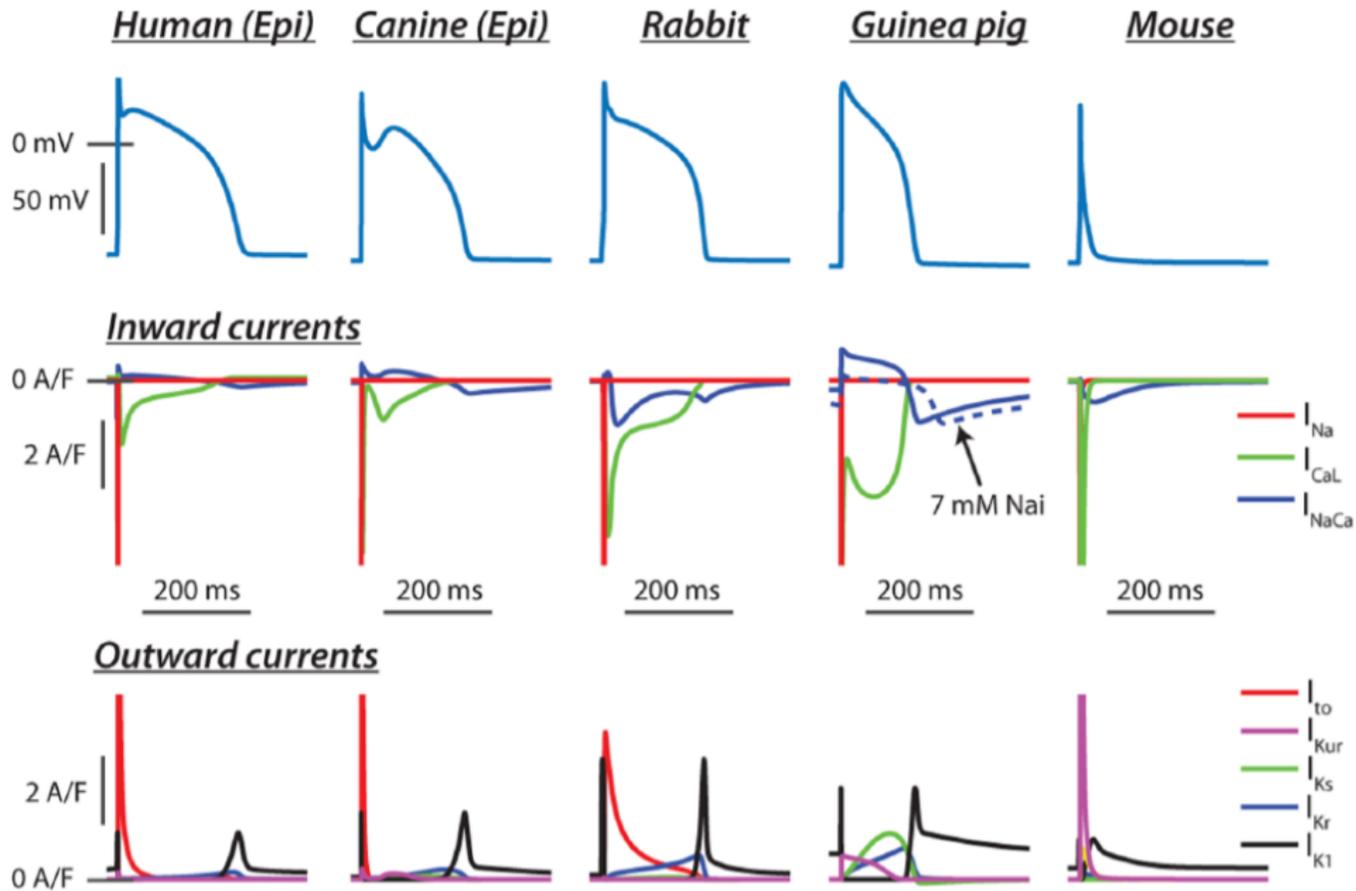
**ECG**



**AP**



# Inter-species differences in ventricular electrophysiology



# Impact on drugs' cardiotoxicity screening

JACC: BASIC TO TRANSLATIONAL SCIENCE

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VOL. 4, NO. 7, 2019

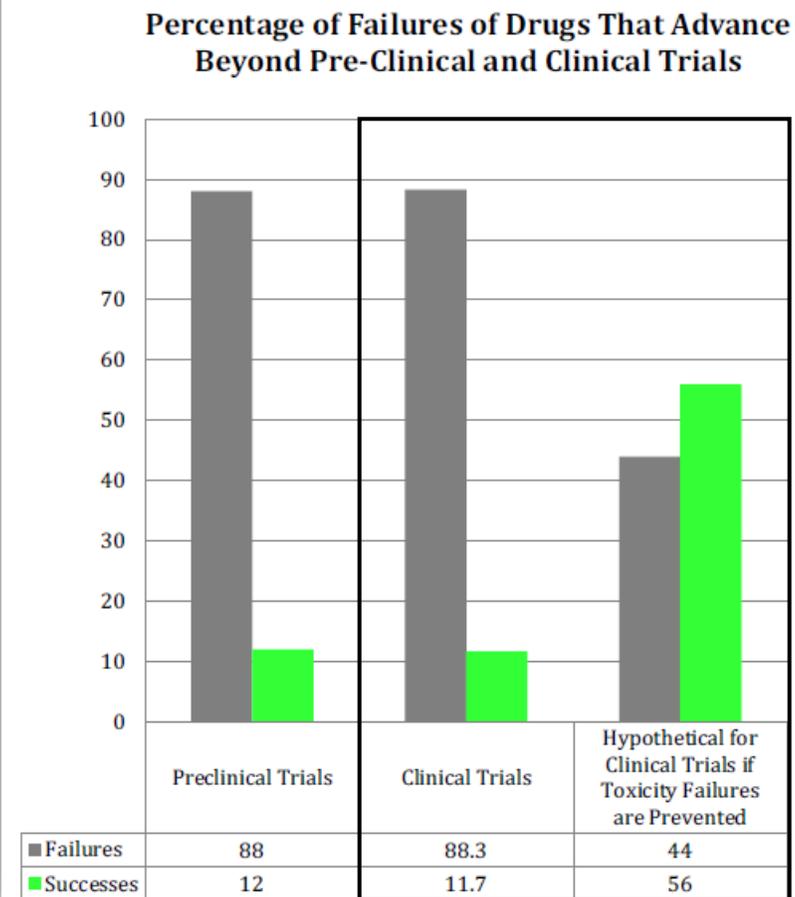
## TRANSLATIONAL PERSPECTIVE

### Limitations of Animal Studies for Predicting Toxicity in Clinical Trials

Is it Time to Rethink Our Current Approach?

Gail A. Van Norman, MD

FIGURE 1 Failures in Translational Research: Preclinical and Clinical Trials



# Impact on drugs' cardiotoxicity screening

JACC: BASIC TO TRANSLATIONAL SCIENCE  
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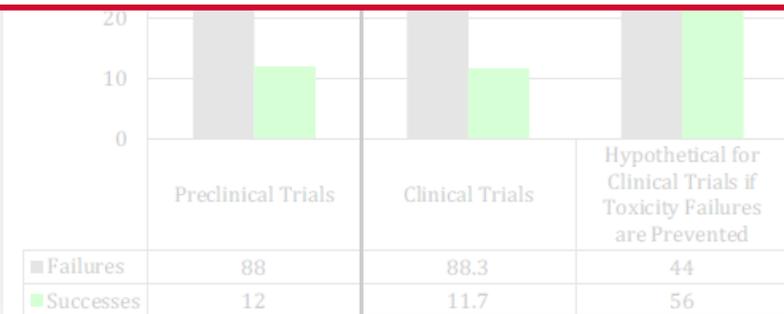
VOL. 4, NO. 7, 2019

## TRANSLATIONAL PERSPECTIVE

FIGURE 1 Failures in Translational Research: Preclinical and Clinical Trials

Percentage of Failures of Drugs That Advance  
Beyond Pre-Clinical and Clinical Trials

***We must systematically characterize  
species-differences in the regulation of  
cardiomyocyte electrophysiology***



# Species-differences in response to $\beta$ -adrenergic stimulation

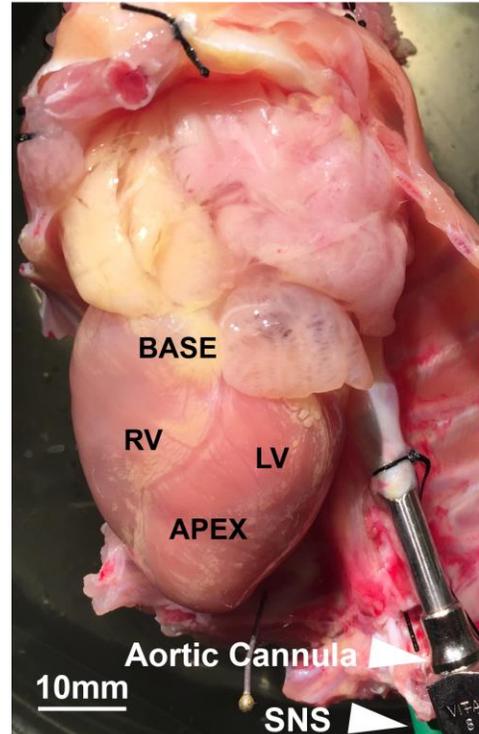


## Sympathetic nerve stimulation (SNS) in whole-heart preparations

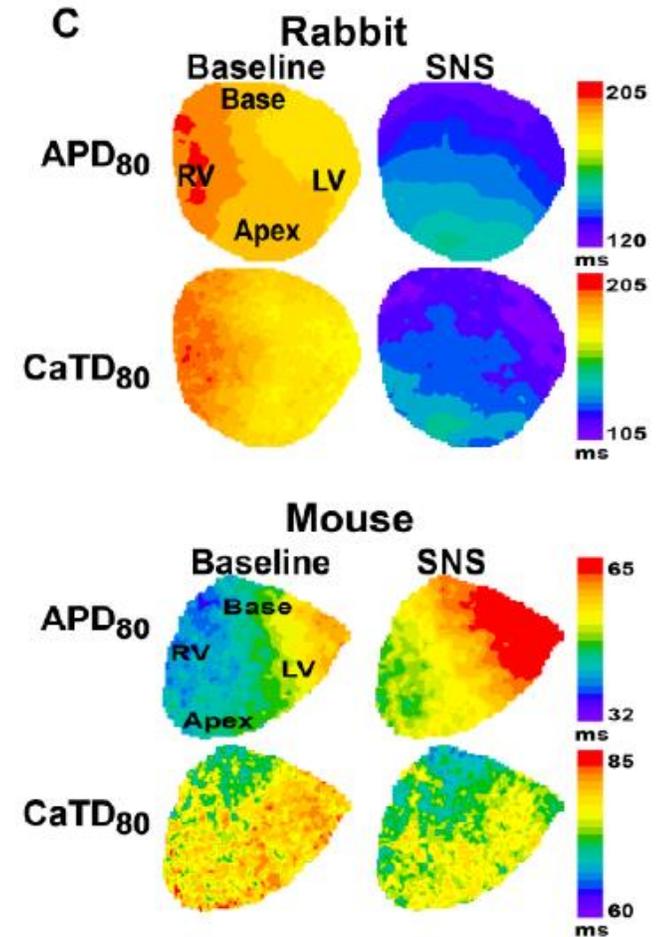
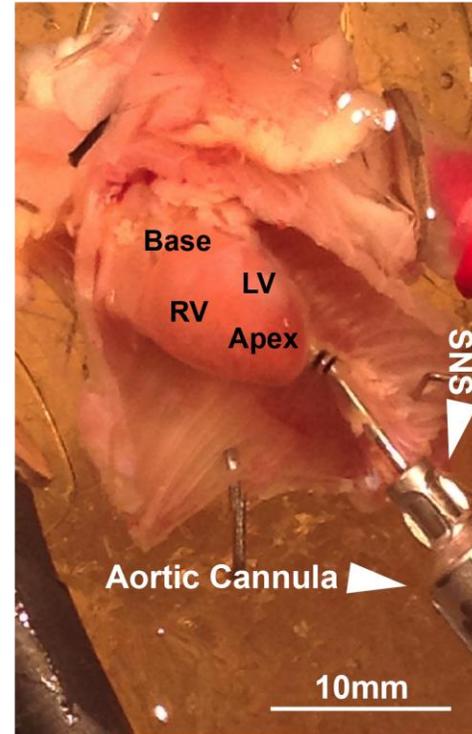
i)  $\beta$ -AR signaling mediates the well-known *fight-or-flight* response, a conserved mammalian behavior

ii)  $\beta$ -AR stimulation is associated with increased propensity for cardiac arrhythmias

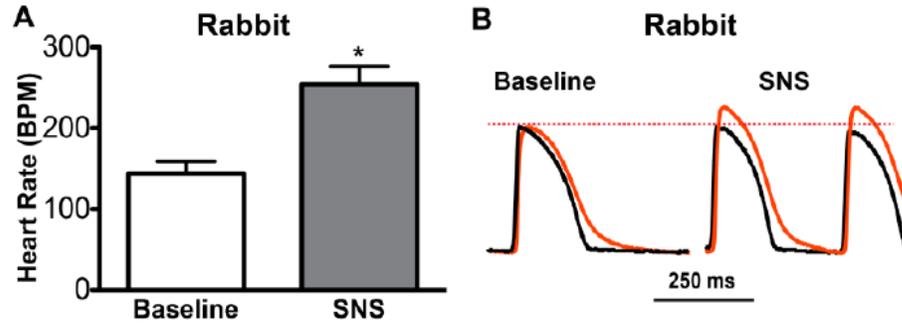
**A** Innervated Rabbit Heart



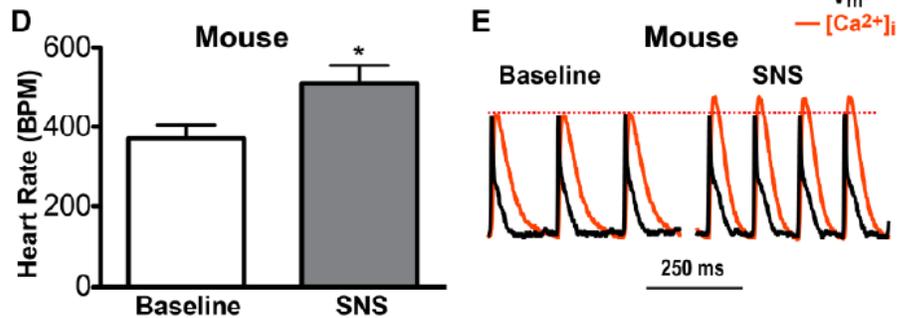
**B** Innervated Mouse Heart



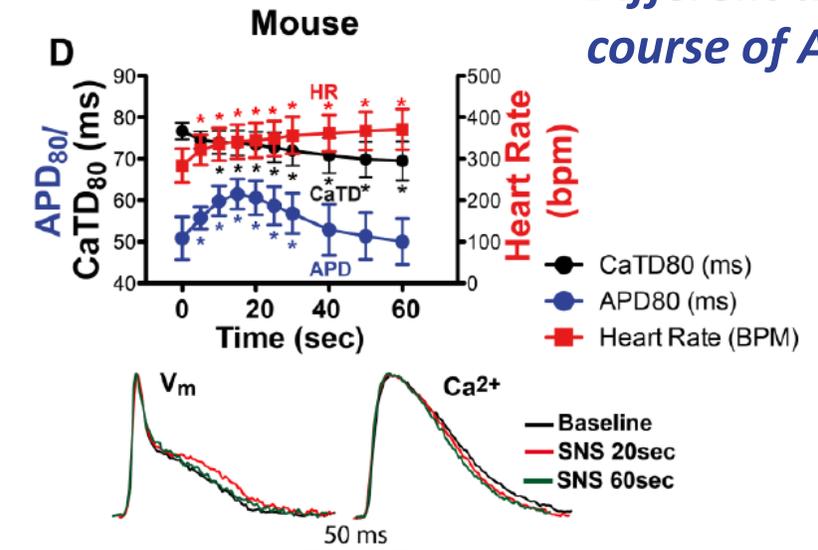
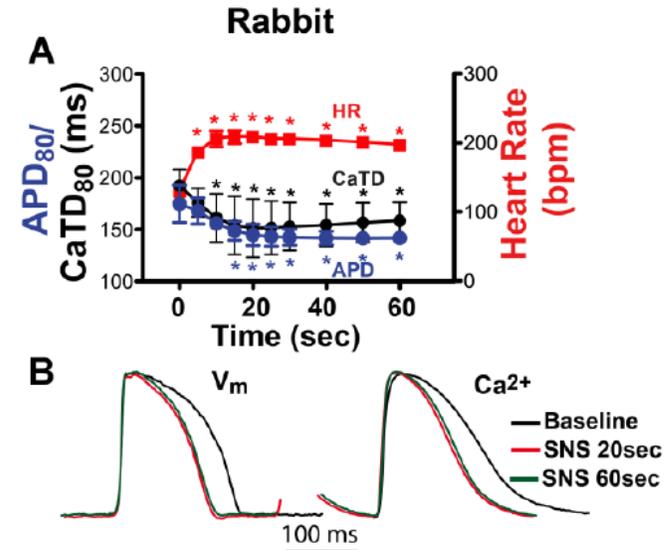
# Consequences of SNS in rabbit vs. mouse



**Conserved chronotropic  
& inotropic response**



**Different time  
course of APD**

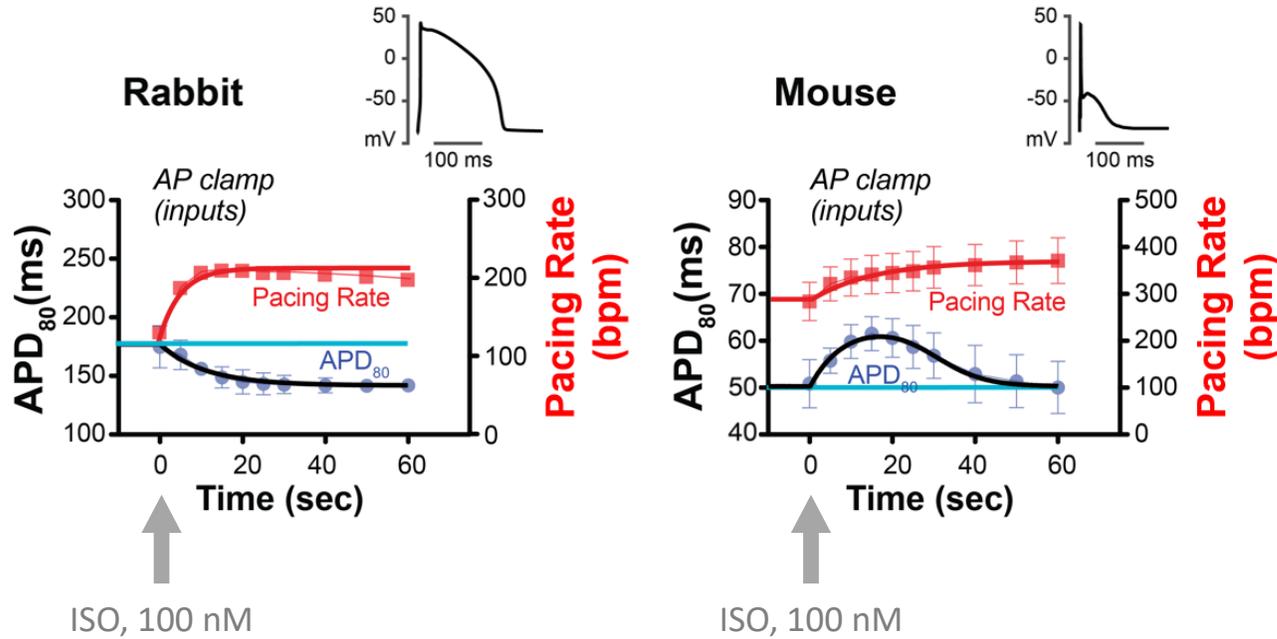


# AP-clamp simulations reveal inter-species differences

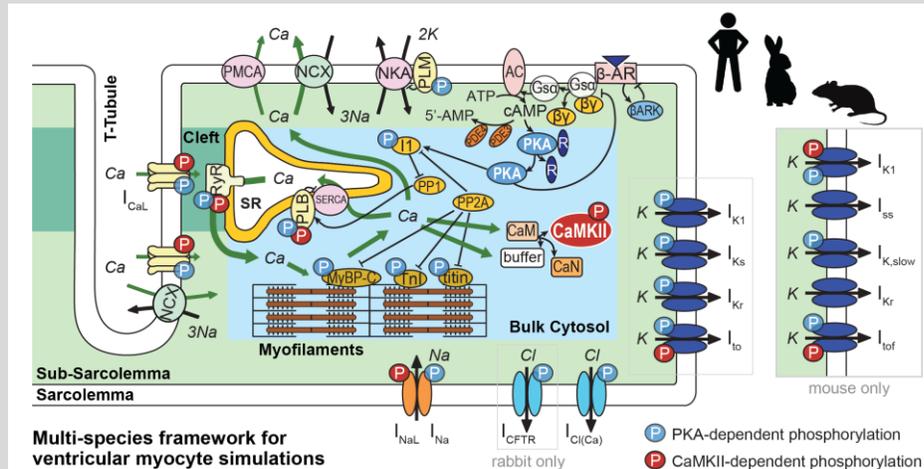


Input:  
time-dependent modulation of pacing rate and APD + ISO administration

Output:  
time course of CaT amplitude (*inotropy*) and time constant of CaT decay (*lusitropy*)



**Deterministic ODE models of AP, Ca transient, & myofilament contraction in ventricular myocytes**



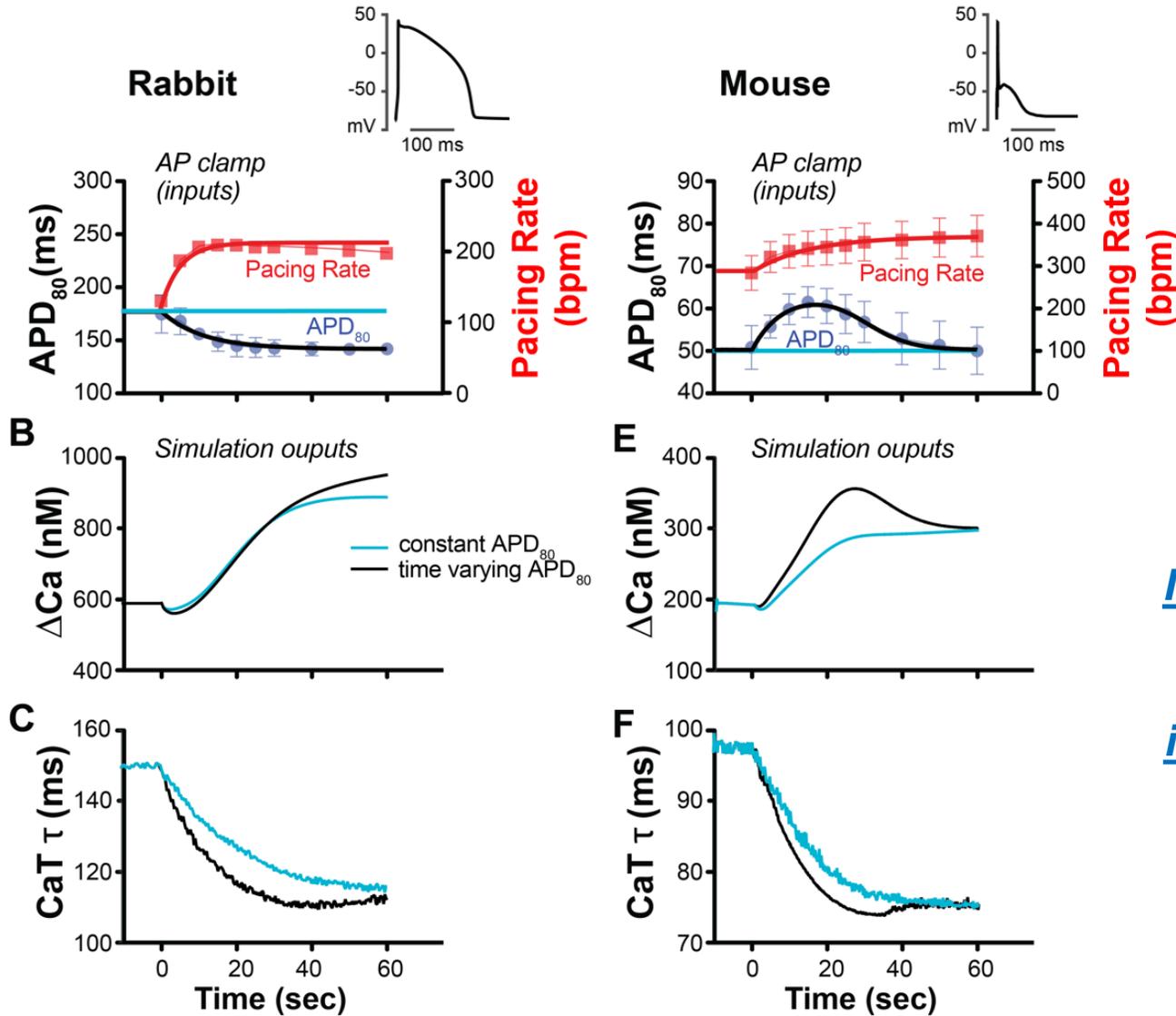
Based on Grandi et al., 2010, Moreno et al., 2013 (**Human**); Shannon et al., 2004, Soltis & Saucerman, 2012, Negroni et al., 2015, Bartos et al., 2017 (**Rabbit**); Morotti et al., 2014, Surdo et al., 2017 (**Mouse**)

# AP-clamp simulations reveal inter-species differences



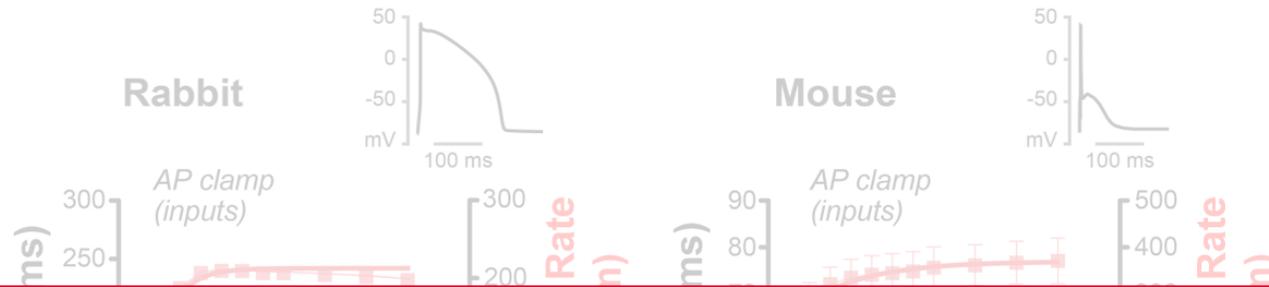
Input:  
time-dependent modulation of pacing rate and APD + ISO administration

Output:  
time course of CaT amplitude (*inotropy*) and time constant of CaT decay (*lusitropy*)



Inter-species differences in AP repolarization lead to optimal enhancement of inotropy & lusitropy during fight-or-flight response

# AP-clamp simulations reveal inter-species differences



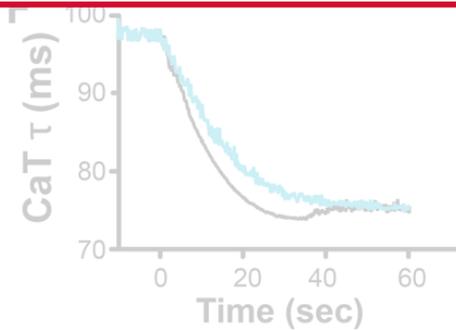
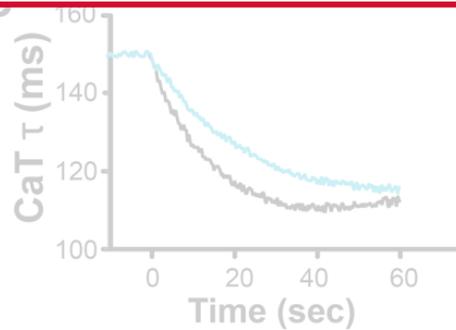
***How can we use our computational models to improve the prediction of human physiology from experiments in animals?***

Input:  
time-dependent  
modulation of  
pacing rate and A  
+ ISO administration

Output:  
time course of C  
amplitude (inotrop  
and time constant  
CaT decay (lusitrop

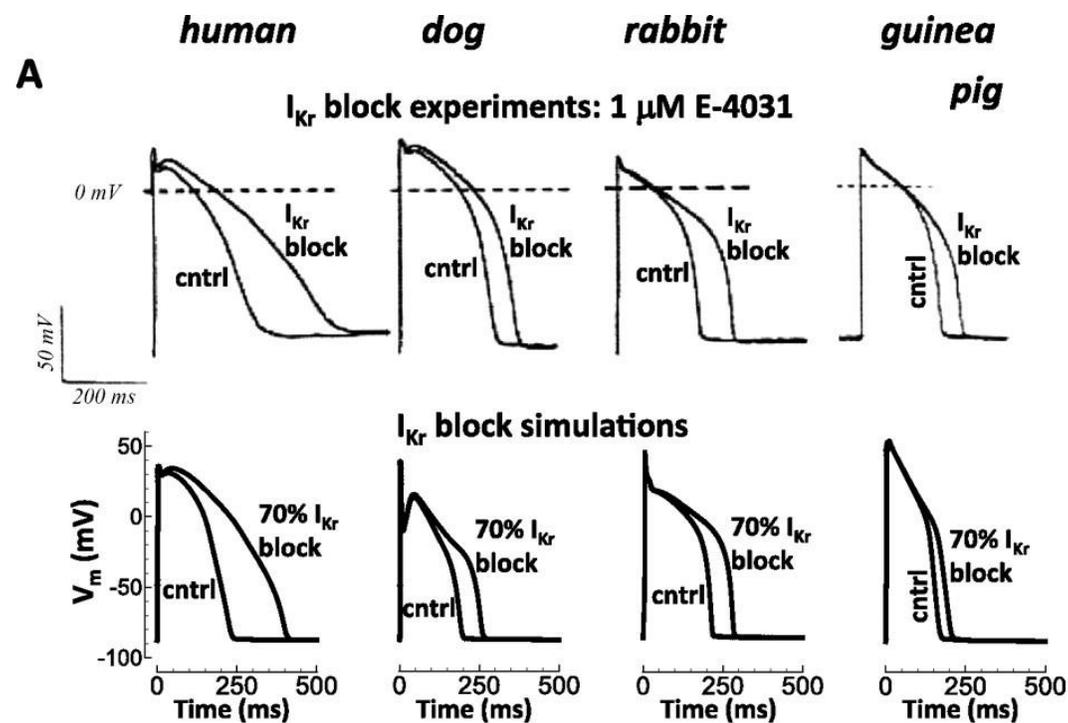
es in  
d to  
t of

inotropy & lusitropy during  
fight-or-flight response



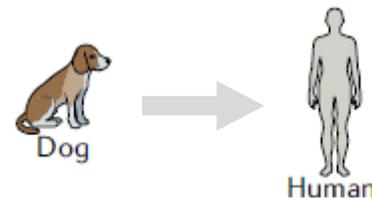
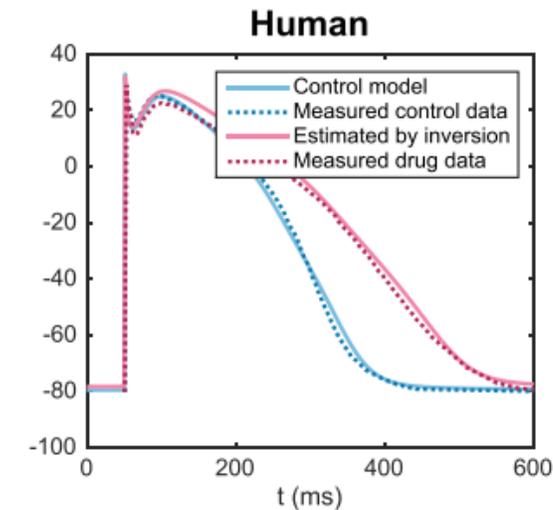
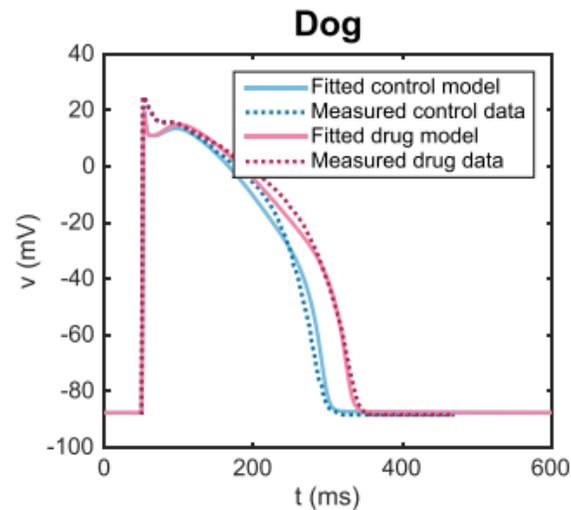
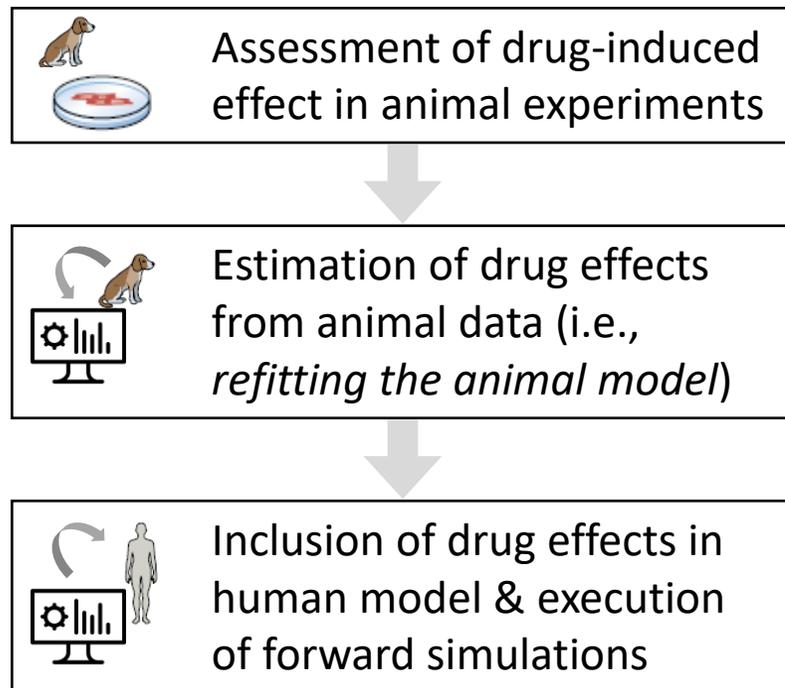
# Existing computational approaches

- Comparing simulations performed in different species



# Existing computational approaches

- Comparing simulations performed in different species
- Cross-species translation based on estimation of drug effects from animal experiments, and execution of new simulations with human model



**SCIENTIFIC  
REPORTS**  
nature research



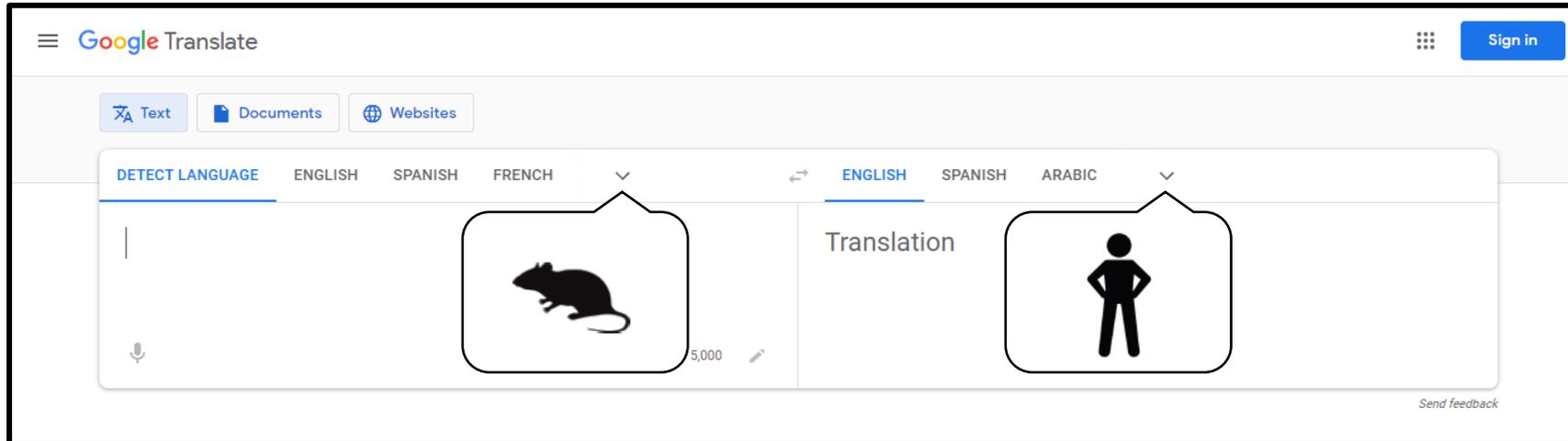
**Computational translation of drug effects from animal experiments to human ventricular myocytes**

Aslak Tveito<sup>1</sup>, Karoline Horgmo Jæger<sup>1</sup>, Mary M. Maleckar<sup>1</sup>, Wayne R. Giles<sup>2</sup> & Samuel Wall<sup>1</sup>

# Goal: to develop an *immediate* cross-species translator



Assessment of drug-induced effect in animal experiments



Quantitative prediction of drug-induced effect in human

# A previously developed *immediate* translator

ARTICLE OPEN

Population-based mechanistic modeling allows for quantitative predictions of drug responses across cell types

Jingqi Q. X. Gong<sup>1</sup> and Eric A. Sobie<sup>1</sup>

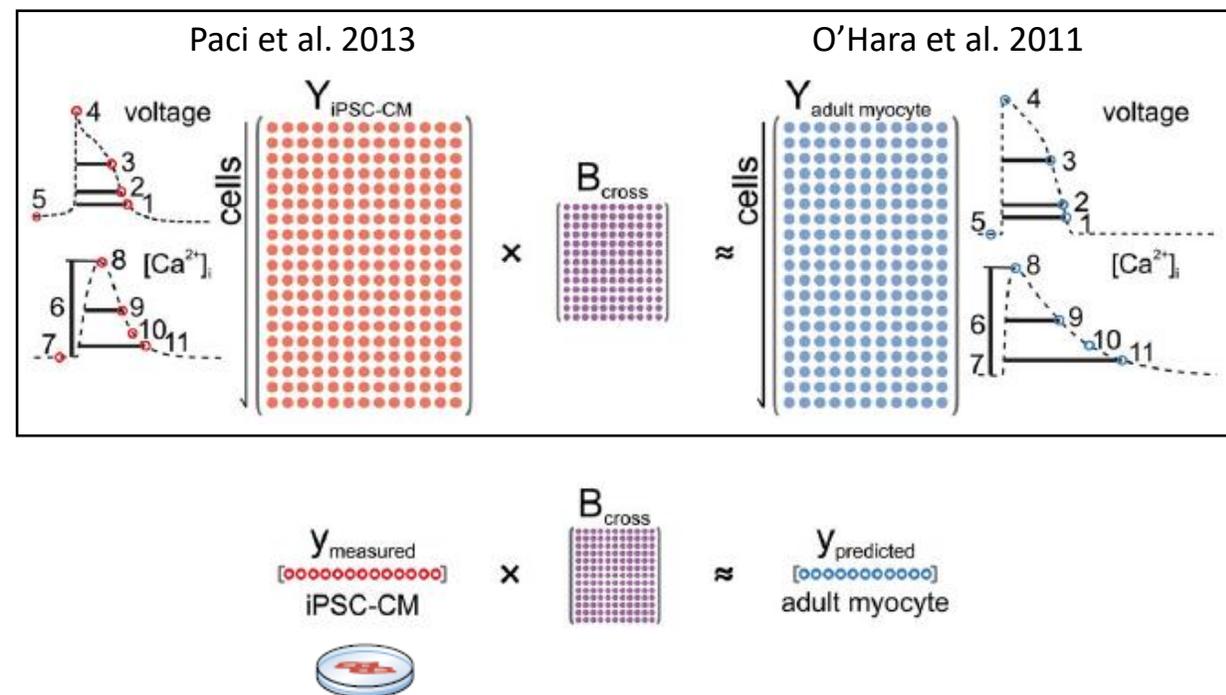
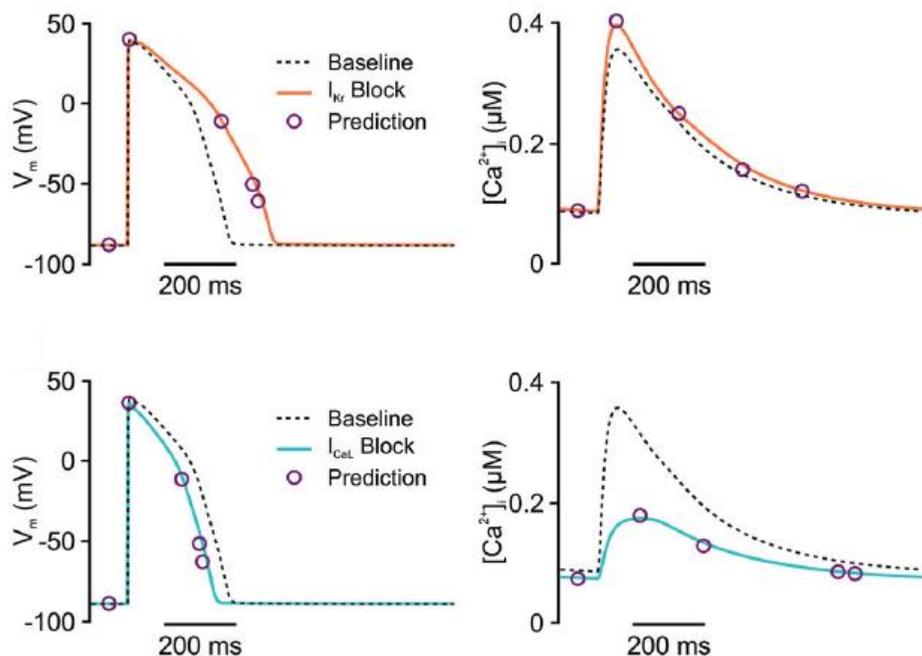
<sup>1</sup>Department of Pharmacological Sciences, Icahn School of Medicine at Mount Sinai, New York, NY, USA  
Correspondence: Eric A. Sobie (eric.sobie@mssm.edu)

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and Applications

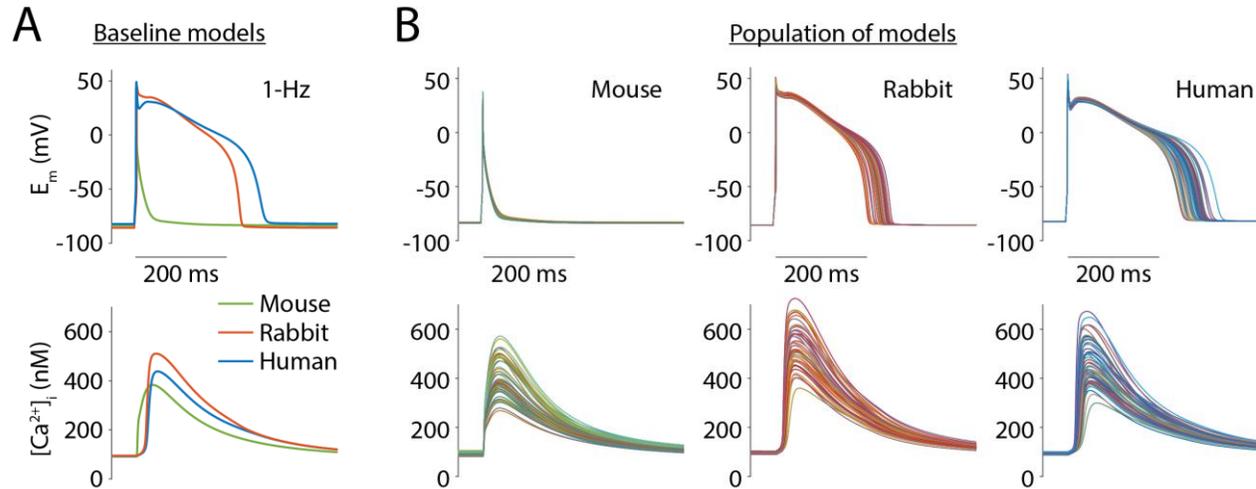
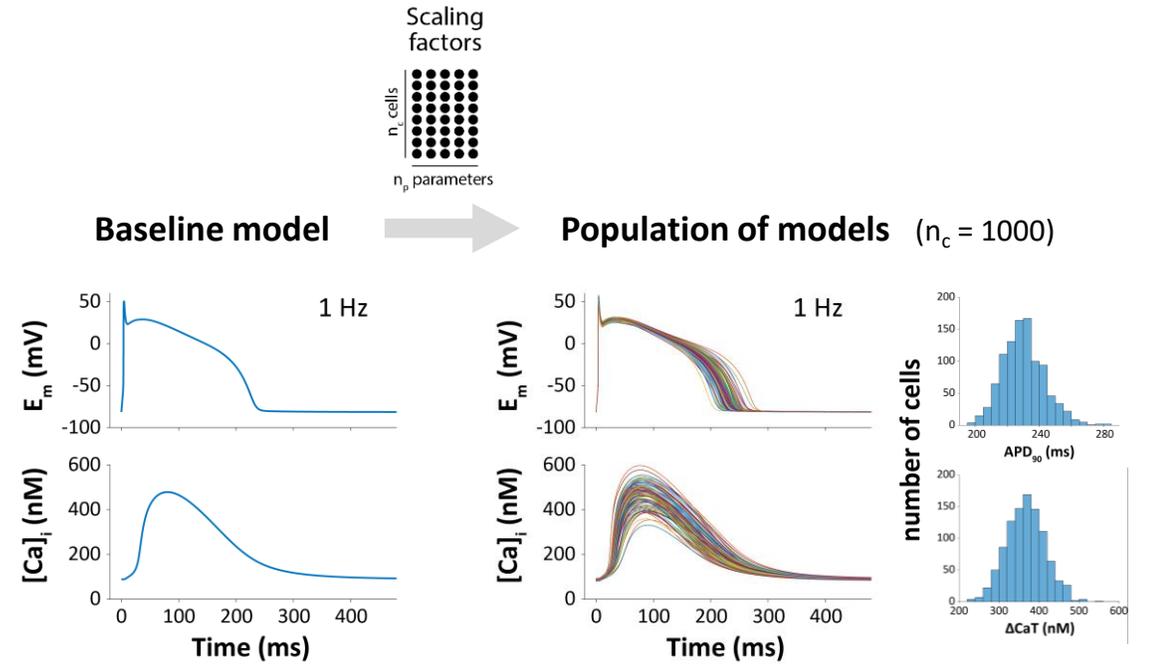
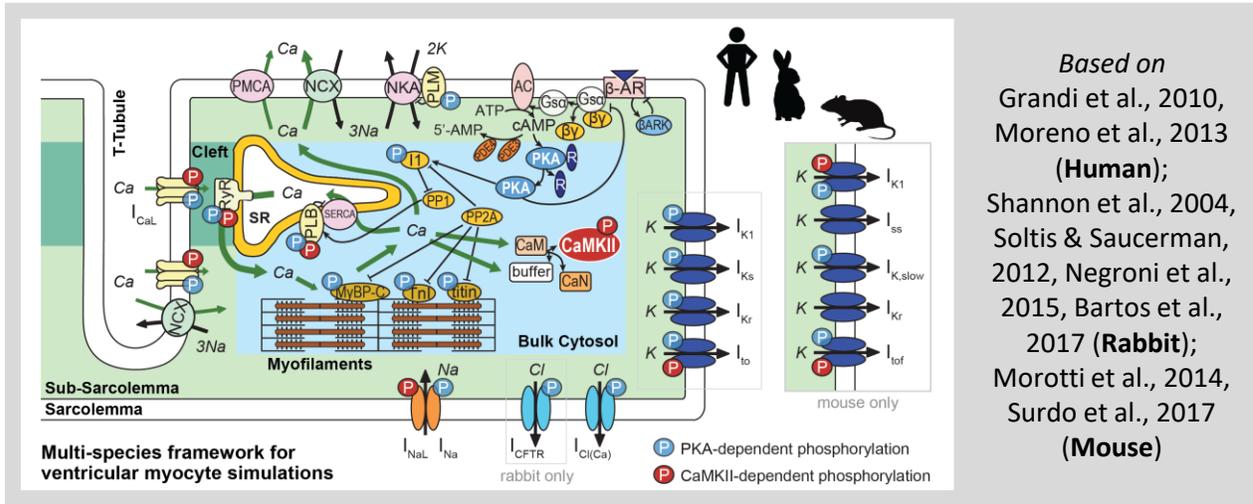
Response in  
hiPSC-CM



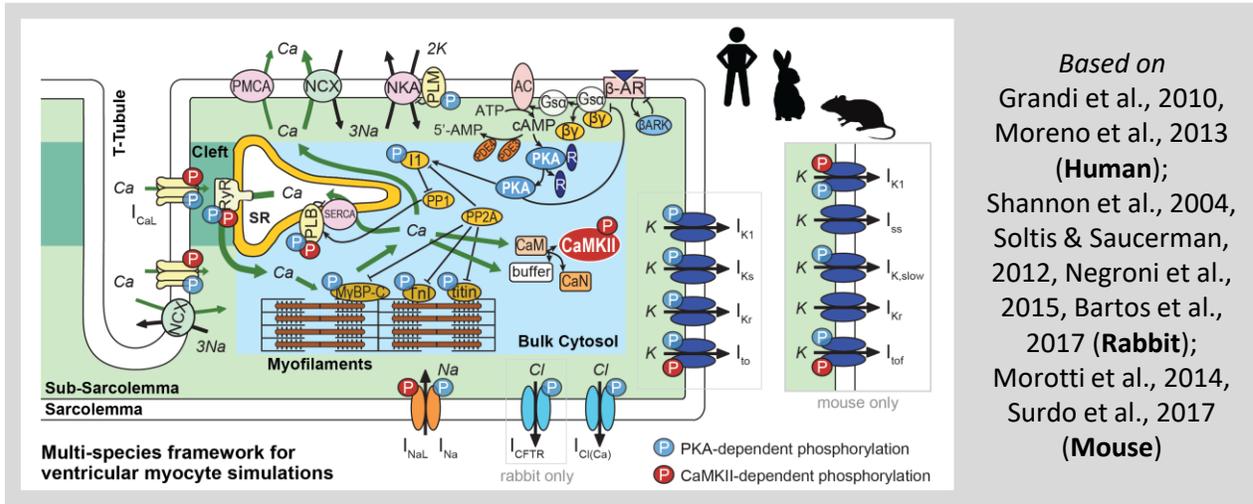
Response in  
adult HVM



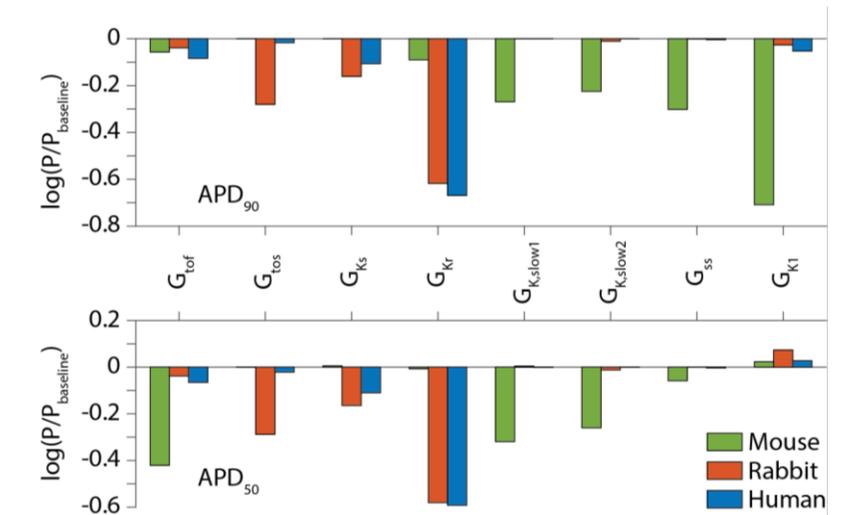
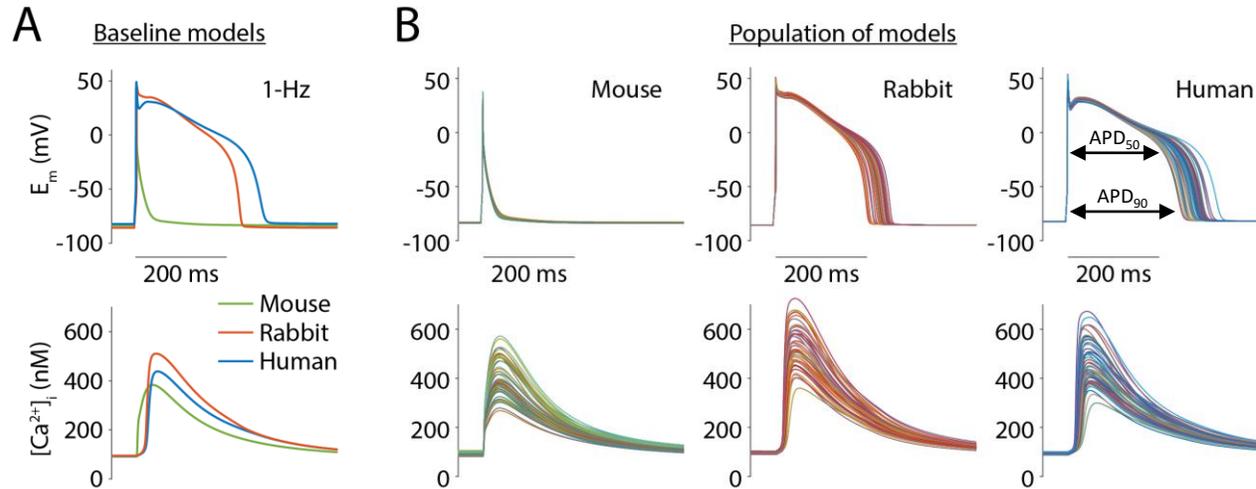
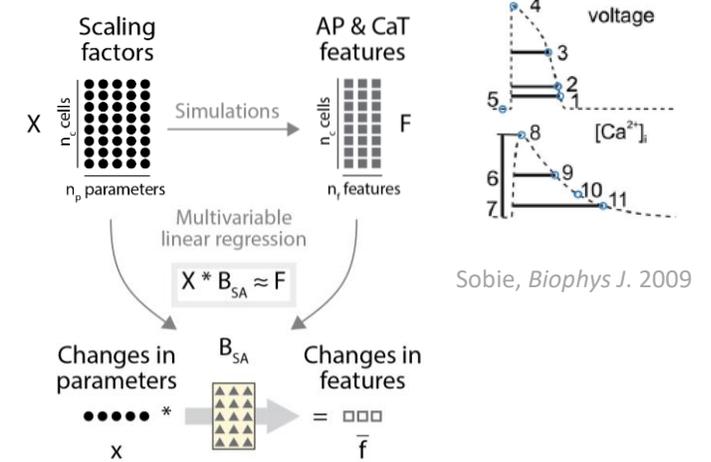
# Methods: building populations of models



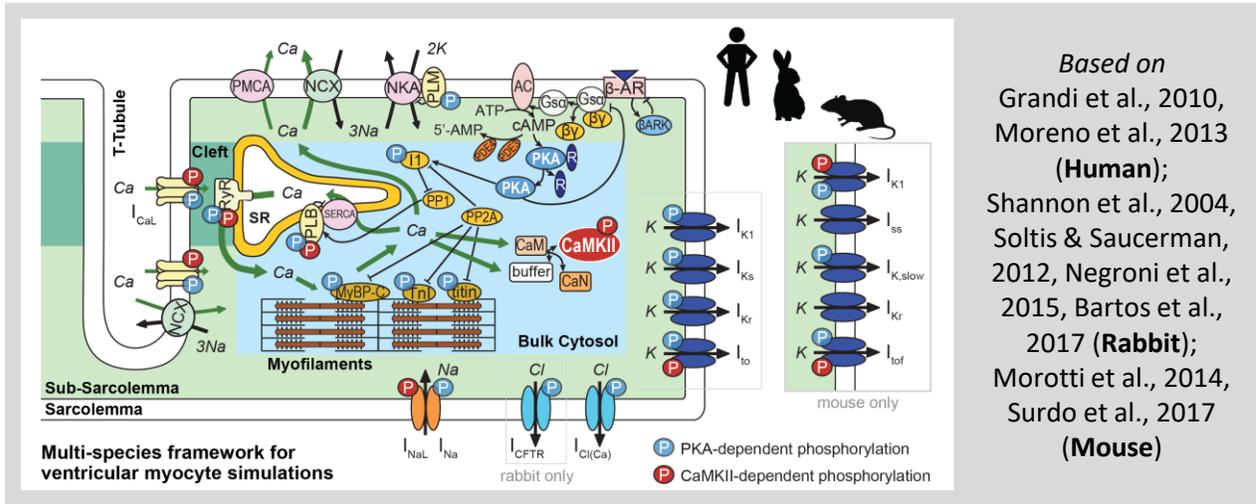
# Methods: performing sensitivity analysis



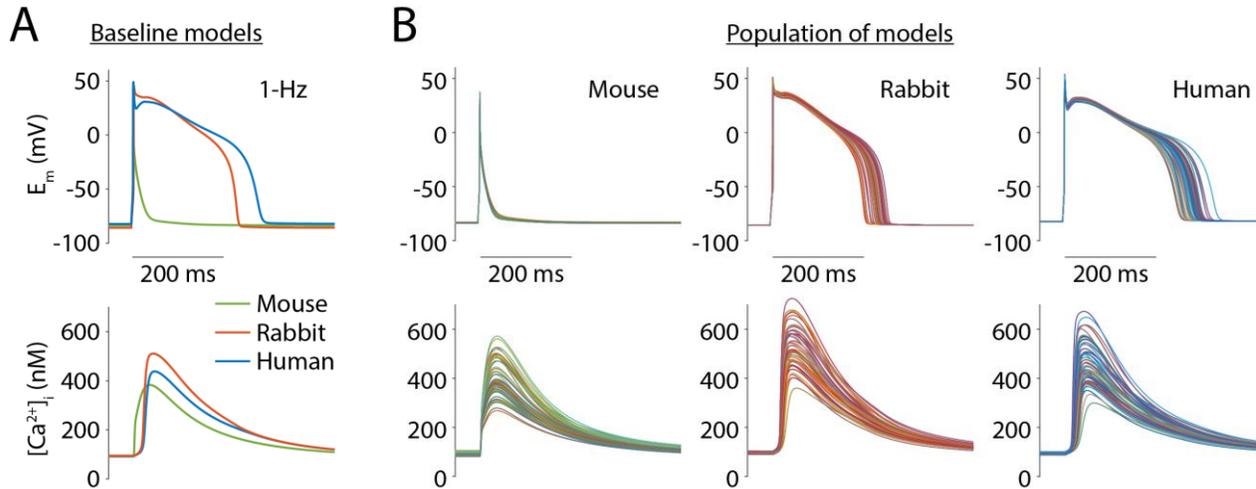
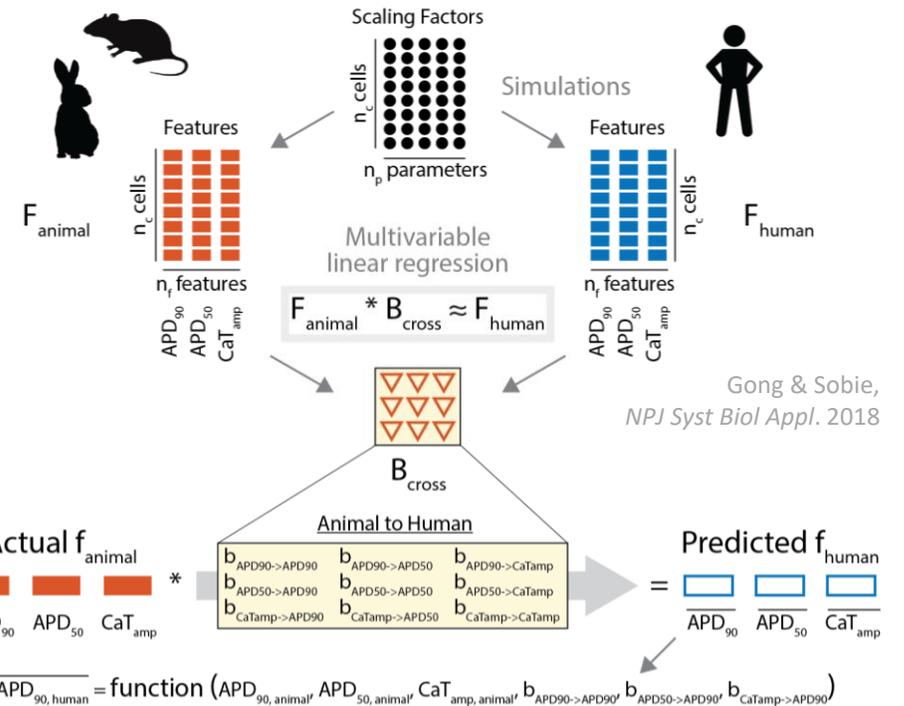
Based on  
 Grandi et al., 2010,  
 Moreno et al., 2013  
 (Human);  
 Shannon et al., 2004,  
 Soltis & Saucerman,  
 2012, Negroni et al.,  
 2015, Bartos et al.,  
 2017 (Rabbit);  
 Morotti et al., 2014,  
 Surdo et al., 2017  
 (Mouse)



# Methods: constructing the translators

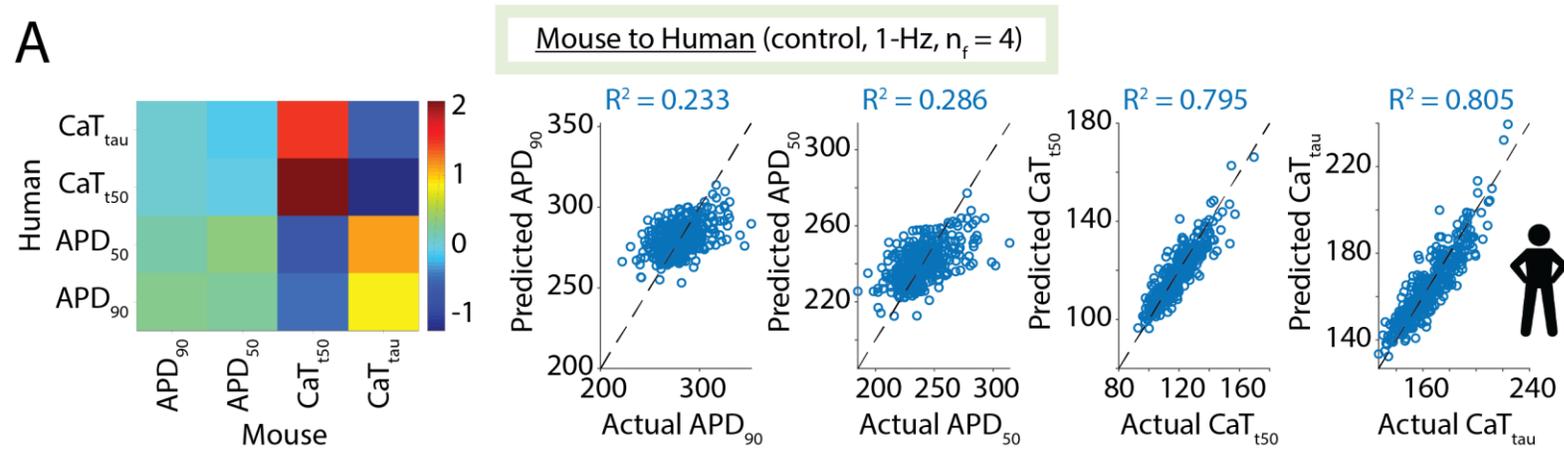


Based on  
 Grandi et al., 2010,  
 Moreno et al., 2013  
 (Human);  
 Shannon et al., 2004,  
 Soltis & Saucerman,  
 2012, Negroni et al.,  
 2015, Bartos et al.,  
 2017 (Rabbit);  
 Morotti et al., 2014,  
 Surdo et al., 2017  
 (Mouse)

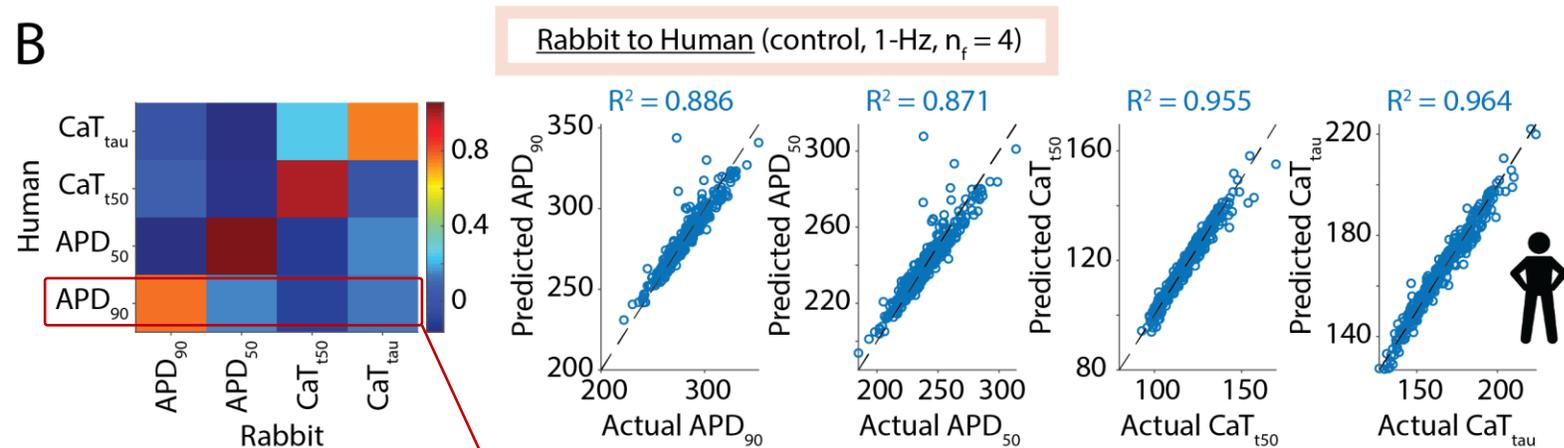


# Development of cross-species translators

A



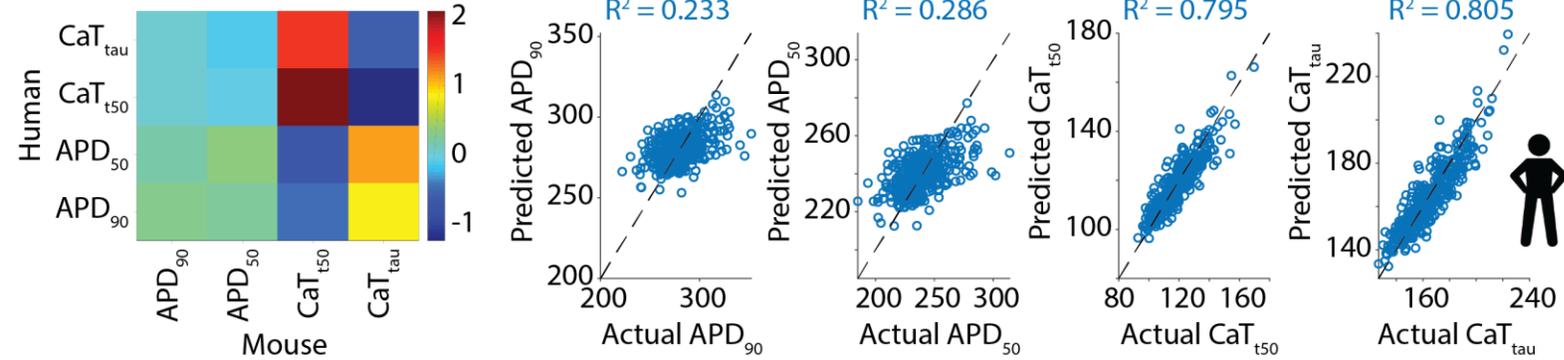
B



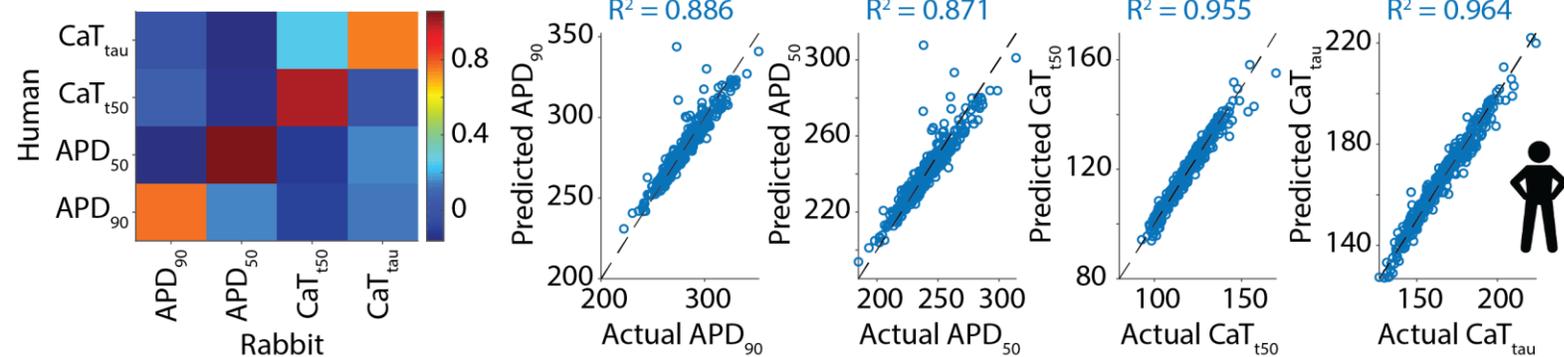
$$APD_{90, human} = function(B_{APD90 \rightarrow APD90}, APD_{90, rabbit}, B_{APD50 \rightarrow APD90}, APD_{50, rabbit}, B_{CaTt50 \rightarrow APD90}, CaT_{t50, rabbit}, B_{CaTtau \rightarrow APD90}, CaT_{tau, rabbit})$$

# Development of cross-species translators

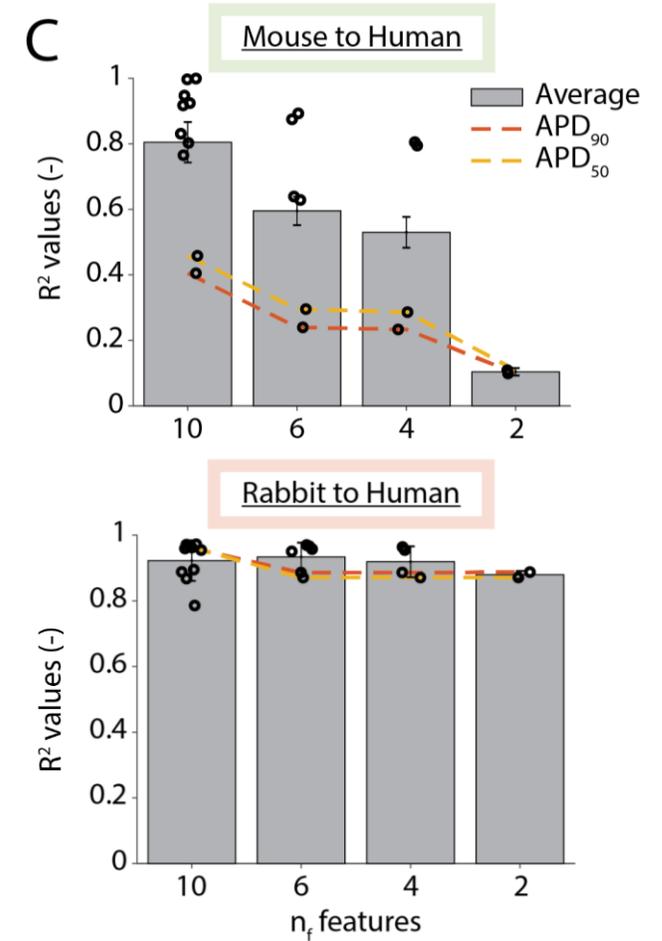
A



B



C



## 10 AP & CaT features

UV, MDP, AP<sub>amp</sub>, APD<sub>90</sub>, APD<sub>50</sub>,  
CaT<sub>min</sub>, CaT<sub>amp</sub>, CaT<sub>ttp</sub>, CaT<sub>t50</sub>, CaT<sub>tau</sub>

## 6 AP & CaT features

APD<sub>90</sub>, APD<sub>50</sub>, CaT<sub>min</sub>, CaT<sub>amp</sub>, CaT<sub>t50</sub>, CaT<sub>tau</sub>

## 4 AP & CaT features

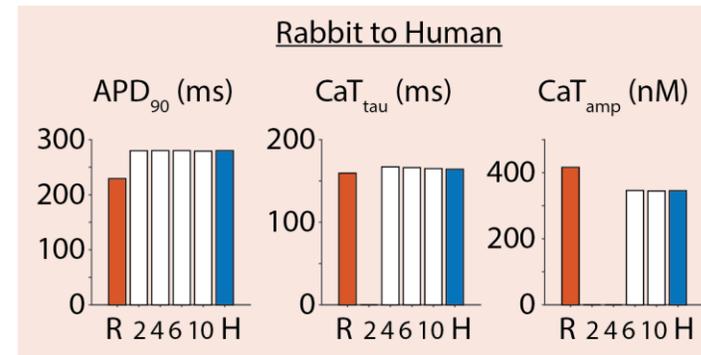
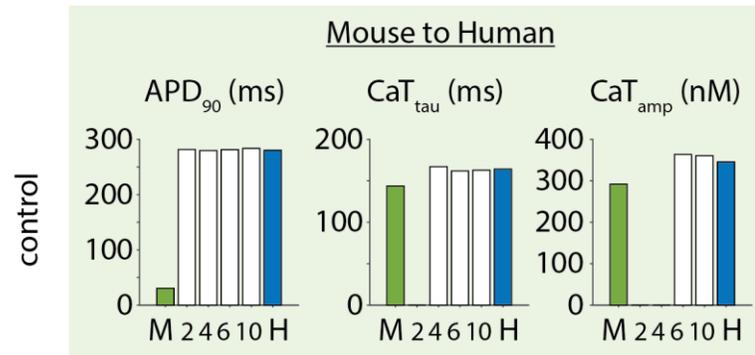
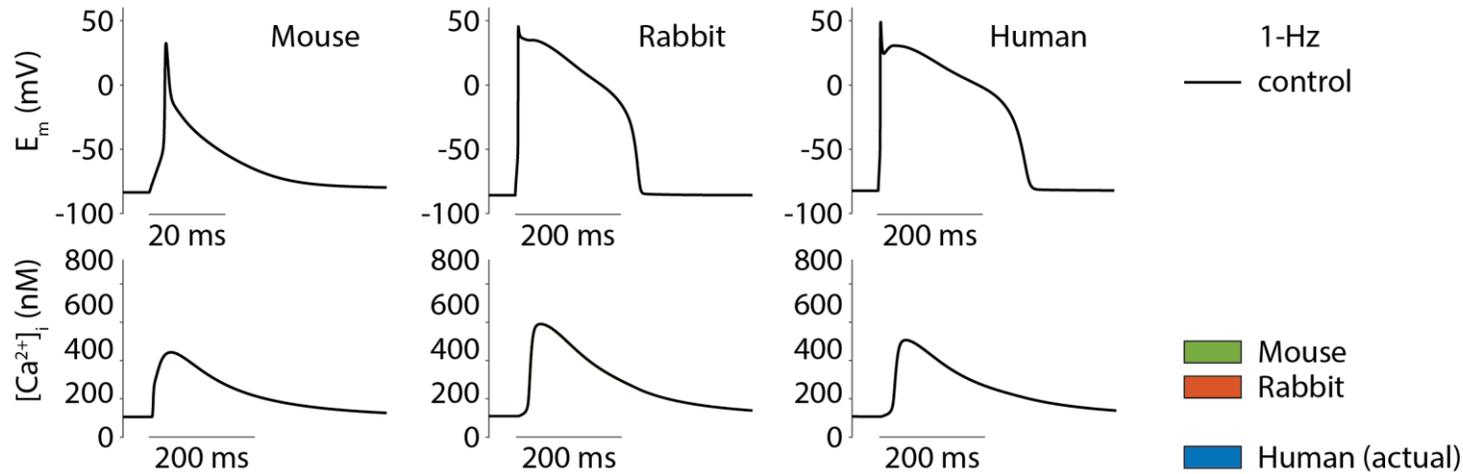
APD<sub>90</sub>, APD<sub>50</sub>, CaT<sub>t50</sub>, CaT<sub>tau</sub>

## 2 AP features

APD<sub>90</sub>, APD<sub>50</sub>

# Validation against simulated data

- Control



**2 AP features**

APD<sub>90</sub>, APD<sub>50</sub>

**4 AP & CaT features**

APD<sub>90</sub>, APD<sub>50</sub>, CaT<sub>t50</sub>, CaT<sub>tau</sub>

**6 AP & CaT features**

APD<sub>90</sub>, APD<sub>50</sub>, CaT<sub>min</sub>, CaT<sub>amp</sub>, CaT<sub>t50</sub>, CaT<sub>tau</sub>

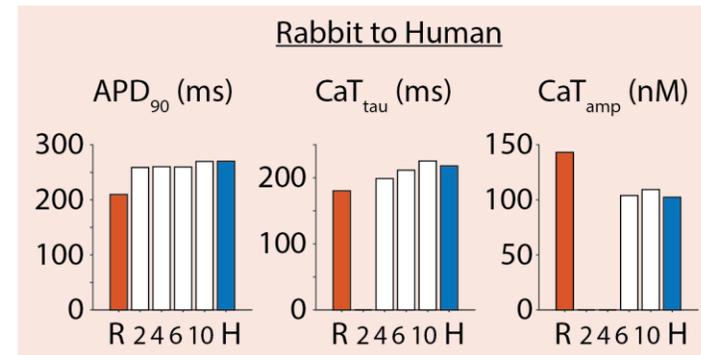
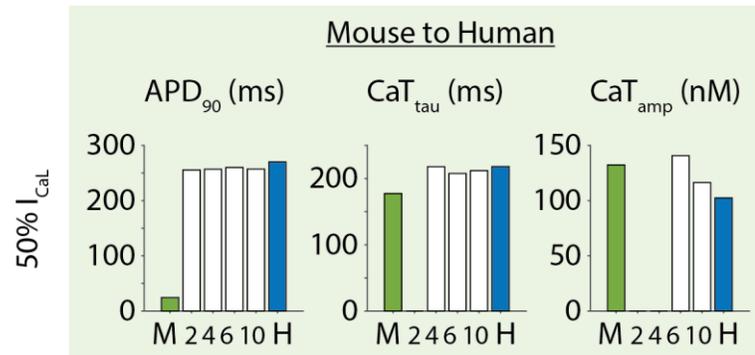
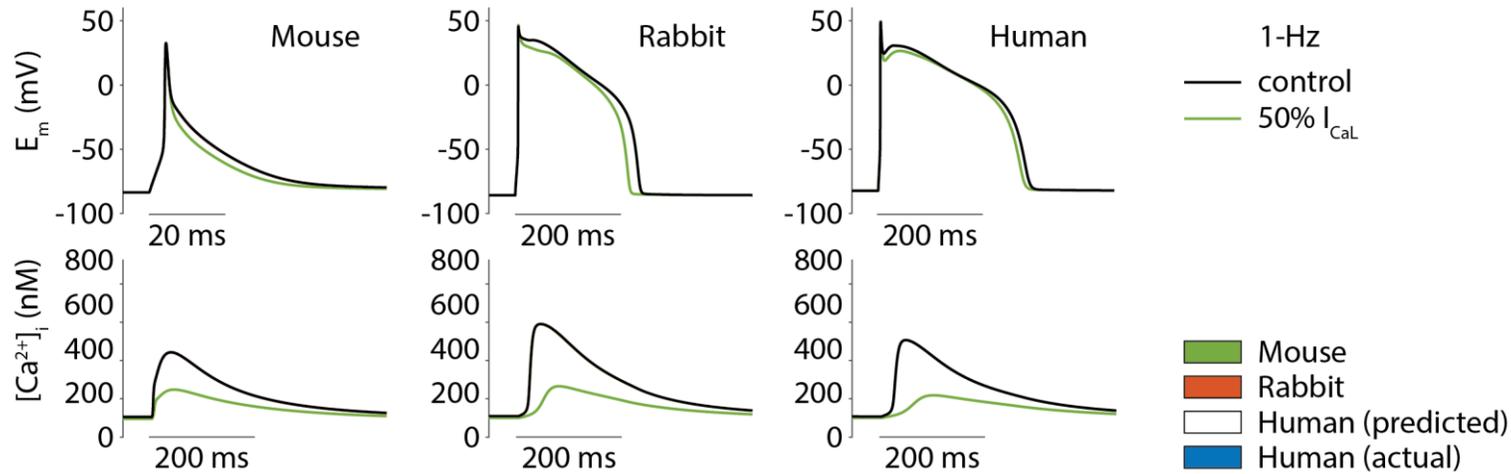
**10 AP & CaT features**

UV, MDP, AP<sub>amp</sub>, APD<sub>90</sub>, APD<sub>50</sub>, CaT<sub>min</sub>, CaT<sub>amp</sub>, CaT<sub>ttp</sub>, CaT<sub>t50</sub>, CaT<sub>tau</sub>

# Validation against simulated data

- *Selective ion channel block*

$I_{CaL}$  block



**2 AP features**

APD<sub>90</sub>, APD<sub>50</sub>

**4 AP & CaT features**

APD<sub>90</sub>, APD<sub>50</sub>, CaT<sub>t50</sub>, CaT<sub>tau</sub>

**6 AP & CaT features**

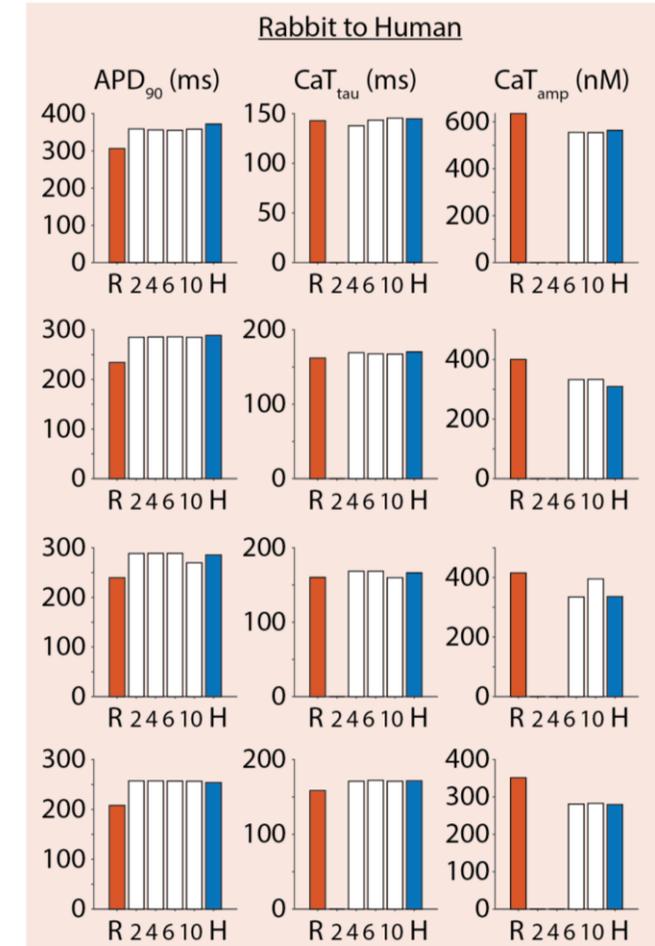
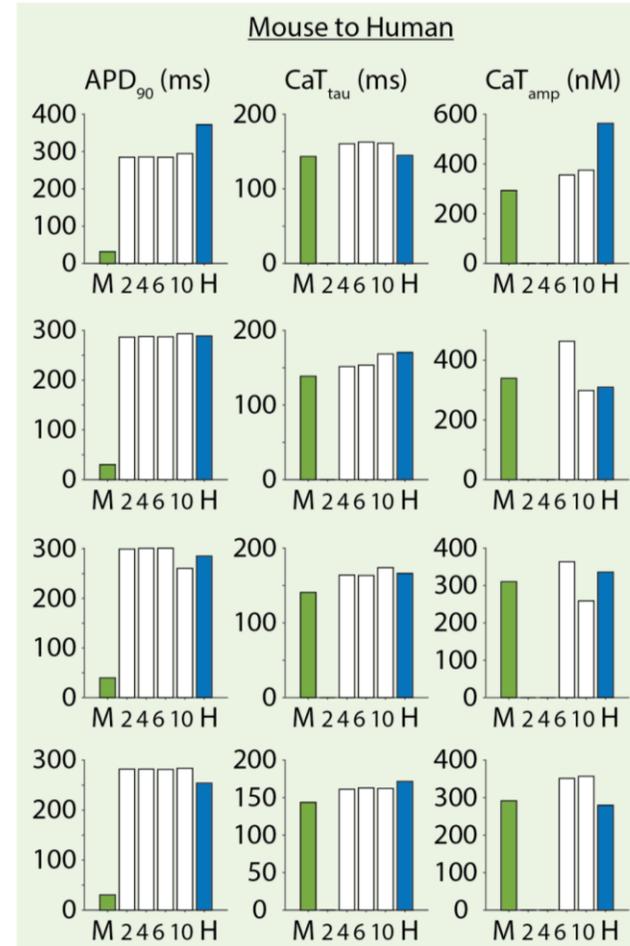
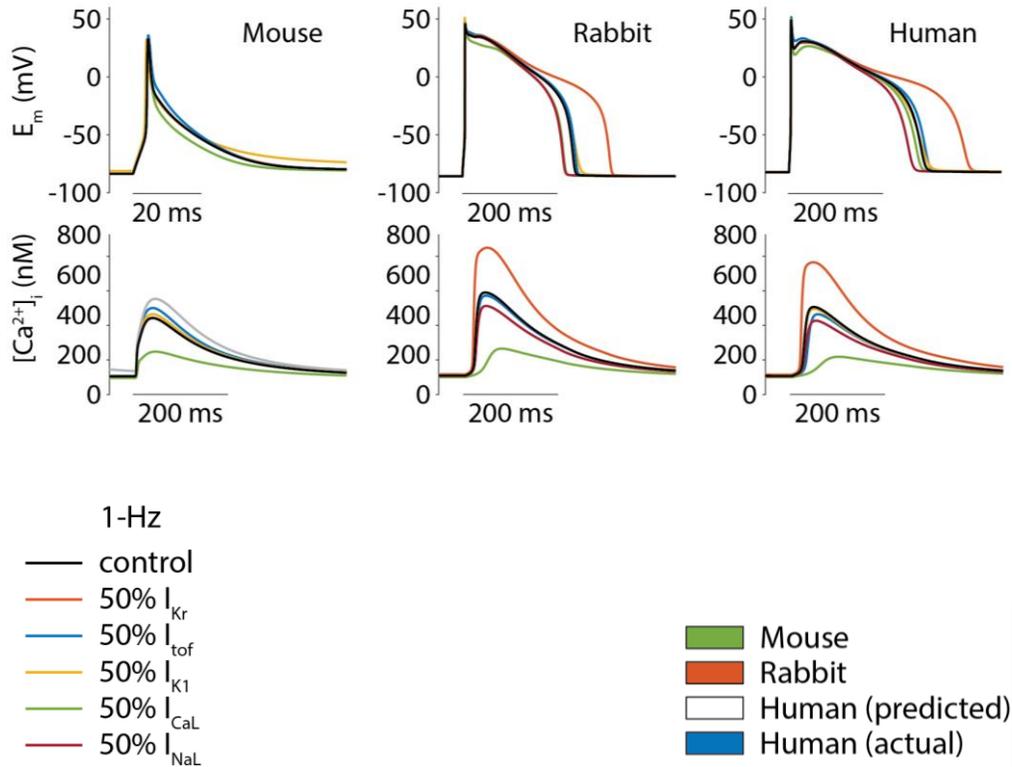
APD<sub>90</sub>, APD<sub>50</sub>, CaT<sub>min</sub>, CaT<sub>amp</sub>, CaT<sub>t50</sub>, CaT<sub>tau</sub>

**10 AP & CaT features**

UV, MDP, AP<sub>amp</sub>, APD<sub>90</sub>, APD<sub>50</sub>, CaT<sub>min</sub>, CaT<sub>amp</sub>, CaT<sub>ttp</sub>, CaT<sub>t50</sub>, CaT<sub>tau</sub>

# Validation against simulated data

- Selective ion channel block*



## 2 AP features

$APD_{90}$ ,  $APD_{50}$

## 4 AP & CaT features

$APD_{90}$ ,  $APD_{50}$ ,  $CaT_{t50}$ ,  $CaT_{\tau}$

## 6 AP & CaT features

$APD_{90}$ ,  $APD_{50}$ ,  $CaT_{min}$ ,  $CaT_{amp}$ ,  $CaT_{t50}$ ,  $CaT_{\tau}$

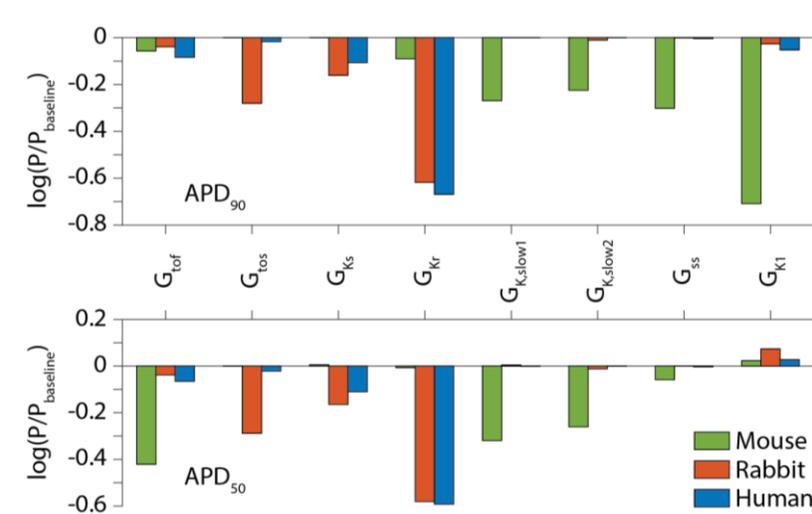
## 10 AP & CaT features

UV, MDP,  $AP_{amp}$ ,  $APD_{90}$ ,  $APD_{50}$ ,  $CaT_{min}$ ,  $CaT_{amp}$ ,  $CaT_{ttp}$ ,  $CaT_{t50}$ ,  $CaT_{\tau}$

# Validation against simulated data

## Limitations of mouse-to-human translation

i) Very different sensitivity to changes in some model parameters  
(e.g.,  $I_{NaL}$ ,  $I_{Kr}$ ,  $I_{K1}$ )



### 2 AP features

$APD_{90}$ ,  $APD_{50}$

### 4 AP & CaT features

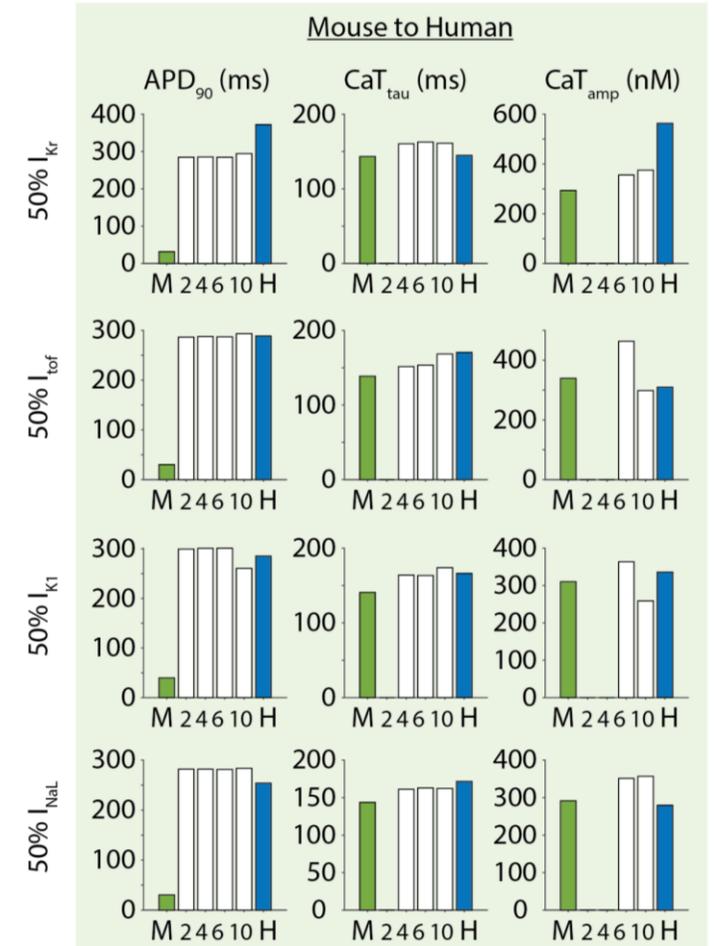
$APD_{90}$ ,  $APD_{50}$ ,  $CaT_{t50}$ ,  $CaT_{tau}$

### 6 AP & CaT features

$APD_{90}$ ,  $APD_{50}$ ,  $CaT_{min}$ ,  $CaT_{amp}$ ,  $CaT_{t50}$ ,  $CaT_{tau}$

### 10 AP & CaT features

UV, MDP,  $AP_{amp}$ ,  $APD_{90}$ ,  $APD_{50}$ ,  $CaT_{min}$ ,  $CaT_{amp}$ ,  $CaT_{ttp}$ ,  $CaT_{t50}$ ,  $CaT_{tau}$

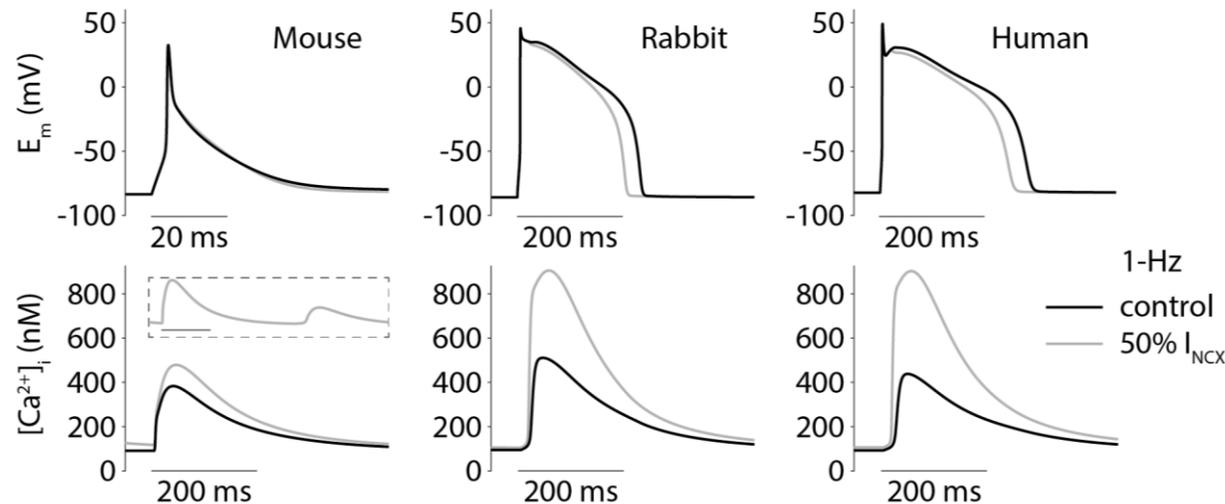


# Validation against simulated data

## Limitations of mouse-to-human translation

i) Very different sensitivity to changes in some model parameters  
(e.g.,  $I_{NaL}$ ,  $I_{Kr}$ ,  $I_{K1}$ )

ii) Different propensity for membrane potential and/or Ca instabilities



### 2 AP features

APD<sub>90</sub>, APD<sub>50</sub>

### 4 AP & CaT features

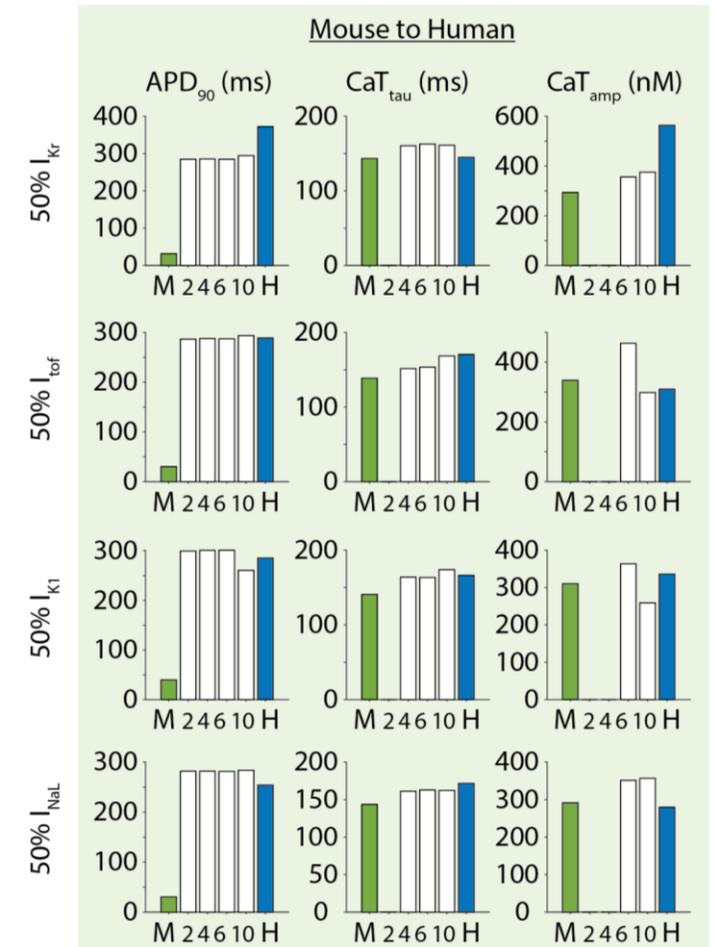
APD<sub>90</sub>, APD<sub>50</sub>, CaT<sub>t50</sub>, CaT<sub>tau</sub>

### 6 AP & CaT features

APD<sub>90</sub>, APD<sub>50</sub>, CaT<sub>min</sub>, CaT<sub>amp</sub>, CaT<sub>t50</sub>, CaT<sub>tau</sub>

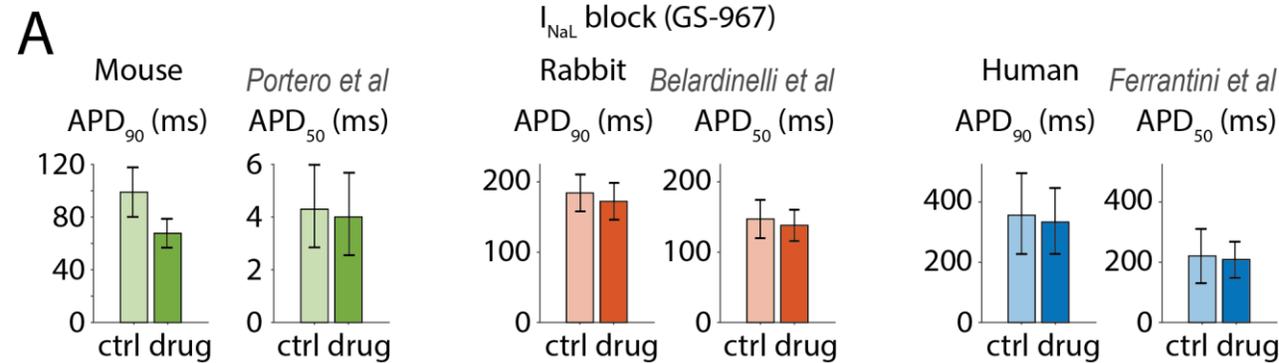
### 10 AP & CaT features

UV, MDP, AP<sub>amp</sub>, APD<sub>90</sub>, APD<sub>50</sub>, CaT<sub>min</sub>, CaT<sub>amp</sub>, CaT<sub>ttp</sub>, CaT<sub>t50</sub>, CaT<sub>tau</sub>



# Experimental validation

## $I_{NaL}$ block



To account for variability among experimental datasets, when applying our translators to experimental data, we use the **relative changes** in the measured AP & CaT properties induced by a perturbation (rather than the absolute values)

$$\text{Actual } f_{\text{animal, drug}} * B_{\text{cross}} = \text{Predicted } f_{\text{animal, drug}}$$

$$\text{APD}_{90, \text{human, drug}} = \text{function} (\text{APD}_{90, \text{animal, drug}}, \text{APD}_{50, \text{animal, drug}}, \text{CaT}_{\text{amp, animal, drug}}, b_{\text{APD90} \rightarrow \text{APD90}}, b_{\text{APD50} \rightarrow \text{APD90}}, b_{\text{CaTamp} \rightarrow \text{APD90}})$$

with

$$\text{APD}_{90, \text{animal, drug}} = \text{APD}_{90, \text{baseline animal model, ctrl}} * \text{APD}_{90, \text{exp, drug}} / \text{APD}_{90, \text{exp, ctrl}}$$

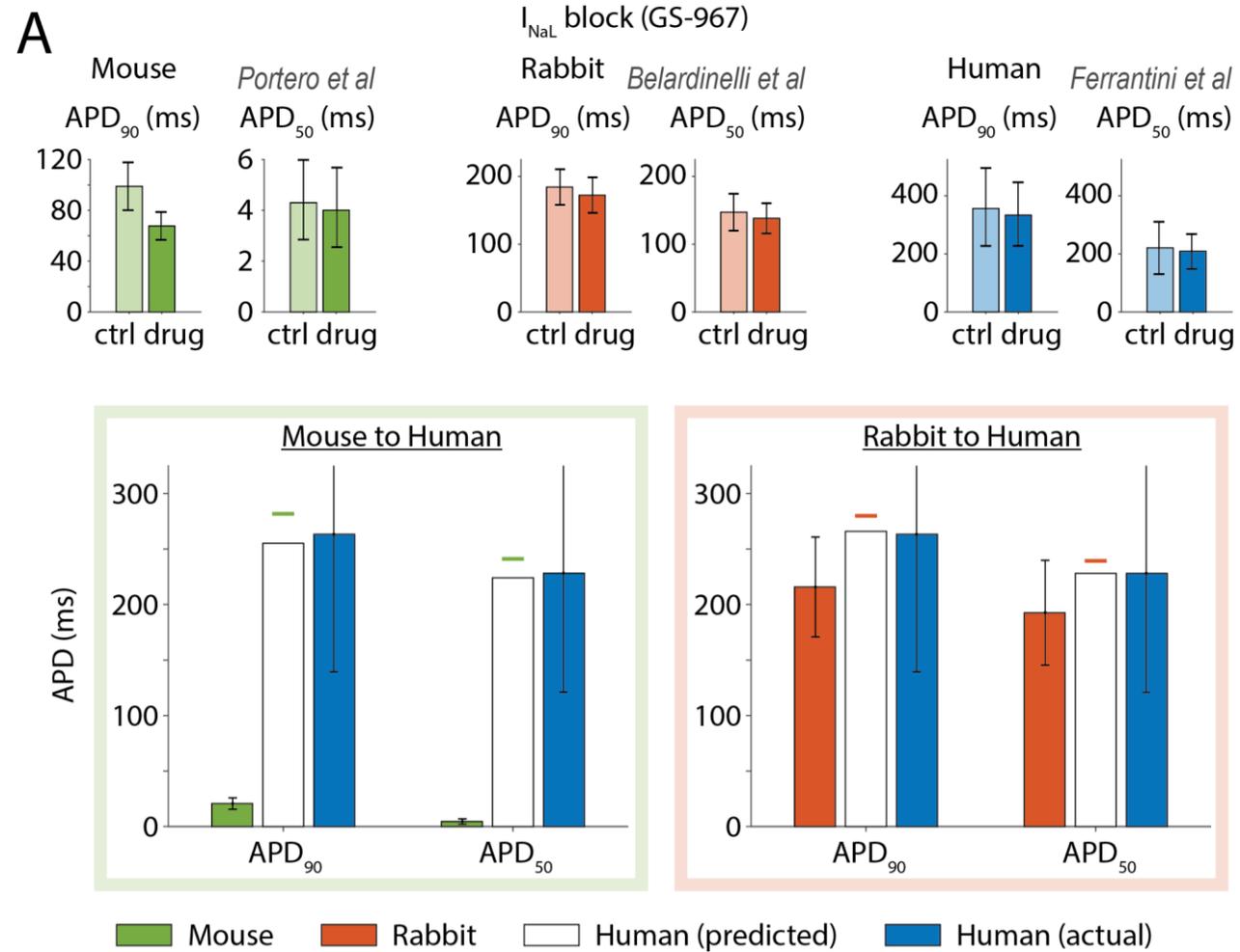
$$\text{APD}_{50, \text{rabbit, drug}} = \text{APD}_{50, \text{baseline animal model, ctrl}} * \text{APD}_{50, \text{exp, drug}} / \text{APD}_{50, \text{exp, ctrl}}$$

$$\text{CaT}_{\text{amp, rabbit, drug}} = \text{CaT}_{\text{amp, baseline animal model, ctrl}} * \text{CaT}_{\text{amp, exp, drug}} / \text{CaT}_{\text{amp, exp, ctrl}}$$

Simulations with baseline model  $\rightarrow$  Experiments  $\rightarrow$

# Experimental validation

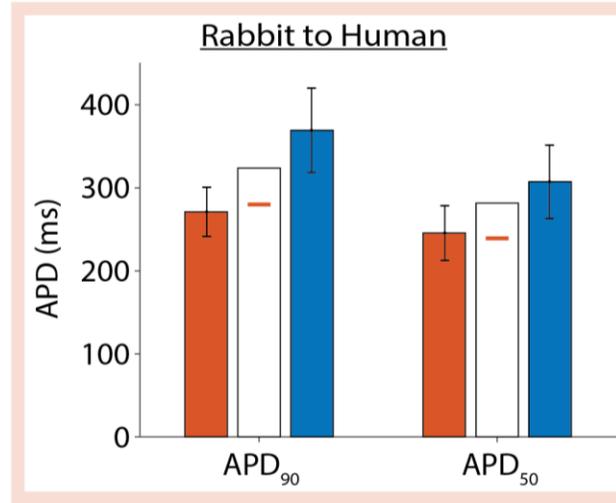
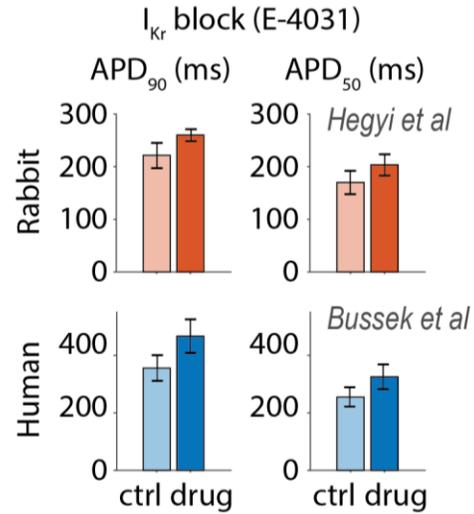
## $I_{NaL}$ block



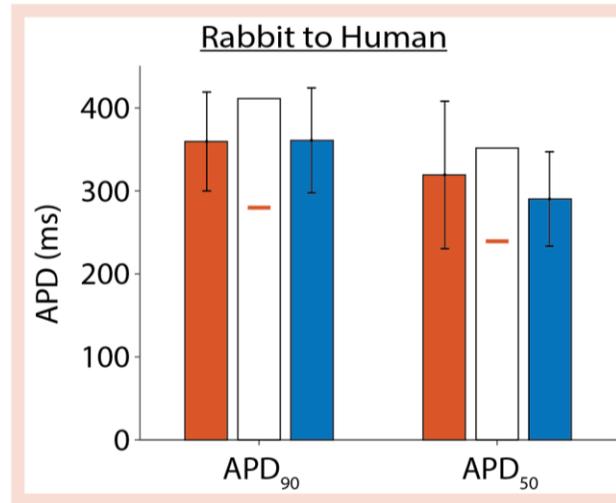
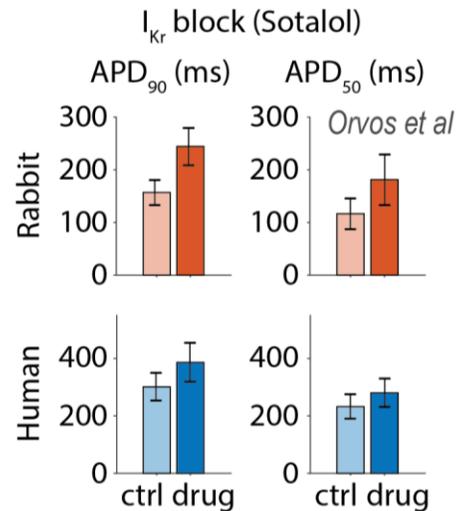
# Experimental validation

$I_{Kr}$  block

B



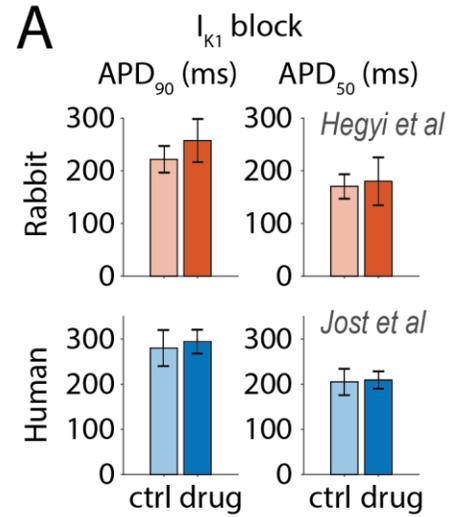
C



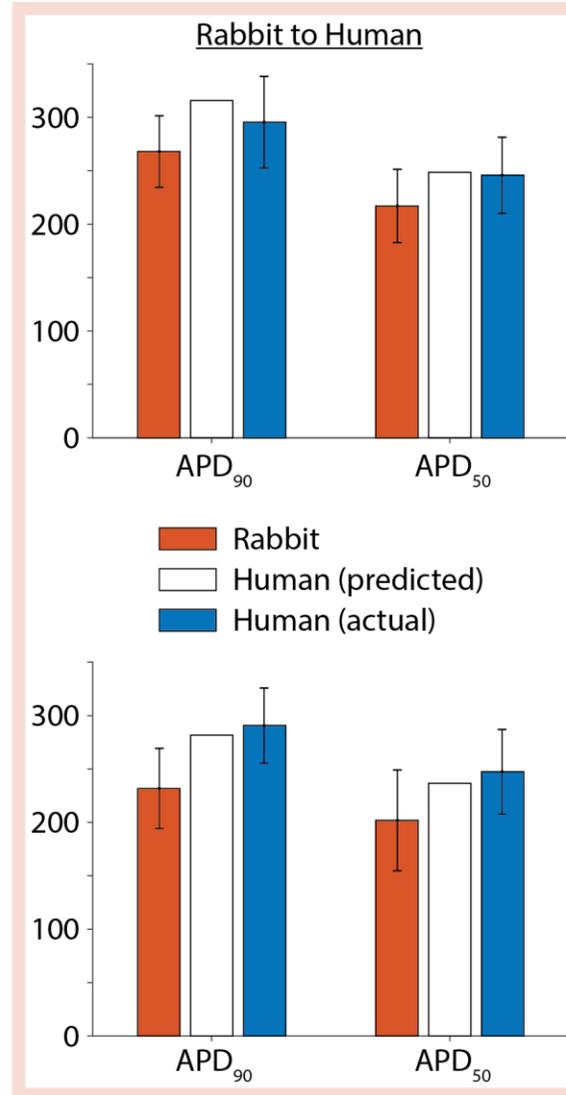
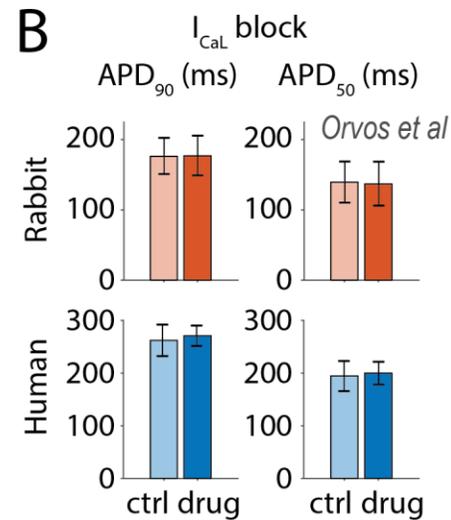
■ Mouse    ■ Rabbit    □ Human (predicted)    ■ Human (actual)

# Experimental validation

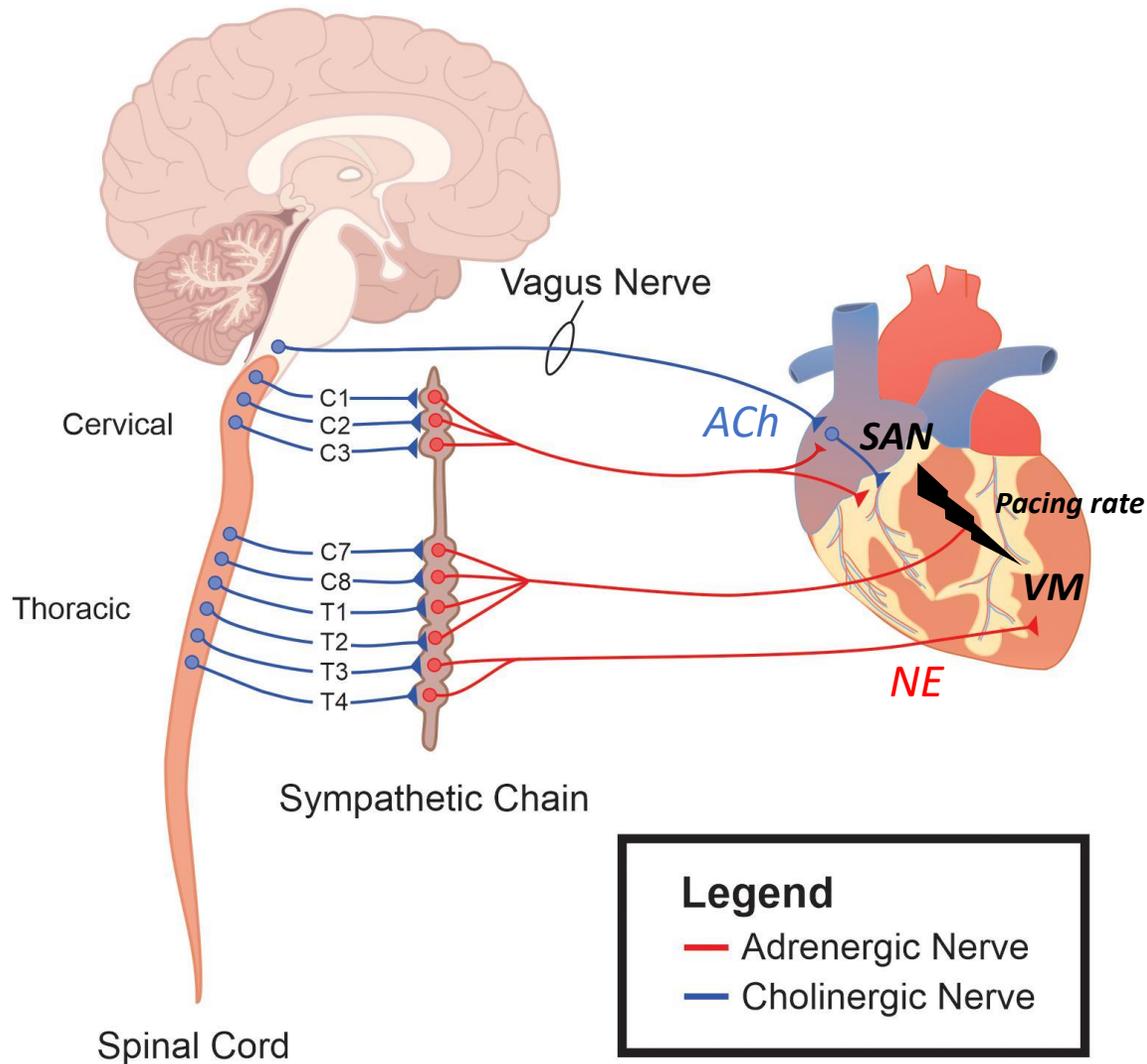
$I_{K1}$  block



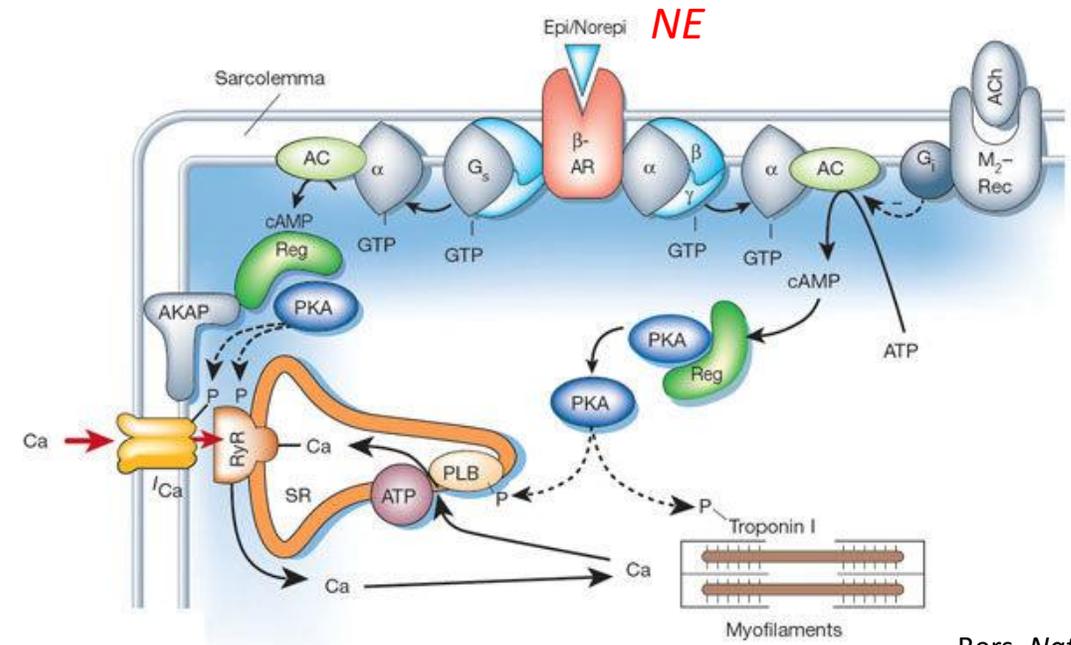
$I_{CaL}$  block



# Prediction of response to sympathetic stimulation

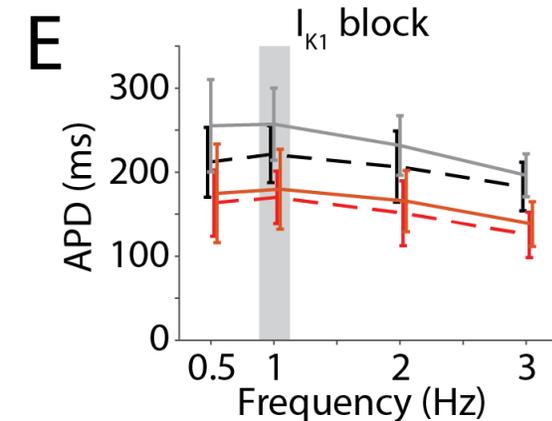
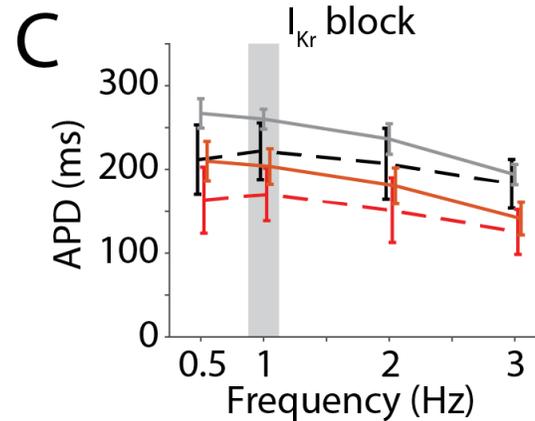
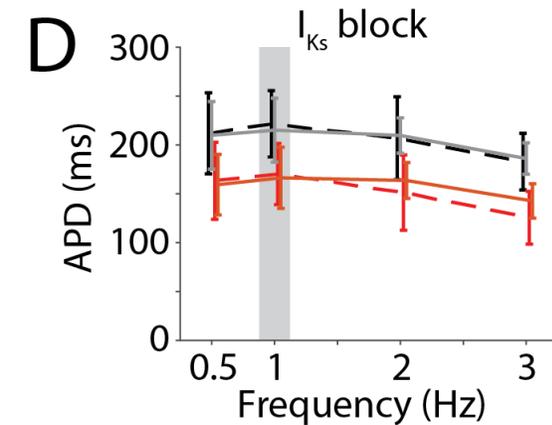
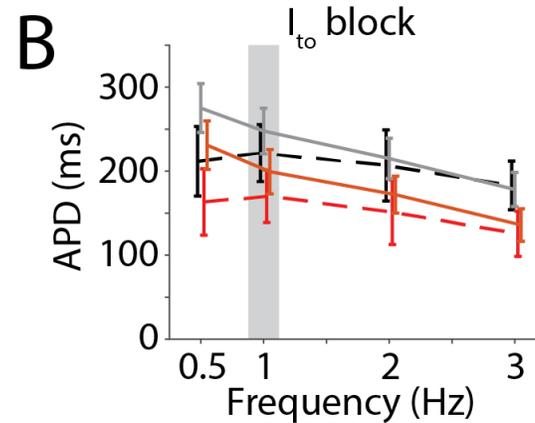
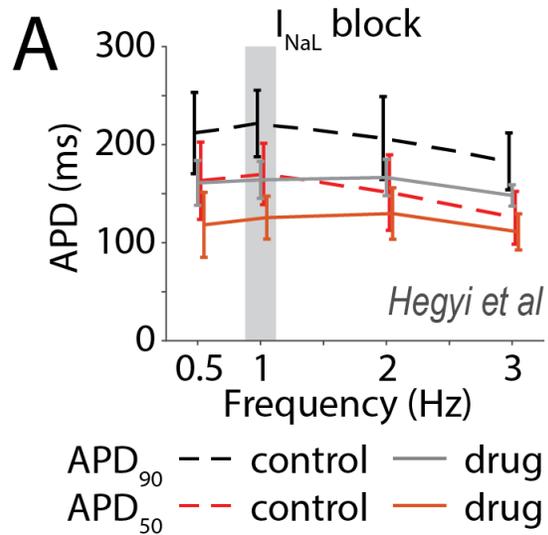


- Ventricular activity is influenced by:
  - Increased beating rate (via SAN)
  - Altered activity of the targets of the  $\beta$ -adrenergic ( $\beta$ -AR) signaling cascade



# Cross-frequency prediction of drug-induced effect

- Frequency-dependence of ion channel block in rabbit ventricular myocytes

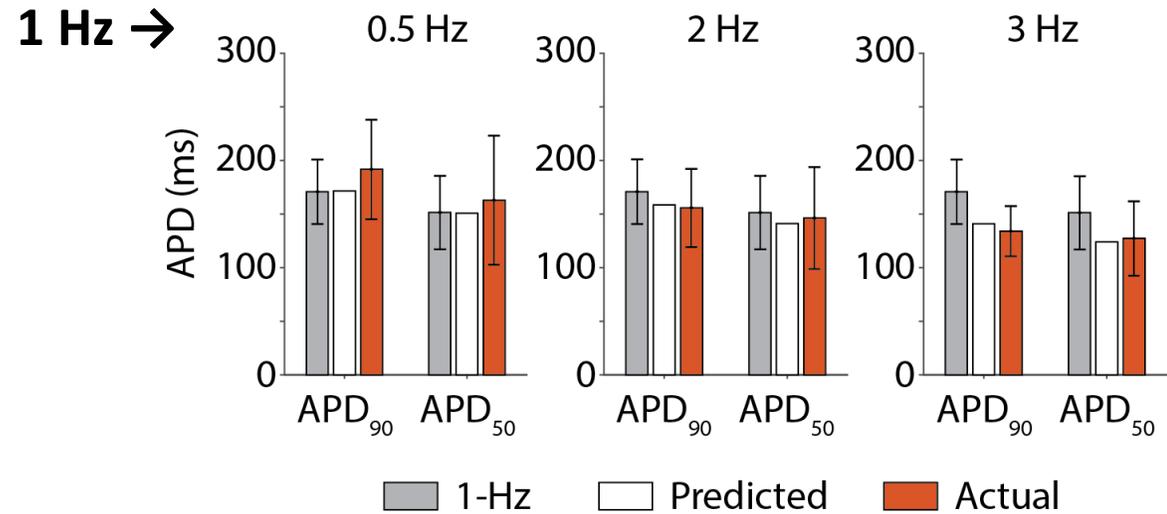
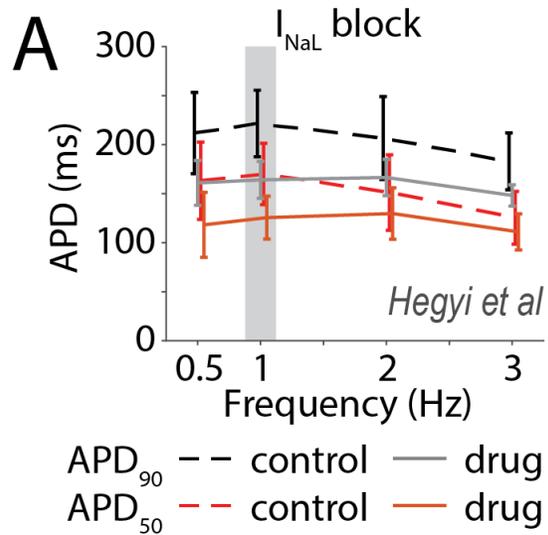


Courtesy of Dr. Bence Hegyi  
Bers Lab, UC Davis



# Cross-frequency prediction of drug-induced effect

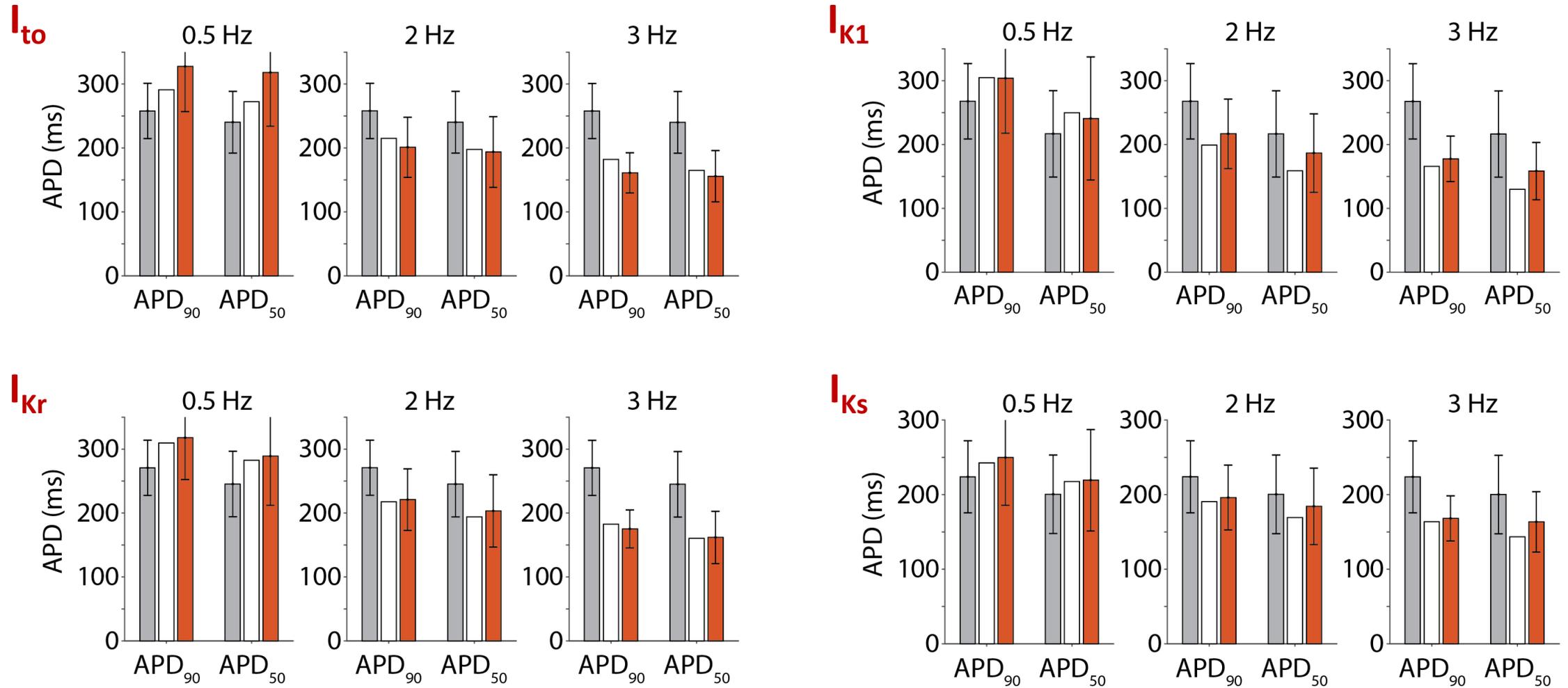
- Predicting the effect of block of  $I_{NaL}$  on APD from 1 to 0.5, 2 & 3 Hz data



Courtesy of Dr. Bence Hegyi  
Bers Lab, UC Davis



# Cross-frequency prediction of drug-induced effect

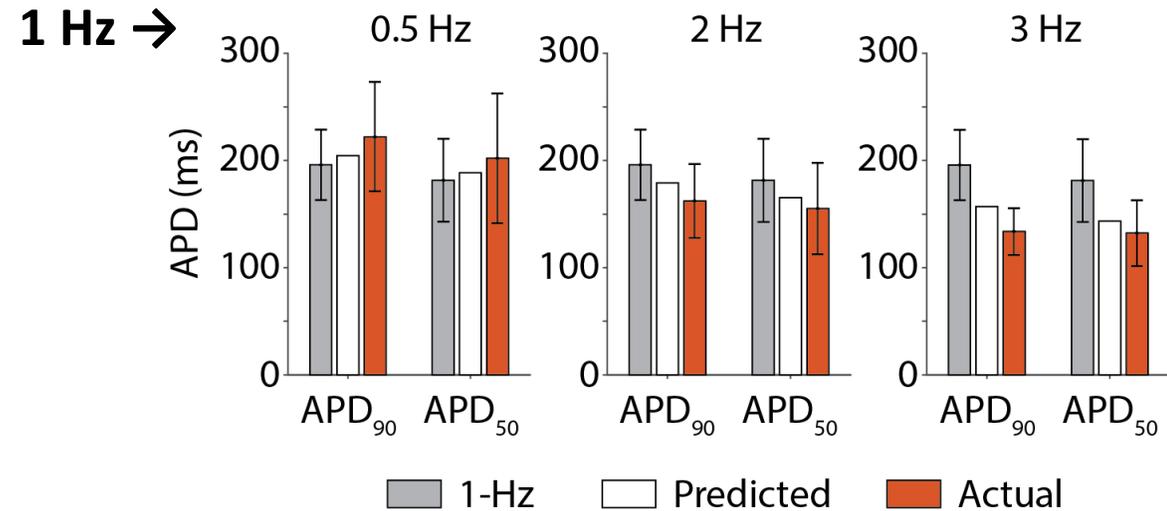
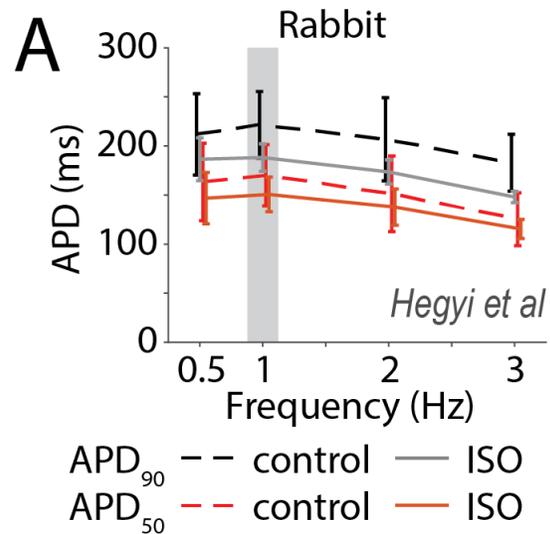


1-Hz Predicted Actual



# Cross-frequency prediction of ISO-induced effect

- Predicting the effect of Isoproterenol (**ISO**) on APD from 1 to 0.5, 2 & 3 Hz data

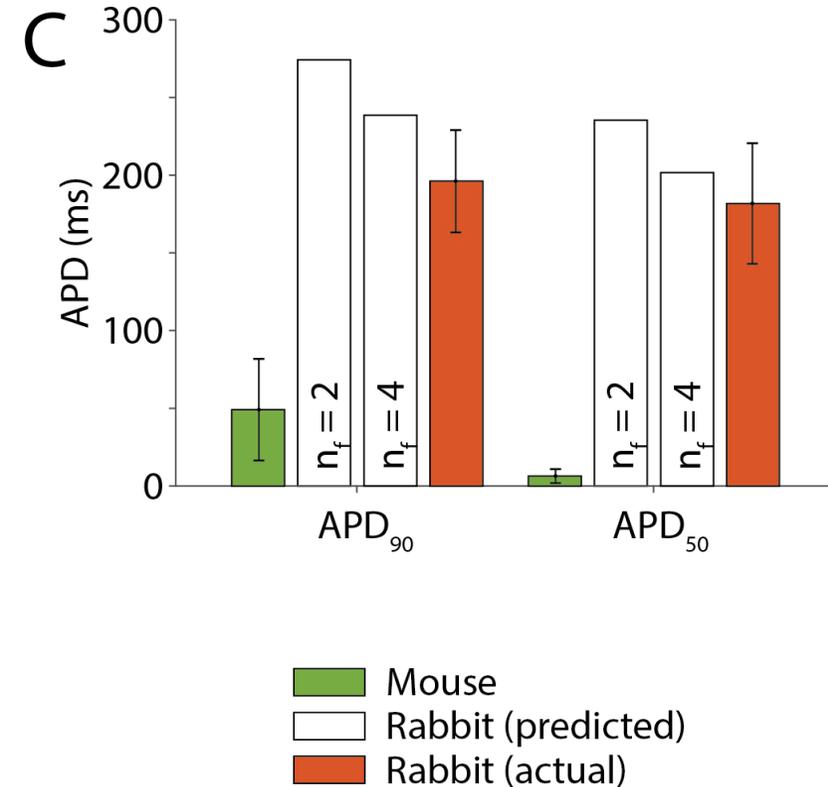
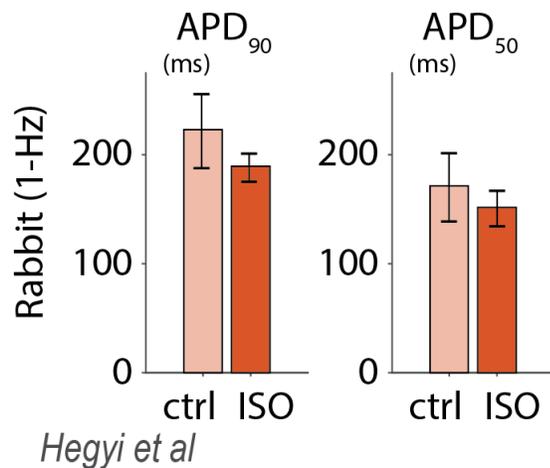
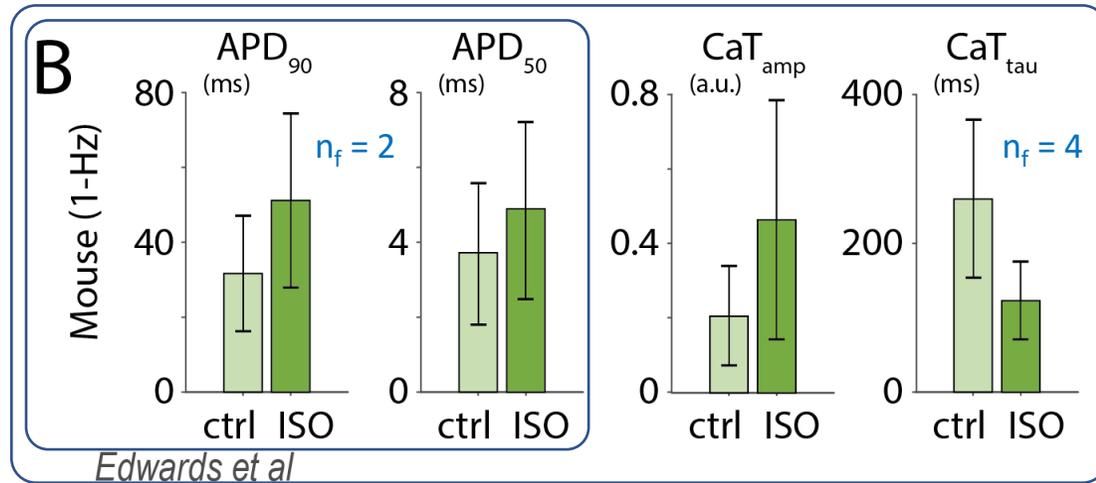


Courtesy of Dr. Bence Hegyi  
Bers Lab, UC Davis

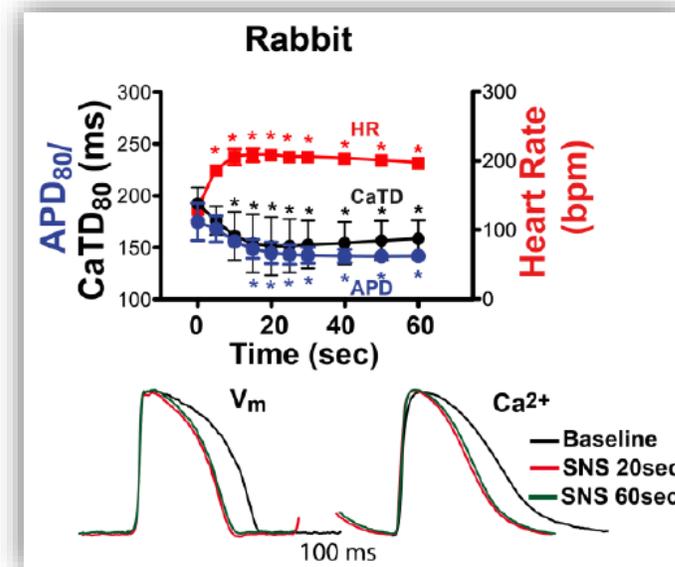
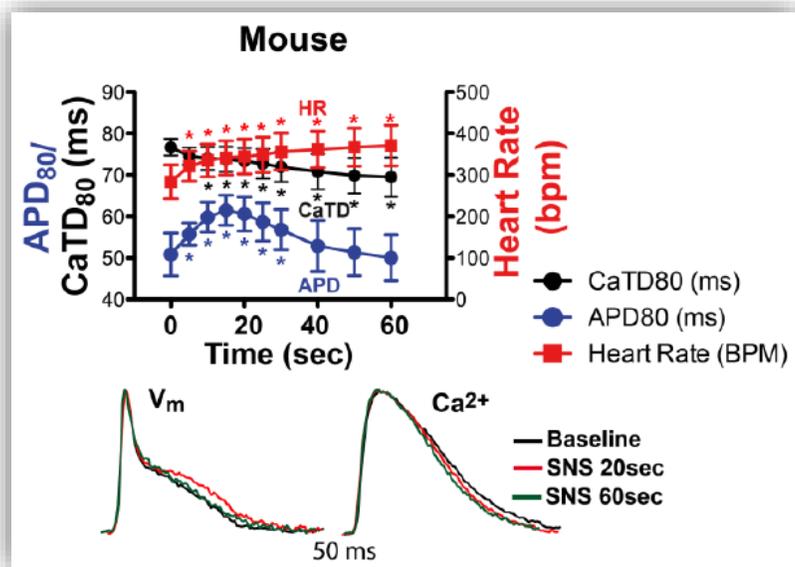


# Cross-species prediction of ISO effect at fixed pacing rate

- Predicting the effect of **ISO** administration on rabbit APD from mouse data (1 Hz)



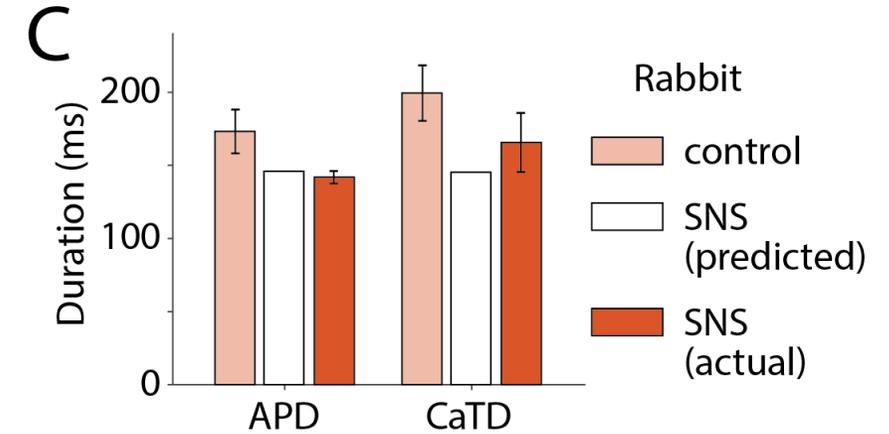
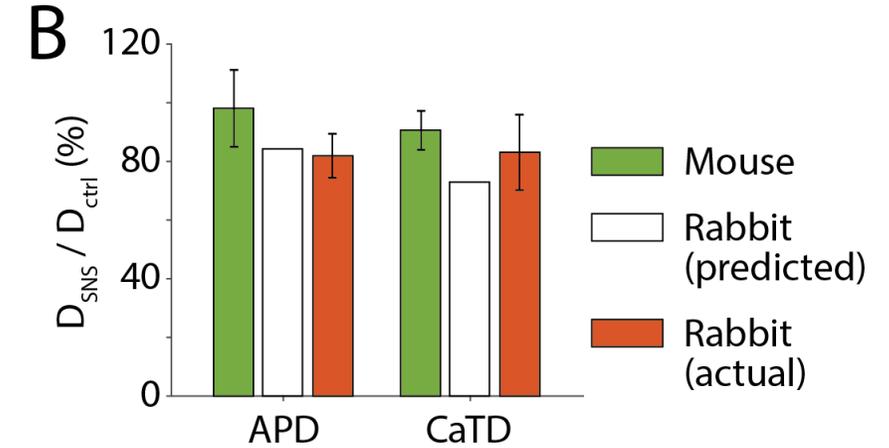
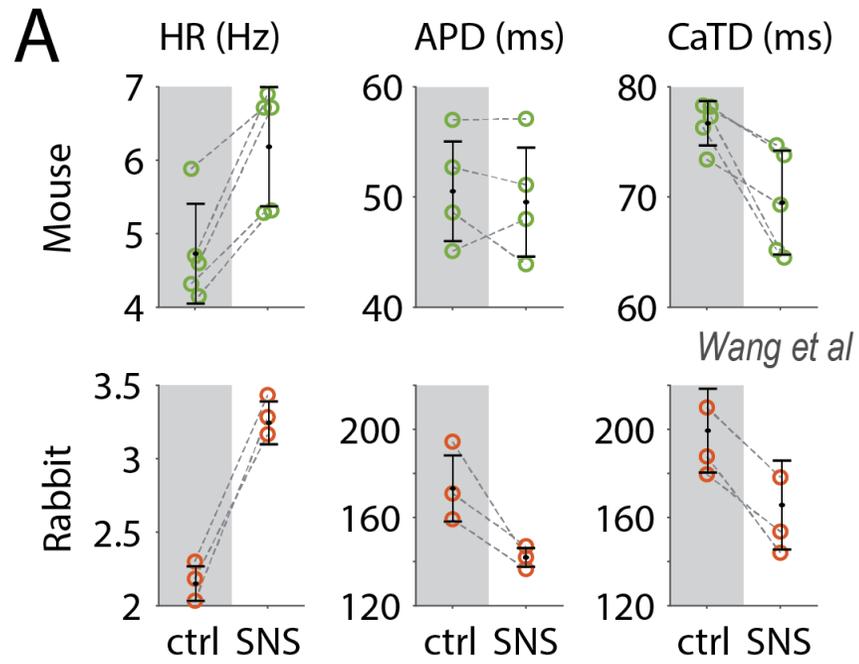
# Sympathetic stimulation in quasi-physiological conditions



# Sympathetic stimulation in quasi-physiological conditions



SNS-specific cross-species translators  
of the relative effect induced by  
sympathetic activation  
(accounting for change in beating rate)



# Experimental validation summary

- Cross-species prediction of drug-induced effect  
(mouse & rabbit to human) ✓
- Cross-frequency prediction of drug-induced effect (rabbit) ✓
- Cross-frequency prediction of ISO-induced effect (rabbit) ✓
- Cross-species prediction of ISO-induced effect at fixed pacing rate  
(mouse to rabbit) ✓
- Cross-species prediction of sympathetic stimulation effect with  
concomitant change in heart rate (mouse to rabbit) ✓

# Conclusions

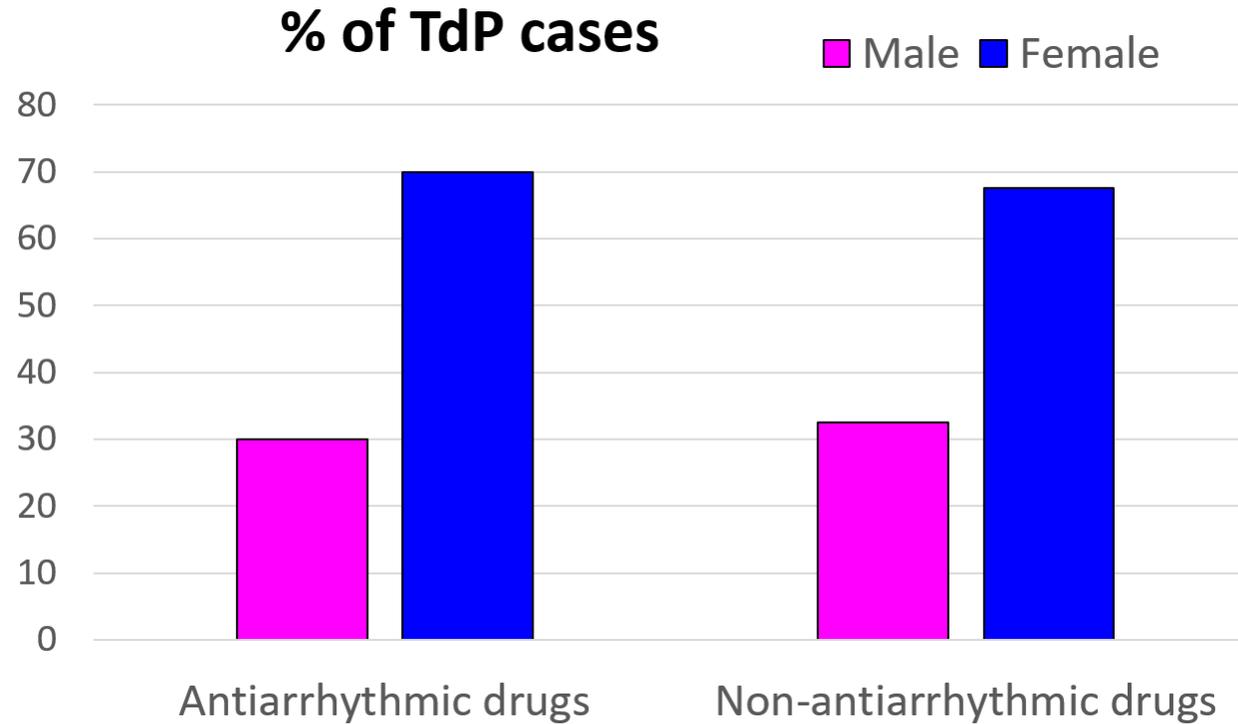
- We constructed a suite of translators for quantitatively mapping electrophysiologic responses across species and experimental conditions
- We trained these statistical operators using a broad dataset obtained simulating populations of our models of mouse, rabbit, and human ventricular myocytes
- We tested our translators against experimental data describing the response to various stimuli (ion channel block, change in beating rate,  $\beta$ -adrenergic challenge)
- **Our work demonstrates that this approach is well suited for predicting the effects of perturbations across different species, thereby suggesting its integration into mechanistic studies and drug development pipelines**

# Future directions

- Further refinement/validation
- Inclusion of more species
- Cross-regional translation (atria $\leftrightarrow$ ventricles)
- **Cross-sex translation**

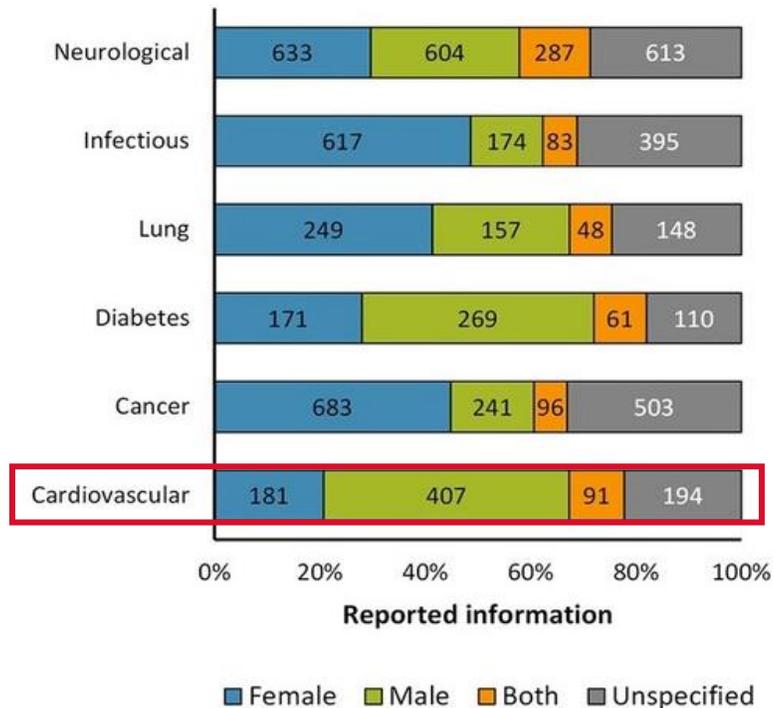
# Cross-sex translation

- Female sex is an independent risk factor for Torsade de Pointes (TdP)



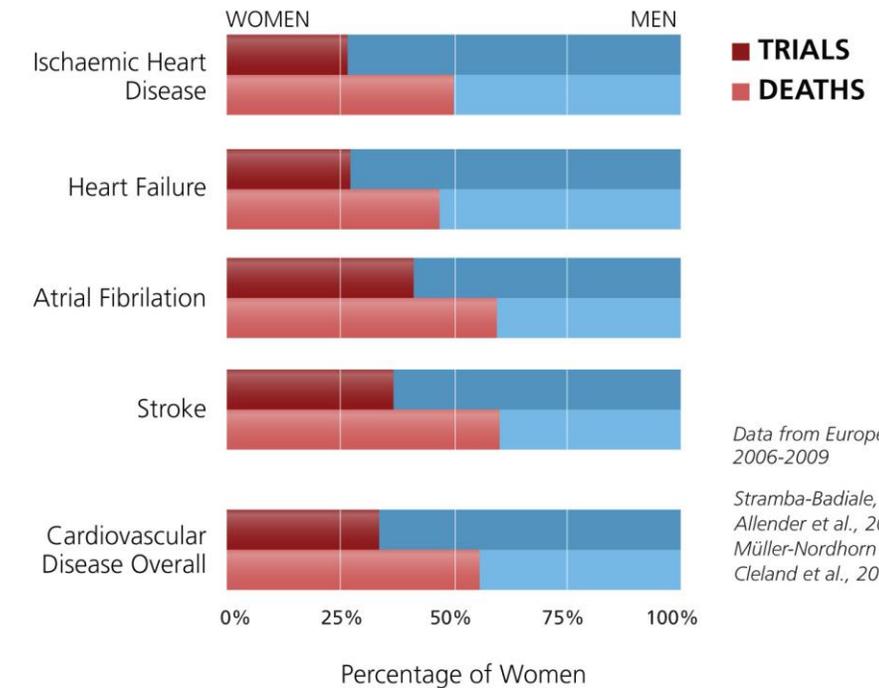
# Cross-sex translation

- Female sex is an independent risk factor for Torsade de Pointes (TdP)
- Female sex is underrepresented in both experimental & clinical studies



## Percentage of Women in CVD Clinical Trials vs. Deaths

Women are underrepresented in CVD clinical trials



Data from European Union, 2006-2009

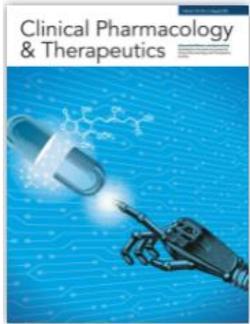
Stramba-Badiale, 2010  
Allender et al., 2008  
Müller-Nordhorn et al., 2008  
Cleland et al., 2003

# Cross-sex translation

ARTICLE

## Sex-Specific Classification of Drug-Induced Torsade de Pointes Susceptibility Using Cardiac Simulations and Machine Learning

Alex Fogli Iseppe<sup>1</sup>, Haibo Ni<sup>1</sup>, Sicheng Zhu<sup>1</sup>, Xianwei Zhang<sup>1</sup>, Raffaele Coppini<sup>2</sup>, Pei-Chi Yang<sup>3</sup>, Uma Srivatsa<sup>4</sup>, Colleen E. Clancy<sup>1,3</sup>, Andrew G. Edwards<sup>1</sup>, Stefano Morotti<sup>1</sup> and Eleonora Grandi<sup>1,\*</sup>



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drug	IC50 (nM)	hERG (nM)	INaI (nM)	ICaL (nM)	INa (nM)	hK1 (nM)	hK2 (nM)
quinidine	992	9417	51923	12329	34874	3958991	4895.9
bepidil	50	1813.9	2888.1	2929.3	8394	0	26282.3
bepidil	0.9	1.4	0.6	1.2	3.5	0	0.7
dofetilide	4.9	251160	2063	288.5	18.8	194.3	0
dofetilide	0	0.3	1.2	0.9	0.8	0.8	0
terfenadine	11660	0	706152	1.1487	4314345	505200	4221856
terfenadine	0	0.8	0	0.9	0.5	0.7	1.2
terfenadine	9292	45596	81919	4335.8	1761871	9269.9	0
chlorpromazine	0	0.8	0.9	0.8	2	0.4	0.7
chlorpromazine	0	0.8	0.9	0.8	2	0.4	0.7
cisapride	10.1	0	925807	0	219112	29498	8119286
cisapride	0	0.7	0	0.4	0.2	0.5	0.3
terfenadine	23	28056	708.4	4882.2	239988	0	399754
terfenadine	0.6	0.6	0.7	1	0.3	0	0.5
ondansetron	1320	191808	225114	57666	1923378	0	509807
ondansetron	0.9	1	0.8	1	1	0	0.7
olmesartan	13150	21868.5	112.1	118859	2.82E+0	9	0
olmesartan	0.9	0.7	0.7	0.7	0.2	0	0

**Experimental Data**  
Ion channel block by drugs

**Clinical Data** on drug toxicity

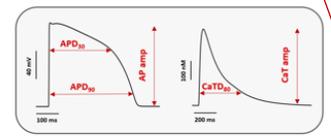
We combined **mechanistic modeling & machine learning** to develop sex-specific TdP classifiers (based on AP & CaT features)

- TdP classifiers require different features in females vs. males
- if applied to female data, male-based classifiers perform poorly and lead to a systematic underestimation of arrhythmic risk

**Simulations** using mechanistic sex-specific models of human cardiac electrophysiology

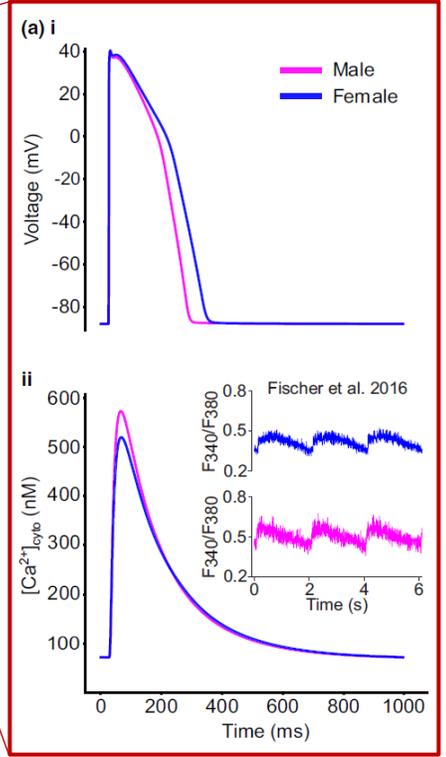
- Various drug concentrations
- Various heart rates
- Various conditions

**Simulated Features**  
APD, Ca<sup>2+</sup> transient properties

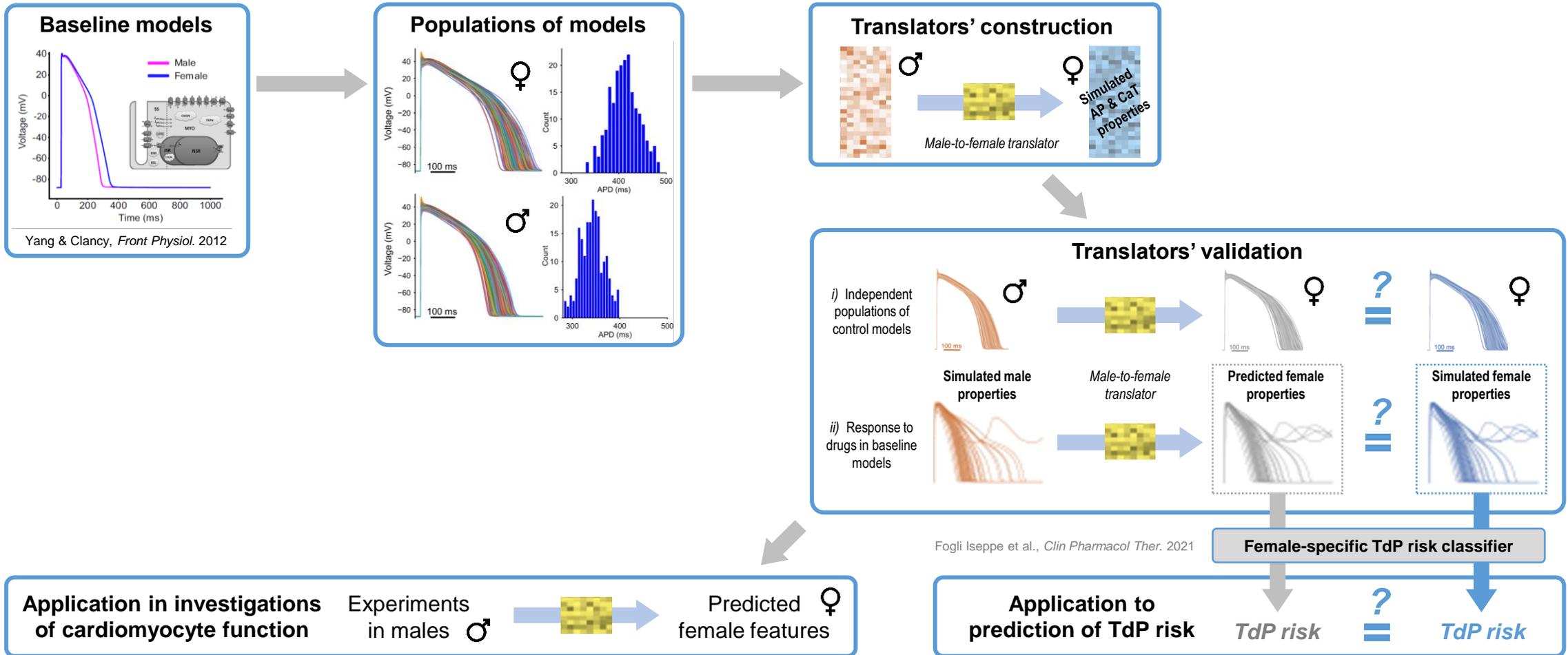


**Statistical Models**  
Using machine learning or logistic regression analysis

**TdP risk?**



# Cross-sex translation



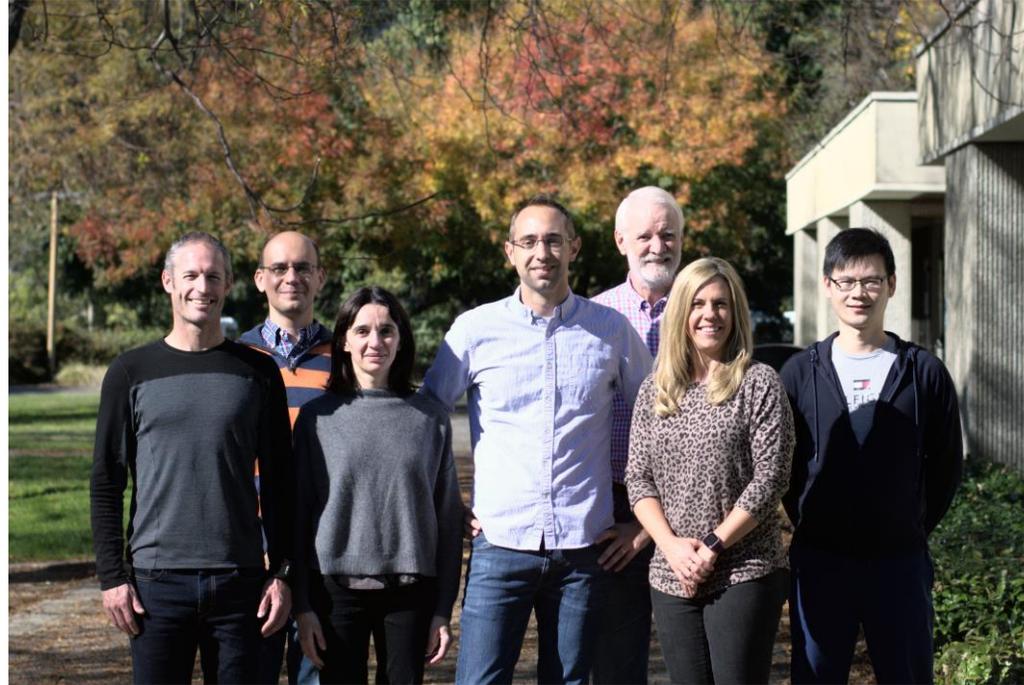
# Acknowledgements

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Lin-Lin Liu

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**HEALTH**



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Lianguo Wang

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Bence Hegyi  
Kim Hellgren



*Marco Pritoni*



**Thank you!**

